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Viral Mimicry of Cdc2/Cyclin-Dependent Kinase 1 Mediates Disruption of Nuclear Lamina during Human Cytomegalovirus Nuclear Egress

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Abstract

The nuclear lamina is a major obstacle encountered by herpesvirus nucleocapsids in their passage from the nucleus to the cytoplasm (nuclear egress). We found that the human cytomegalovirus (HCMV)-encoded protein kinase UL97, which is required for efficient nuclear egress, phosphorylates the nuclear lamina component lamin A/C in vitro on sites targeted by Cdc2/cyclin-dependent kinase 1, the enzyme that is responsible for breaking down the nuclear lamina during mitosis. Quantitative mass spectrometry analyses, comparing lamin A/C isolated from cells infected with viruses either expressing or lacking UL97 activity, revealed UL97-dependent phosphorylation of lamin A/C on the serine at residue 22 (Ser22). Transient treatment of HCMV-infected cells with maribavir, an inhibitor of UL97 kinase activity, reduced lamin A/C phosphorylation by approximately 50%, consistent with UL97 directly phosphorylating lamin A/C during HCMV replication. Phosphorylation of lamin A/C during viral replication was accompanied by changes in the shape of the nucleus, as well as thinning, invaginations, and discrete breaks in the nuclear lamina, all of which required UL97 activity. As Ser22 is a phosphorylation site of particularly strong relevance for lamin A/C disassembly, our data support a model wherein viral mimicry of a mitotic host cell kinase activity promotes nuclear egress while accommodating viral arrest of the cell cycle.

Introduction

Human cytomegalovirus (HCMV) is a pathogen that is especially dangerous in immunocompromised individuals [1]. As is true for all viruses, HCMV replication depends on the interplay between viral and host cell functions. An important example of this interplay is nuclear egress, a stage during which herpesviral DNA-containing capsids (nucleocapsids) exit the nucleus [2]. An important obstacle for the exiting nucleocapsids is a meshwork underlying the inner nuclear membrane known as the nuclear lamina, whose principal components are intermediate-filament proteins known as lamins [3,4]. There are two major classes of lamins in mammalian cells: A-type lamins, which comprise the four lamins encoded by alternative splicing from the LMNA gene, lamin A, LMNA, LMNB1, and LMNB2 genes. A major function of lamins is to help maintain the structure of the nuclear envelope. Accordingly, along with the nuclear envelope, the nuclear lamina must be disassembled during mitosis and then reassembled after mitosis. These dynamic processes are regulated by phosphorylation of lamins. In particular, it is well established that Cdc2/cyclin-dependent kinase (CDK) 1 disassembles nuclear lamina by phosphorylation of specific sites on lamins during mitosis [5,6,7]. CDK1 phosphorylation of lamin A/C at Ser22, and of lamin B at the equivalent position, have been shown to be especially crucial for lamin disassembly [3,8]. It is thought that phosphorylation at this site interferes with head-to-tail interactions between lamins (reviewed in [3,4]). HCMV arrests...
Human cytomegalovirus (HCMV) causes life-threatening disease in transplant patients and people with AIDS, and is also an important cause of birth defects. Like all viruses, HCMV must have a way to leave the host cell, so that it can infect new cells. Moreover, as a member of the herpesvirus family, HCMV replicates its DNA in the nucleus, so it must have mechanisms to ensure that its genetic material can exit from the nucleus (nuclear egress). HCMV encodes a protein kinase, UL97, which is required for efficient nuclear egress. We found that UL97 aids nuclear egress by mimicking a host cell enzyme that normally helps break down a protein meshwork in the nucleus during cell division. The enzyme activity of UL97 poke holes in the meshwork that allow nascent HCMV virions to gain access to the nuclear membrane. UL97 is also an important target for drugs for treating HCMV disease. This work not only helps explain how these drugs act, but also highlights the potential of targeting nuclear egress for the discovery of new drugs.

**Author Summary**

Human cytomegalovirus (HCMV) causes life-threatening disease in transplant patients and people with AIDS, and is also an important cause of birth defects. Like all viruses, HCMV must have a way to leave the host cell, so that it can infect new cells. Moreover, as a member of the herpesvirus family, HCMV replicates its DNA in the nucleus, so it must have mechanisms to ensure that its genetic material can exit from the nucleus (nuclear egress). HCMV encodes a protein kinase, UL97, which is required for efficient nuclear egress. We found that UL97 aids nuclear egress by mimicking a host cell enzyme that normally helps break down a protein meshwork in the nucleus during cell division. The enzyme activity of UL97 pokes holes in the meshwork that allow nascent HCMV virions to gain access to the nuclear membrane. UL97 is also an important target for drugs for treating HCMV disease. This work not only helps explain how these drugs act, but also highlights the potential of targeting nuclear egress for the discovery of new drugs.

**Results**

Identification of lamin A/C as a candidate substrate of UL97

To investigate the role(s) of UL97 during HCMV infection, we searched for candidate substrates of UL97 by implementing a proteomics strategy. Human foreskin fibroblast (HFF) cells infected with wild-type (wt) HCMV strain AD169 (multiplicity of infection [MOI] = 1), with a UL97 deletion mutant (RCΔ97) [24], or with wt virus under conditions where UL97 was pharmacologically inhibited using maribavir [23] were radiolabeled with 32P orthophosphate, and the proteins were separated on 2-dimensional gels (Figure S1). Spots containing labeled phosphoproteins from wt-infected cells that differed from those from the other conditions were excised from the gels, digested with trypsin, and analyzed by liquid chromatography-tandem mass spectrometry (LC-MS/MS). A number of polypeptides were identified; these are tabulated in Table S1. The protein for which the most peptides were identified was lamin A/C, which is a major component of the nuclear lamina [3,4]. Given that HCMV UL97 is important at the stage of nuclear egress [19] and that the nuclear lamina forms a barrier to nuclear egress, we hypothesized that UL97 phosphorylates lamin A/C to mediate nuclear egress.

**Purified UL97 phosphorylates lamin A in vitro on sites phosphorylated by CDK1**

A previous report [20] described the phosphorylation of lamins in anti-FLAG immunoprecipitates from lysates of cells in which FLAG-UL97 was expressed by transfection, but only when lamins were simultaneously immunoprecipitated by anti-lamin antibodies. No controls in which lamins were immunoprecipitated, but UL97 was absent or inactive were shown. Thus, it was not demonstrated that UL97 directly phosphorylates lamins. Additionally, sites of phosphorylation were not reported.

