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Heritable Stochastic Switching Revealed by Single-Cell Genealogy

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The partitioning and subsequent inheritance of cellular factors like proteins and RNAs is a ubiquitous feature of cell division. However, direct quantitative measures of how such nongenetic inheritance affects subsequent changes in gene expression have been lacking. We tracked families of the yeast Saccharomyces cerevisiae as they switch between two semi-stable epigenetic states. We found that long after two cells have divided, they continued to switch in a synchronized manner, whereas individual cells have exponentially distributed switching times. By comparing these results to a Poisson process, we show that the time evolution of an epigenetic state depends initially on inherited factors, with stochastic processes requiring several generations to decorrelate closely related cells. Finally, a simple stochastic model demonstrates that a single fluctuating regulatory protein that is synthesized in large bursts can explain the bulk of our results.

Introduction

Inheritance is more than the faithful copying and partitioning of genomic information. When cells divide, the mother cell passes numerous other cellular components to the freshly born daughter, including nucleosomes, transcription factors, mitochondria, and substantial fractions of its proteome and transcriptome. In this way, an entire pattern of gene expression can be passed from mother to daughter, a phenomenon known as epigenetic or non-Mendelian inheritance. Classic examples permeate the literature and include the sex-ratio disorder in Drosophila [1], the yellow-tip phenotype in melons [2], the telomere position effect in yeast [3] and mouse [4], and prions such as Psi+ in yeast [5].

The time scale over which epigenetic phenotypes may persist spans many orders of magnitude and depends strongly on the physical mechanism used by the cell [6]. In general, however, epigenetic phenotypes are substantially less stable than chromosomally inherited ones are [6,7], and can change reversibly in single cells [3,8,9] during development [10,11], or in mature organisms [12].

Beginning with landmark studies on the lac operon in the 1950s, positive transcriptional feedback loops have emerged as a means to store cellular memory [13–15]. Such epigenetic inheritance systems are frequently described as “bistable,” meaning that transcriptional activity of genes in the network tends to become fixed in single cells around one of two stable levels (ON and OFF), each of which is able to stably persist for many generations [8,16,17]. Stochastic fluctuations in the creation or decay of the proteins involved [18–34], or changes in external cues (e.g., a changing environment), are responsible for causing transitions between the two states [8,13,16,17].

This flexible strategy, which is present in both prokaryotes and eukaryotes, allows genetically identical cells to diversify their population, possibly allowing them to exploit new environmental niches or to survive in a fluctuating environment [35]. Feedback-based cellular memories show an exceptional range of stability; depending on the strength of the feedbacks, cells may display memory of a previous expression state as short as a single generation to as far back as many thousands of generations [17]. However, quantitative measurements of phenotype stability, switching, and heritability are rare, both because detailed genealogical relationships are challenging to produce in single cells [36] and because reporters indicating degree of inheritance are not always available.

To measure how a dynamic gene expression state is inherited, we focused on an engineered version of the galactose utilization (GAL) pathway in the yeast Saccharomyces cerevisiae (Text S1). We disrupted the pathway’s major negative feedback loop and grew cells in conditions where only a single positive-feedback loop was operational (see Materials and Methods). Under these conditions, cells stochastically transition between two distinct expression states even in the absence of an extracellular trigger. These infrequent switching events therefore likely arise from fluctuations in concentrations of regulatory proteins within the individual cells [37]. We are able to monitor transitions between ON and OFF using a fluorescent reporter (see Materials and Methods, Figure S3). Together, these attributes make our network an ideal model system to investigate the interplay between genetic and epigenetic inheritance.

Abbreviations: GAL, galactose utilization; GM-GD, grandmothers with granddaughters; M-D, mothers with daughters; S1-S2, older siblings with younger siblings

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When cells divide, not only DNA but an entire pattern of gene expression can be passed from mother to daughter cell. Once cell division is complete, random processes cause this pattern to change, with closely related cells growing less similar over time. We measured inheritance of a dynamic gene-expression state in single yeast cells. We used an engineered network where individual cells switch between two semi-stable states (ON and OFF), even in a constant environment. Several generations after cells have physically separated, many pairs of closely related cells switch in near synchrony. We quantified this effect by measuring how likely a mother cell is to have switched given that the daughter cell has already switched. This yields a conditional probability distribution that is very different from the exponential one found in the entire population of switching cells. We measured the extent to which this correlation between switching cells persists by comparing our results with a model Poisson process. Together, these findings demonstrate the inheritance of a dynamic gene expression state whose post-division changes include both random factors arising from noise as well as correlated factors that originate in two related cells’ shared history. Finally, we constructed a model that demonstrates that our major findings can be explained by burst-like fluctuations in the levels of a single regulatory protein.

Author Summary

In this work, we find that not only is the epigenetic phenotype itself heritable, but that the stability of this phenotype is likewise a heritable quantity. In other words, when cells divide, the nascent daughter cell assumes both the expression state of the mother cell as well as its tendency to switch epigenetic states at a similar time in the future. This is surprising, especially considering that individual cells viewed outside their genealogical context appear to switch completely at random. We resolve this apparent contradiction using a simple stochastic model.

Results

Heterogeneous Populations Are Generated from Single Progenitors that Spontaneously Switch between Two Phenotypes

We first set out to quantify, using fluorescence microscopy, the infrequent switching events that occur at random times. All experiments began with a single cell confined between a cover slip and a thick agar pad. Over a period of about 920 min (>15 h) each cell grew and divided to eventually form a small colony of 50–100 cells. Throughout the measurement period, these cells diverged in behavior, with some increasing in fluorescence and others decreasing. We repeated this process with more than 100 progenitor cells, so in sum our data represent many thousand single-cell trajectories.

We present two examples of the experimental procedure in Figure 1. In Figure 1A, an initially bright cell develops into a small colony with distinct subpopulations. The dim cells in the lower subpopulation continue to diminish in fluorescence with each successive cell division as the remaining molecules of green fluorescent protein (GFP) dilute. In Figure 1B, an initially faint cell likewise gives rise to a variegated colony with cells both dim and bright. Together, these two processes generate a broad bimodal steady-state distribution.
reverse switching distribution, composed of ON cells switch-
ing into the OFF state, could not be obtained in this simple
way, because in this scenario the long life of the fluorescent
proteins makes it difficult to determine the exact moment
cells cease production of yellow fluorescent protein (YFP).

Apparently Random Switches Are Heritable

This exponentially distributed switching pattern applies to
cells chosen at random without regard to genealogy. How-
ever, measuring cells instead on the basis of their family
history paints a very different picture. To demonstrate this
difference, we asked how likely a mother and a daughter cell
were to have both switched within a small window of time
after the cells divided. We selected all daughter cells with
marginal switch times below some value $T$, and then we
measured what percent of their mothers had also switched at
or before that time. The results, summarized in Figure 3B
(open circles), show that when a daughter switches shortly
after cell division, its mother cell is overwhelmingly likely to
switch as well. For example, of the daughters who switch within
400 min of cell division (about two generations), their
mothers have approximately a 50% chance of switching in
that same period. This represents a 2-fold increase in the
switching rate for a typical unrelated cell. As $T$ grows to