We wished to determine whether UL97 can directly phosphorylate the largest of the A-type lamins, lamin A, in vitro. We expressed lamin A fused to a histidine tag at its N-terminus, and purified it from *E. coli* (Figure S2). When incubated with purified enzymatically active UL97 fusion protein (GST-UL97) and γ-[32P]-ATP, both GST-UL97 and lamin A became radiolabeled (Figure 1A). However, labeling of lamin A was not observed in the absence of added enzyme, and was almost completely abolished when an equivalent quantity of a catalytically deficient mutant form of GST-UL97 (K355Q) was used in place of wild-type GST-UL97 (Figure 1A). Moreover, treatment with maribavir, a specific inhibitor of UL97 activity, reduced phosphorylation of lamin A by GST-UL97 with a dose-dependence very similar to its inhibition of autophosphorylation of GST-UL97 (Figure 1B). Thus, UL97 phosphorylates lamin A in *vivo*.

To map the sites on lamin A that are phosphorylated by UL97 in vitro, proteins from kinase reactions, such as those conducted in Figure 1A — except using unlabeled ATP — were digested with trypsin. Phosphopeptides were enriched, and the peptides analyzed by LC-MS/MS. Two major phosphorylated peptides on lamin A were reproducibly detected following phosphorylation by GST-UL97. One of these peptides (residues 12–25) was unambiguously phosphorylated on Ser22, a site whose phosphorylation by CDK1 is crucial for lamin disassembly [5]. A representative MS/MS spectrum showing Ser22 phosphorylation in tryptic peptides released from lamin A incubated with GST-UL97 is presented in Figure 2A. A second major peptide including Ser390 and Ser392 was completely phosphorylated, but in the initial analysis there was ambiguity as to which of these serines was phosphorylated (see further analysis below). Three other phosphorylated peptides—one containing Ser19, one Ser23, and one Ser552 (Table 1) — were detected more than once, but less frequently than the two major peptides. No phosphopeptides were reproducibly detected in reactions using GST-UL97 K355Q or in reactions containing GST-UL97 and 1 μM maribavir (data not shown).
In order to definitively assign the site of in vitro phosphorylation by GST-UL97 on the second major peptide, we used an isotopically labeled synthetic peptide phosphorylated at Ser\(^{390}\) to identify fragment ions diagnostic for this phosphorylation site. As an additional control, we also treated lamin A with a commercial preparation of CDK1/cyclin B\(_1\) complex. For both GST-UL97 (GST97 WT), GST-UL97 K355Q or wild-type GST-UL97 were also incubated in kinase buffer without lamin A. Following termination of kinase reactions, proteins were resolved by SDS-PAGE. Signal from incorporation of \(^{32}\)P was detected by exposure to a phosphoscreen (top panel), and total protein was detected by Coomassie brilliant blue staining (bottom panel). The positions of radiolabeled GST-UL97 (GST97) and lamin A, and Coomassie stained lamin A are indicated. (The amounts of GST-UL97 were too small to see on the stained gel.) (B) UL97 autophosphorylation and labeling of lamin A were quantified following in vitro kinase reactions in the presence of varying concentrations of maribavir (MBV). Signal detected from \(^{32}\)P incorporation for autophosphorylation of GST-UL97 and phosphorylation of His-tagged lamin A are plotted as a percent of the signal detected in the absence of drug. The results taken together show that UL97 phosphorylates lamin A in vitro.

Figure 1. In vitro phosphorylation of lamin A by GST-UL97. (A) Recombinant His-tagged lamin A was incubated in kinase reaction buffer in the presence of \(^{\gamma}\)\(^{32}\)P-ATP either alone (no kinase), with catalytically deficient GST-UL97 K355Q (K355Q), or with wild-type GST-UL97 (GST97 WT). GST-UL97 K355Q or wild-type GST-UL97 were also incubated in kinase buffer without lamin A. Following termination of kinase reactions, proteins were resolved by SDS-PAGE. Signal from incorporation of \(^{32}\)P was detected by exposure to a phosphoscreen (top panel), and total protein was detected by Coomassie brilliant blue staining (bottom panel). The positions of radiolabeled GST-UL97 (GST97) and lamin A, and Coomassie stained lamin A are indicated. (The amounts of GST-UL97 were too small to see on the stained gel.) (B) UL97 autophosphorylation and labeling of lamin A were quantified following in vitro kinase reactions in the presence of varying concentrations of maribavir (MBV). Signal detected from \(^{32}\)P incorporation for autophosphorylation of GST-UL97 and phosphorylation of His-tagged lamin A are plotted as a percent of the signal detected in the absence of drug. The results taken together show that UL97 phosphorylates lamin A in vitro.

doi:10.1371/journal.ppat.1000275.g001

In order to definitively assign the site of in vitro phosphorylation by GST-UL97 on the second major peptide, we used an isotopically labeled synthetic peptide phosphorylated at Ser\(^{390}\) to identify fragment ions diagnostic for this phosphorylation site. As an additional control, we also treated lamin A with a commercial preparation of CDK1/cyclin B\(_1\) complex. For both GST-UL97 and CDK1/cyclin B\(_1\) treated samples of recombinant lamin A, no phosphopeptides matching the fragmentation pattern for phosphorylation at Ser\(^{392}\) were detected. Instead, a tryptic peptide (387–397) with phosphorylation at Ser\(^{392}\) with phosphorylation at Ser\(^{390}\) was detected (Table 1, Figure 2B and 2C). Evidently, phosphorylation at Ser\(^{390}\) by UL97 was again detected (Figure 2D provides an example from an iTRAQ labeling experiment — see below). A phosphopeptide containing either Ser\(^{390}\) or Ser\(^{392}\) was also detected, but in this case, with the aid of the isotopically labeled synthetic peptide phosphorylated at Ser\(^{392}\), the site of phosphorylation was unambiguously identified as Ser\(^{392}\) (Figure 2E). The longer tryptic peptide containing Ser\(^{390}\) that was observed following in vitro phosphorylation was not detected in the lamin A/C immunoprecipitates. Four other phosphorylation sites were also detected (Ser\(^{391}\), Ser\(^{400/401}\), Ser\(^{423}\), Thr\(^{424}\)), which are either identical or in close proximity (Ser\(^{401}\)) to sites previously detected in studies of phosphorylated proteins from human cells not infected by HCMV [26,27,28,29].

As mutational analysis implicates Ser\(^{22}\) and to a lesser extent Ser\(^{392}\) in lamin A/C disassembly [5], we employed an iTRAQ\(^{TM}\)-based quantitative MS approach [30] to evaluate whether levels of phosphorylation on these Ser residues were influenced by UL97 during HCMV replication. Cells were infected at an MOI of 1 with wt HCMV or a UL97 deletion mutant, Δ97 [31] or a virus in which the deletion mutation was rescued [31] and lamin immunoprecipitates were prepared at 72 hours p.i. Following digestion with trypsin, peptides from each sample were labeled with a sample-specific iTRAQ reagent. Stable isotopes incorporated into these reagents permit pooling of samples after labeling and subsequent relative quantification of phosphopeptide abundances by LC-MS/MS. In two separate experiments, while levels of Ser\(^{392}\) phosphorylation did not vary between lamin A/C isolated from wt-infected vs. Δ97-infected cells, there were 2–3 fold higher levels of phosphorylation of Ser\(^{22}\) in lamin A/C from wt HCMV-infected cells compared to lamin A/C from Δ97-infected cells (Table 2). Furthermore, lamin A/C from cells infected with the virus in which the deletion was rescued also showed approximately 2-fold higher levels of Ser\(^{22}\) phosphorylation compared to that from cells infected with Δ97 (Table 2). Another pair of previously described mutant and rescuant viruses was also compared, one encoding a catalytically-deficient form of UL97 (AD K355Q), and the other a rescuant of AD K355Q, in which sequences encoding a wild-type UL97 were restored (AD Q355K) [22]. In this experiment, lamin A/C from cells infected with the mutant (AD Q355K) had 4.1-fold higher levels of Ser\(^{22}\) phosphorylation than that from AD K355Q-infected cells (Table 2). From these results, we conclude that there is UL97-dependent phosphorylation of lamin A/C at Ser\(^{22}\) during HCMV infection.

Transient inhibition of UL97 decreases lamin A/C phosphorylation during viral replication

As a second approach to investigating the dependence of lamin A/C phosphorylation on UL97, we performed radiolabeling
experiments with wt HCMV infected cells (MOI = 3) in the presence or absence of 1 μM maribavir, which specifically inhibits UL97 [23,32], or 15 μM roscovitine, which specifically inhibits CDKs including CDK1 [33,34]. To minimize the effects of kinase inhibition on earlier stages of the HCMV replication cycle, infection was allowed to proceed for 69 hours p.i., at which time nuclear egress has commenced. The infected cells were then treated with the inhibitors or mock-treated with DMSO-

Figure 2. Phosphorylation sites detected on lamin A/C from HCMV-infected cells and from UL97 treated lamin A in vitro. (A) Mass spectrum from electrospray ionization (ESI)-MS-MS of SGAQASSTPLpSPTR tryptic peptide (amino acids 12–25 of native lamin A) after phosphorylation of His-lamin A in vitro with GST-UL97 indicating phosphorylation at Ser22. Diagnostic fragment ions are shown below the spectrum. (B) Mass spectrum from ESI-MS-MS of LRLpSPSPTSQR tryptic peptide (amino acids 386–397 of native lamin A) after phosphorylation of His-lamin A in vitro with GST-UL97, indicating phosphorylation at Ser390. Diagnostic fragment ions are indicated as above. (C) Mass spectrum from ESI-MS-MS of LRLpSPSPTSQR tryptic peptide (amino acids 386–397 of native lamin A) after phosphorylation of His-lamin A in vitro with recombinant human CDK1/cyclin B complex, indicating phosphorylation at Ser390. Diagnostic fragment ions are indicated as above. (D) Representative mass spectrum from ESI-MS-MS of SGAQASSTPLpSPTR tryptic peptide (amino acids 12–25 of native lamin A) after immunoprecipitation of lamin A/C from HCMV-infected cells and iTRAQ labeling. Diagnostic fragment ions are presented as above. Note that the iTRAQ label alters the mass of the b-ions. The masses of the ions from this spectrum are presented in Figure S3. (E) Representative mass spectrum from ESI-MS-MS of LSppSPTSQR tryptic peptide (amino acids 389–397 of native lamin A) after immunoprecipitation of lamin A/C from HCMV-infected cells. Diagnostic fragment ions are indicated as above. The results show that UL97, like CDK1 phosphorylates lamin A on Ser22 and Ser390 in vitro, but that phosphorylation occurs on Ser392 (and Ser22) in infected cells. doi:10.1371/journal.ppat.1000275.g002
were resolved using SDS-PAGE, and transferred to a membrane. Immunoprecipitated from each lysate, the immunoprecipitates of protein were present in each lysate (Figure 3C), Lamin A/C was detected in HCMV infected cells are indicated with a +. Those that were found in one setting, but not the other are indicated with a –. (Singly phosphorylated peptides with phosphorylation at Ser403, Ser404, Ser406, Ser407, or Thr409 were each detected. Phosphorylation could not be confidently assigned between Ser423 or Thr424. doi:10.1371/journal.ppat.1000275.t001)

| Table 1. A-type lamin phosphopeptides detected by mass spectrometry. |
|----------------|----------------|----------------|
| Position | Sequence | in vitro (UL97) | Infected cells |
| Ser22 | SGQAASSTIPspSPTTR | + | + |
| Ser390 | LRLpSPTSPSQR | + | – |
| Ser392 | LSpsPSTPSQR | – | + |
| Ser403-Thr409 | ApSpSHpSpSpTPQQGGSsVTk | – | + |
| Ser414 | A5HHSQQTQGGSsVTk | + | – |
| Ser423-Thr424 | KLEpSpTRES | + | + |
| Ser628 | pSVGSSGGGSSFGDNLVTR | + | – |
| Ser652 | SYLLGSNspSPR | + | + |

A-type lamin phosphopeptides detected after treatment with UL97 in vitro or detected in HCMV infected cells are indicated with a +. Those that were found in one setting, but not the other are indicated with a –.

containing vehicle for 1 hour, and then, in the presence or absence of the inhibitors, labeled with [32P]-orthophosphate for 2 hours. Following radiolabeling, cells were lysed. Probing western blots of the lysates with anti-actin antibodies showed that similar amounts of the inhibitors, labeled with [32P]-orthophosphate for 2 hours. Incorporation of radioactive phosphate into HCMV UL44 was reduced by 60–70% (Figure 3A, upper panel). (Again, there was more lamin A/C, present, radioactive phosphate incorporation was reduced by 60–70% (Figure 3A, upper panel). When both drugs were treated with maribavir compared to mock-treated infected cells, many of which exhibited a kidney shape. This shape was mirrored by those of the replication compartments stained with anti-UL44. Wt-infected cells treated with the UL97 inhibitor maribavir (Figure 4B) and cells infected with a UL97 null mutant virus (RCΔ97) (Figure 4C) showed much less dramatic shape changes, and while nuclei were observed to be somewhat enlarged relative to mock-infected cells, they often retained an oval shape. In addition, the anti-lamin staining appeared more uniform around the rim of the nuclei in mutant-infected cells, maribavir-treated wt-infected cells, or mock-infected cells than in wt-infected cells (compare Figure 4A to 4B, 4C and 4D). These kinds of changes in shape and

UL97 is required for alterations in the nuclear lamina during HCMV replication

Morphological deformations of cell nuclei are common in cells that are defective for lamin A/C [35]. Infection by HCMV has long been known to cause distortions in nuclear shape [36], and another study has reported that UL97 mutants do not induce such deformations [37]. Given our identification of lamin A/C as a substrate for UL97, we investigated whether these distortions were associated with alterations in the nuclear lamina. Therefore, we examined nuclear morphology during infection (MOI = 1) by staining the nuclear lamina with a polyclonal antibody against lamin A/C, while co-staining for sites of HCMV DNA replication (replication compartments) using a monoclonal antibody against the viral DNA polymerase subunit, UL44 (Figure 4). In immunofluorescence images of mock-infected cells (Figure 4D), the nuclei stained with the lamin antibody and were oval-shaped with no anti-UL44 staining. In wt HCMV-infected cells at 96 hours p.i. (Figure 4A), anti-lamin A/C staining revealed deformed nuclei, many of which exhibited a kidney shape. This shape was mirrored by those of the replication compartments stained with anti-UL44. Wt-infected cells treated with the UL97 inhibitor maribavir (Figure 4B) and cells infected with a UL97 null mutant virus (RCΔ97) (Figure 4C) showed much less dramatic shape changes, and while nuclei were observed to be somewhat enlarged relative to mock-infected cells, they often retained an oval shape. In addition, the anti-lamin staining appeared more uniform around the rim of the nuclei in mutant-infected cells, maribavir-treated wt-infected cells, or mock-infected cells than in wt-infected cells (compare Figure 4A to 4B, 4C and 4D). These kinds of changes in shape and

Table 2. Comparison of lamin A/C phosphorylation at positions Ser22 and Ser392 during replication of wild-type and UL97 mutant HCMVs.

<table>
<thead>
<tr>
<th>Position</th>
<th>Phosphopeptide abundance</th>
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<tbody>
<tr>
<td></td>
<td>AD169rv/197</td>
</tr>
<tr>
<td></td>
<td>Experiment 1</td>
</tr>
<tr>
<td>Ser22</td>
<td>2.3</td>
</tr>
<tr>
<td>Ser392</td>
<td>0.9</td>
</tr>
</tbody>
</table>

The ratios of abundances of the indicated phosphopeptides of lamin A/C from cells infected for 72 hours (MOI = 1) with the indicated pairs of viruses were determined using iTRAQ labeling and LC-MS/MS. doi:10.1371/journal.ppat.1000275.t002
anti-lamin staining could be observed in wt HCMV-infected cells by 24 hours p.i., although less consistently (data not shown).

We then used the same antibodies and confocal microscopy to investigate these staining patterns and their requirement for active UL97 in more detail. In wt HCMV-infected cells (MOI = 1; 96 hours p.i.), we observed thinning, invaginations (as recently reported [17]), and, in some cells, discrete gaps in anti-lamin staining (Figure 5A and 5B) compared to mock-infected cells (Figure 5E). In contrast, when wt HCMV infection was performed in the presence of maribavir or when UL97 was ablated genetically (two independent isolates of a UL97 deletion mutant), the anti-lamin staining of the nuclear rim resembled that of mock-infected cells with oval shaped nuclei and little or no thinning and few or no gaps (Figure 5C and 5D). To quantify the UL97-dependent changes in nuclear morphology, we surveyed approximately one hundred nuclei from each infection condition by confocal microscopy. These results indicated that nuclear deformities (Figure 6A) and gaps in the nuclear lamina large enough to be visible by light microscopy (Figure 6B) were each significantly more frequent under conditions where UL97 kinase activity was present ($p<0.001$). Thus, UL97 is required for nuclear deformation and disruptions of lamina during HCMV infection.

**Discussion**

We found that UL97 can directly phosphorylate lamin A/C in vitro on sites phosphorylated in vitro by Cdk1, including Ser22. We also detected UL97-dependent phosphorylation of Ser22 on lamin A/C in HCMV-infected cells. Additionally, phosphorylation of
lamin A/C was reduced by approximately two-fold when UL97 kinase activity was transiently inhibited in HCMV-infected cells. These findings taken together strongly suggest that lamin A/C is a bona fide, direct substrate of UL97 in infected cells.

Moreover, we found that UL97 is required in infected cells for discrete changes in the nuclear lamina, including gaps visible by confocal light microscopy. Given our results on lamin phosphorylation by UL97, we propose a model in which UL97 phosphorylates lamin A/C on Ser\textsuperscript{22}, a site whose phosphorylation is known to mediate disassembly of nuclear lamina [5,7]. These phosphorylation events contribute to localized disruptions in the nuclear lamina. These disruptions permit access of HCMV nucleocapsids to the inner nuclear membrane for primary envelopment and budding into the space between the inner and outer nuclear membranes, and thus nuclear egress. This model explains the requirement for UL97 for efficient nuclear egress [19].

We now discuss our results in terms of this model and previous results regarding lamin phosphorylation and nuclear egress.

UL97 mimics CDK1 for direct phosphorylation of lamin A/C

UL97 directly phosphorylates E. coli expressed lamin A in vitro with Ser\textsuperscript{22} and Ser\textsuperscript{390} being the major sites phosphorylated. Neither of the major sites conforms to a preference of UL97 for basic residues in the P+ position [32], although the sites do contain basic residues at P+6 and P+7. A minor site (Ser\textsuperscript{414}) phosphorylated by UL97 has an Arg at P+5. More importantly, the major sites phosphorylated by UL97 were also phosphorylated in vitro by CDK1/cyclin B\textsubscript{1}. This result and previous results with Rb [22] indicate that UL97 can exhibit at least some preference for sites that CDKs prefer.

UL97-dependent phosphorylation of lamin A/C at Ser\textsuperscript{22} in infected cells

We observed UL97-dependent phosphorylation in infected cells of a site that UL97 phosphorylates directly in vitro, and we found that transient inhibition of UL97 with maribavir could inhibit lamin A/C labeling by ~50%. The simplest interpretation of our data is that UL97 directly phosphorylates lamin A/C in HCMV-infected cells. An alternative interpretation is that UL97 acts indirectly, for example, by increasing the activity and/or expression of host cell kinases that phosphorylate lamin A/C, in particular CDK1. However, if UL97 were acting indirectly to increase lamin A/C phosphorylation by increasing CDK activity or expression, then roscovitine should have been at least as effective as at inhibiting lamin phosphorylation as maribavir. Our finding that maribavir was more effective than roscovitine is consistent with UL97 directly phosphorylating lamin A/C rather than increasing the activity or expression of a CDK. This interpretation is also consistent with a study that found that expression of a dominant negative CDK1 did not adversely affect replication of HCMV [30]; if CDK1 were required for disruption of the nuclear lamina during nuclear egress, such expression might have been expected to decrease HCMV replication.

Although only one site phosphorylated by UL97 in vitro showed dependence on UL97 for its phosphorylation in virus infected cells, it is striking that this position — Ser\textsuperscript{22} — is the lamin A/C phosphoacceptor site of greatest established relevance to dissolution of the nuclear lamina [5]. In particular, substitution of Ser\textsuperscript{22} with Ala results in a dominant negative mutant protein that inhibits lamina disassembly, while substitutions of Ser\textsuperscript{390} exert little effect on lamina disassembly by themselves [5]. Similarly, substitution of...
lamin B at Ser16, the position equivalent to Ser22 of lamin A/C, renders lamin polymers resistant to phosphorylation-mediated disassembly, while substitutions of serines equivalent to Ser200 and Ser392 have little or no effect [8]. Therefore, our results together with these previous studies strongly suggest that UL97 phosphorylation of lamin A/C in infected cells at Ser22 drives lamin A/C disassembly.

We also detected UL97-independent phosphorylation of lamin A/C in our infected cell preparations. This could be due to the activities of other kinases in infected cells, due to phosphorylated lamin A/C that existed prior to infection, and/or due to phosphorylation occurring in uninfected cells in our samples.

**Relationship of UL97 effects on lamins to nuclear egress**

In previous studies, it was reported that wt HCMV infection resulted in drastic reductions in the amounts of A-type lamins detected on western blots [20,39], and almost complete loss of lamin staining as detected by immunofluorescence [20], but that neither was observed during infection by a UL97 mutant [20]. However, we observed copious expression of A-type lamins during wt HCMV infection, and, similar to the recent report of Camozzi et al. [17], we observed only discrete changes in lamin staining in immunofluorescence experiments. The failure of others to detect lamin A/C in infected cells may be due to using antibodies that fail to recognize hyperphosphorylated A-type lamins, as previously asserted [20], or some other form of lamin A/C. Additionally, it has been reported that expression of UL97 following transient transfection induces a redistribution of lamin A/C from the nuclear rim to granular intranuclear structures [20,40]. Whether this was due to lamin A/C phosphorylation or due to toxicity (e.g. apoptosis) following UL97 overexpression was unclear. Regardless, we showed here that UL97 is required for disruptions in the nuclear lamina in HCMV-infected cells.

The simplest explanation for this requirement is that limited phosphorylation of lamins by UL97 causes local disassembly of the lamina. Although we detected gaps visible by confocal light microscopy in only a minority of cells, we emphasize that such gaps are much larger than an HCMV nucleocapsid. We think it is highly likely that nearly all infected cells contain smaller gaps in the lamina that could readily permit access of HCMV nucleocapsids to the inner nuclear membrane during nuclear egress. It may be advantageous for HCMV to induce only localized disruptions of the nuclear lamina. Lamins are important for cellular chromatin organization, DNA synthesis and transcription [3,33], and thus may play important roles in herpesvirus infection in addition to their service as barriers for nuclear egress. Indeed, during HSV infection, lamin A/C is required for proper targeting of the viral genome and for reduction of heterochromatin formation on viral promoters at early times of infection [41]. Nevertheless, there is more extensive disruption of the nuclear lamina during HSV infection than what we observed during HCMV infection [42,43].

The requirement for UL97 for the characteristic kidney-shape of nuclei in HCMV-infected cells is consistent with the results of Azzeh et al. [37]. The concave invaginations found in these nuclei are typically adjacent to juxtanuclear structures that appear to be cytoplasmic sites of virion assembly [44], and whose morphology depends on UL97 [37]. It is possible that the earliest UL97-mediated lamin disruptions (we detected gaps in some cells at 24 hours p.i. (data not shown)) direct the passage of primary enveloped virions into a particular portion of the perinuclear cytoplasm, and this contributes to the organization of the juxtanuclear structure. Alternatively or additionally, UL97 may prevent aggregation of tegument proteins in the nucleus [45] or be more directly required for formation of the juxtanuclear structure [37]. As noted by Azzeh et al. [37], the juxtanuclear structure seems to impact the nucleus. Thus, it may exert the force that causes deformation of the lamin-depleted nuclear envelope.

A considerable body of evidence establishes that HCMV UL50 and UL53 and their homologs in other herpesviruses play important roles in nuclear egress (reviewed in [2]). It has been proposed that these proteins and their homologs alter the nuclear lamina by recruiting PKCs to phosphorylate lamins [15,16,17,46,47]. However, PKC phosphorylation of nuclear lamins normally occurs during interphase and is not sufficient to cause dissolution of lamina [6,25]. Moreover, we did not observe phosphorylation of lamin A/C in HCMV-infected cells at sites known or likely to be phosphorylated by PKC, including Ser5, Thr480, Ser652, Thr656, Thr660, and Ser652 [7,25]. It may be that UL50 and UL53 act by recruiting PKC to disassemble nuclear lamina, but that the relevant substrate of PKC is lamin B [48] or other nuclear envelope components such as emerin [49,50]. Alternatively, the roles of UL50 and UL53 during nuclear egress may be independent of protein kinase recruitment. Recently, Camozzi et al. [17] reported that transient overexpression of HCMV UL50 and UL53 was sufficient to induce changes in the distribution of lamin A/C akin to what is observed during HCMV infection. Regardless, during HCMV infection, in the absence of UL97, nuclei remain oval and lamin staining remains intact despite the presence of UL50 and UL53. Thus, much as CDK1 is the crucial kinase for dissolution of lamina during mitosis, UL97, which mimics CDK1 for phosphorylation of lamin A/C, is the crucial kinase for nuclear egress.

**Why mimic CDKs?**

Why has HCMV evolved to encode a kinase that mimics CDK1 for phosphorylation of lamin A/C? One possible explanation stems from HCMV arresting the cell cycle at the G1/S boundary [9,10,11]. CDK1 is most active in phosphorylating nuclear lamina during M-phase [6]. There are elevated levels of CDK1 and cyclin B1 in HCMV-infected cells [12,13,14], which may account for the phosphorylation of Ser22 and Ser392 on lamin A/C that we detected even in the absence of UL97. It is also possible that this phosphorylation contributes to disruption of the nuclear lamina. However, expression of a dominant negative CDK1 did not decrease HCMV replication [38], CDK1 and cyclin B do not appear to accumulate in the nuclei of infected cells to the extent seen in mitotic cells [14], and their elevated levels evidently are not sufficient to disrupt the nuclear lamina by themselves. Interestingly, CDK1 does appear to be required in HCMV-infected cells for the formation of so-called pseudomitoses, in which aberrant spindle poles and chromosome condensation occur [38], and CDKs that are sensitive to roscovitine are involved in several phases of HCMV replication [51].

UL97 mimicry of a CDK for phosphorylation of lamin A/C explains at least part of this viral enzyme’s role in HCMV infection. It is notable that UL97 is also required for phosphorylation of a second CDK substrate, Rb, in infected cells [22,52]. As yet, the importance of phosphorylation of Rb for HCMV replication has not been established. It will be interesting to elucidate further how HCMV uses its CDK-mimic to promote its propagation.

**Materials and Methods**

**Reagents**

All reagents were from Sigma unless otherwise specified.

**Cells and viruses**

HFF cells, isolate Hs27, (American Type Culture Collection) were cultured in Dulbecco’s modified Eagle’s medium (DMEM)
was kept for subsequent MS analysis. There were visualized by exposure to a phosphorscreen or Bio-Max Film.

Dilutions of the AD169 and containing the AD169rv, were used: A97, a BAC derived UL97 deletion mutant, FLAG97 a resistant of A97, AD K355Q, and AD Q355K [22,31]. Viral stocks were propagated and titrated as previously described [31].

Protein expression and purification

Two-dimensional (2D) gel analysis of phosphoproteins

Protein expression was induced with 0.5 mM isopropyl-β-D-thiogalactopyranoside (IPTG) for 20 h at 16°C. The resulting pellets were isolated and dissolved in 8 M urea, 50 mM Tris-HCl pH 8.0, 0.5 M NaCl, 1 mM dithioretilo (DTT), 20 mM imidazole and Complete™

Protein kinase assays

For radioactive in vitro kinase assays, 32 ng of GST-UL97 or GST-UL97 K355Q was used with 0.6 μg of His-lamin A per 20 μL reaction in 30 mM Tris (pH 6.5 at 37°C), 125 mM NaCl, 5% glycerol, 10 mM MgCl₂, 2 mM DTT, 5 mM betaglycerophosphate, 100 μM unlabeled ATP and 0.5 μL of [γ-32P]-ATP (3,000 to 6,000 Ci/mmol) (Perkin Elmer Inc., Waltham, MA) and incubated at 37°C for 90 min. For non-radioactive kinase reactions submitted for MS analysis, reactions were scaled up to 120 μL and 2 μg of UL97 or 160 ng of CDK1/cyclin B complex (Cell Signaling Technology, Inc., Danvers, MA) was used, and the final ATP concentration was adjusted to 200 μM. For comparison of maribavir inhibition of UL97 autophosphorylation versus lamin A phosphorylation, the same reaction conditions were used except 50 ng GST-UL97, 0.5 μg of His-lamin A, 50 mM Tris (pH 9.0 at 25°C), 300 mM NaCl, 1 mM DTT, 10 mM MgCl₂, 0.5 μL of [γ-32P]-ATP (3,000 to 6,000 Ci/mmol) (Dupont NEN) and specified concentrations of maribavir were combined in a final volume of 10 μL and incubated at 37°C for 2 h. Reactions were terminated by the addition of concentrated SDS-PAGE loading buffer. Samples were heated at 95°C for 5 min and proteins were resolved by SDS-PAGE. Gels were dried onto blotting paper under vacuum, and incorporated 32P was quantified with a phosphorimager (Molecular Imager FX System; Bio-Rad Laboratories Inc., Hercules, California).

Immunoprecipitation and western blotting

For immunoprecipitations of lamin A/C for MS analysis, HFF were infected at an MOI of 1.0 (confirmed by back-titration). At 72 h post infection, cells were rinsed. In non-radioactive experiments, cells were rinsed twice in ice-cold Dulbecco’s phosphate buffered saline prior to lysis. For radioactive experiments, cells were rinsed three times in 0.5 mL of ice-cold Dulbecco’s buffered saline (25 mM Tris pH 8.0, 4 mM KCl, 137 mM NaCl) prior to lysis. Cells were lysed for 30 min at 4°C in 0.5 mL of ice-cold modified radio-immunoprecipitation assay (RIPA) buffer per well. The RIPA buffer used was similar to one previously described [57] except that 300 mM NaCl was used, Triton-X 100 (1%) was used in place of NP-40, and the following components were added: 10 mM EDTA, 10% glycerol, 10 mM betaglycerophosphate, 5 mM NaF, 10 μg/mL each of leupeptin and aprotinin, 1 μg/mL pepstatin A, 10 μM E-64, 2 mM imidazole, 1.2 mM sodium molybdate, 0.5 mM sodium orthovanadate, 4 mM sodium taatrte, 1 mM DTT, and Calbiochem Phosphatase Inhibitor Set I (EMD Chemicals Inc., Gibbstown, New Jersey) was added at 1:100. Lysates were collected and clarified at 10,000 g at 4°C and supernatants were transferred to new tubes, flash frozen on liquid N₂, and stored at ~−80°C until use.

Two hundred microliters of each thawed lysate was pre-cleared, with rotation, for 30 min at 4°C with 10 μg each of purified mouse IgG₁ and IgG₂a (Bethyl Labs Inc., Montgomery, Texas) and 20 μL of settled Immunopure Protein A/G (Pierce) in a final volume of 0.5 mL (adjusted by adding RIPA buffer). Then, 220 μL of each sample was added to 15 μL of Protein A/G, which
had been pre-incubated with 10 μL JOL2 and 5 μL JOL4 mouse anti-lamin A/C monoclonal antibodies (Millipore Inc., Billerica, Massachusetts); IP reactions were adjusted to a final volume of 0.5 mL with RIPA buffer and allowed to rotate for 1 h at 4°C. IP reactions were then subjected to four 5 min washes in 0.5 mL RIPA buffer and then incubated at 05°C for 5 min in 80 μL of 2× SDS-PAGE sample buffer [57] supplemented with 5% betamercaptoethanol. Immunoprecipitation of lamin A/C from non-radioabeled cells was performed essentially as above except without pre-clearing.

For immunoprecipitation of HCMV UL44, each sample of cells was lysed in 1 mL EBC2 lysis buffer (50 mM Tris [pH 8.0], 30 mM NaCl, 0.5% NP-40, 2 mM EDTA and 2 mM DTT) supplemented with 10 μg/mL each of leupeptin and aprotonin, 1 μg/mL pepstatin, 25 mM betaglycerophosphate, 0.5 mM sodium orthovanadate, and 1:100 of Phosphatase Inhibitor Cocktail 1 and 2 (Sigma). Lysates were pre-clear and then reacted with anti-UL44 antibody 28-21 (kindly provided by William Britt, U. of Alabama) pre-conjugated to Protein A beads. The immunoprecipitations were washed four times in 1 mL cold EBC2 lysis buffer, and resuspended in 40 μL 2× SDS-PAGE sample buffer.

SDS-PAGE and western blotting procedures were carried out as previously described [58] using goat anti-lamin A/C polyclonal antibody N-18 (Santa Cruz Biotechnology), anti-CMV ICP36 (UL44) monoclonal antibody-VirusSelect (Virsys), or anti-beta-actin mouse monoclonal antibody (Abcam) to probe immunoblots.

**MS analysis**

In early experiments, following SDS-PAGE, lamin A from in vitro phosphorylation reactions or lamin A/C from infected cells in gel slices was red and alkylated by DTT and iodoacetamide, respectively. The gel slices were then dehydrated in acetonitrile, and the protein digested with trypsin (500 ng/g slice) in NHCl/COH buffer (50 mM, 50 μL/slice) overnight at 37°C. Trypsin-digested peptide samples were then enriched for phophopeptides using a phopho-peptide isolation kit (Pierce). The samples were mixed with binding buffer (10% acetic acid), followed by addition of sample mix (50 μL) to SwellGel Disc (Pierce) resin. The sample-resin mixture, whose pH was maintained below 3.5, was incubated for 8 minutes at room temperature with regular gentle stirring. Resin was washed twice in 50 μL of 0.1% acetic acid, and twice in 50 μL of 0.1% acetic acid, 10% acetonitrile. Phosphopeptides were eluted in elution buffer (50 μL, 100 mM ammonium bicarbonate, pH 9.0) after 5 minutes incubation. Eluted phosphopeptides were analyzed by LC-MS/MS at the Taplin Biological Mass Spectrometry Facility at Harvard Medical School.

Subsequent experiments were performed using different protocols depending on whether phosphorylation was assessed following in vitro phosphorylation or following phosphorylation in infected cells. For analyses of in vitro phosphorylated samples, the kinase reactions were treated with 1 μg trypsin (Promega) in 100 mM ammonium bicarbonate pH 8.0 (overnight, 37°C), and lyophilized. Free peptide carboxylate groups were converted to their corresponding methyl esters and the derived peptides were subjected to phosphopeptide enrichment as described previously [59]. Captured phosphopeptides were eluted with 50 mM PO4 buffer pH 8.0 directly onto a 100 μm (i.d.)×8 cm fused silica capillary desalting precolumn (PC) packed with 10/25 irregular-shaped C18 beads. The PC was then connected to a 50 μm×65 cm (5 μm spherical C18 beads) analytical column. LC-MS/MS was performed on a QSTAR XL (MDS SCIEX). The HPLC solvent gradient was 0–7% B in 5 min, 7–63% B in 30 min; solvent A was 0.2 M acetic acid and solvent B was 70% acetonitrile/0.2 M acetic acid.

In later experiments aimed at comparing the phosphorylation sites of UL97 and CDK1 on lamin A/C, in vitro phosphorylated lamin A/C was digested as described above and each digest was analyzed separately on a LTQ-Orbitrap mass spectrometer using a method consisting of two data-dependent MS/MS scans, followed by targeted MS/MS scans on precursors corresponding to lamin A/C peptides containing Ser25, Ser392, their phosphorylated counterparts, and a synthetic phosphopeptide containing 613C and 4 15N atoms. This synthetic peptide was loaded onto the PC as an internal control. For analysis of lamin A/C phosphorylated in cells, lamin A/C immunoprecipitates was processed for subsequent iTRAQ (Applied Biosystems) isotope labeling according to the manufacturer’s protocol. Briefly, each sample was separately reduced using 5 mM Tris-(2-carboxyethyl)phosphine (TCEP) (1 hr, 60°C), alkylated using 10 mM methyl methane-thiosulfonate (MMTS) (10 min, room temperature) and digested on the beads with trypsin (Promega) in 0.5 M triethylammonium bicarbonate pH 8.5 (overnight, 37°C). The resulting peptides were then labeled with iTRAQ-114 (mock), iTRAQ-115 (AD K355Q), iTRAQ-116 (AD) and iTRAQ-117 (AD169or) reagents in 70% ethanol, respectively, for 1 hr at room temperature. Following iTRAQ labeling, all four samples were combined and dried by centrifugal concentration (Thermo Savant, Holbrook, New York). To enrich for phosphorylated lamin A/C peptides, 100 μl of 100 mg/ml MassPREP Enhancer in 80% acetonitrile/0.1% trichloroacetic acid (TFA) were added. After sonication (20 min), the mixed sample was loaded on a well of a TiO2 96-well plate (Wars, Milford, Massachusetts). Peptides were eluted with 100 μl of 300 mM ammonium hydroxide. Following elution, 3 μl of TFA were added to the sample to bring the pH to 2.0 and the sample volume was reduced to ~20 μl by centrifugal concentration. LC-MS/MS was performed on a QSTAR XL using a Top 3 method. A similar protocol was followed for analysis of AD169 (iTRAQ 114), A97 (iTRAQ 115), FLAG97 (WT rescue of A97) (iTRAQ 116), mock-iTRAQ 117, AD K355Q-iTRAQ 114 and AD Q355K-iTRAQ 115 HCMV-infected cells, except that a targeted MS/MS method (on phosphorylated Ser25 and Ser392) was used for the phosphopeptides and a Top 6 was used for the TiO2 loading flowthrough. Non-phosphorylated lamin peptides detected in the loading flowthrough were used to normalize abundances of phosphorylated peptides. As described above, an isotopically labeled synthetic phosphopeptide containing a phosphate on Ser392 was loaded onto the PC as an internal standard. MS/MS spectra were searched against an in-house lamin A/C database and the human RefSeq database using the Mascot algorithm. Putative lamin A/C phosphopeptide sequences were manually confirmed.

**Transient inhibition of protein kinase activities in cells**

2×10^3 HFF per well of a 24 well cluster plate were infected at a MOI of 3 (confirmed by back titration) and then incubated at 37°C in DMEM supplemented with 5% FBS. Sixty-nine hours post infection, medium was removed and cells were washed with phosphate-free DMEM (Invitrogen Inc.) and incubated for 1 hour in 0.5 mL phosphate free medium containing 2% FBS, supplemented with 15 μM roscovitine or 1 μM maribavir, both drugs, or no drug. The concentration of DMSO was adjusted to a final concentration of 0.2% (vol/vol) to control for any effects of the carrier. Five hundred microcuries of 32P-labeled orthophosphoric acid (8500–9120 Ci/mmol) was then added in 0.5 mL of phosphate free medium containing 2% FBS and the same drug condition used during the pre-incubation step and left on the cells.
for 2 h. Lamin A/C and HCMV UL44 were immunoprecipitated as described above and analyzed by SDS-PAGE and autoradiography and western blotting as described above.

**Immunofluorescence**

HFF were seeded at 1 × 10⁵ cells/well on glass coverslips in 24-well dishes and allowed to attach overnight prior to infection at an MOI of 1. At 96 h post infection, cells were fixed in 4% formaldehyde in PBS for 20 min (unless otherwise indicated, all steps were performed at room temperature). Cells were permeabilized with acetone at −20°C for 2 min. Following several washes in PBS, cells were blocked overnight in IF buffer (PBS containing 4% FBS [Sigma]). Primary antibodies were diluted in IF buffer and incubated with fixed cells for 1 h. Lamin A/C goat polyclonal antibody N-18 (Santa Cruz Biotechnology, Inc.) was used at 1:10 dilution and UL44 mouse monoclonal antibody 10-C50 ( Fitzgerald-Ald Industries International Inc.) was used at 1:100. Secondary antibodies (Alexa Fluor 568-rabbit anti-mouse IgG and Alexa 488-chicken anti-goat IgG, Invitrogen) were applied at 1:1,000. Cells were then washed in IF buffer three times for 5 min per wash, and mounted on slides in Prolong Antifade reagent (Molecular Probes, Inc.). Fluorescence microscopy was performed in the Nikon Imaging Center at Harvard Medical School, using a Nikon TE2000U inverted microscope in conjunction with a PerkinElmer Ultraview spinning disk confocal system equipped with a Hamamatsu Orca ER cooled-CCD camera. Images were acquired using a 60× differential interference contrast oil immersion objective lens and analyzed using Metamorph software from Universal Imaging, Inc. (Downington, Pennsylvania). Fisher’s exact test was performed using Prism 4 (GraphPad Software, Inc.) for Macintosh.

**Supporting Information**

Table S1 Candidate UL97 substrates. Protein description, database hit, and peptides detected for those polypeptides for which at least two distinct peptides were detected in one or more experiments are shown. Found at: doi:10.1371/journal.ppat.1000275.s001 (0.05 MB DOC)

Figure S1 2D electrophoresis. Samples from HFF cells infected with HCMV wt strain AD169 in the absence of drug (left-most panels), UL97 null mutant RCA97 in the absence of drug (middle panels), and AD169 in the presence of 1 μM maribavir (MBV; right-most panels) that had been radiolabeled with 32P orthophosphate were resolved by SDS-PAGE alongside molecular weight markers in the second dimension (arrows to the left of the panels point to lower molecular weight [Mr]; Top panels (A) show autoradiograms of the 2D gels; the red rectangles indicate the areas from which spots from wt-infected cells differed from those in the other two samples were taken. Bottom panels (B) show an expanded region of the gels in the top panels. Spots numbered 1–6 from wt-infected cells were excised from the gels and submitted for MS analysis. Lamin A/C peptides were found in spot 6. Found at: doi:10.1371/journal.ppat.1000275.s002 (8.95 MB TIF)

Figure S2 Purification of recombinant lamin A. His-lamin A was expressed in E. coli, the bacterial cells were lysed and His-lamin A/C was purified as a fusion protein using nickel affinity chromatography as described in Materials and Methods. Aliquots of the initial crude protein extract derived from inclusion bodies (crude), the flow through, the first wash (wash 1), the second wash (wash 2), and the eluate (lanes indicated above the image of the gel) were resolved by SDS-PAGE alongside molecular weight markers (M) and detected by Silver Blue staining. The position of His-lamin A is indicated to the right of the gel. By this procedure, sufficient amounts of reasonably pure lamin A were obtained. Found at: doi:10.1371/journal.ppat.1000275.s003 (0.03 MB PDF)

Figure S3 Diagram of predicted fragment ions from spectrum in Fig. 2D. The sequence of the peptide is shown on the top line, with the position of the iTRAQ label indicated. Below are the masses of the various series of ions. Found at: doi:10.1371/journal.ppat.1000275.s004 (2.11 MB TIF)

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**Author Contributions**

Conceived and designed the experiments: SH JPK YMNC AJL WJJ MCB DMK DEG JAM DMC. Performed the experiments: SH JPK YMNC AJL WJJ MCB LAS DMK JAM DMC. Contributed reagents/materials/analysis tools: DMK DEG JAM. Wrote the paper: SH JPK YMNC JAM DMC. Suggested revisions: WJJ SN LAS MCB SN LAS MSH DMK DEG JAM DMC.

**References**


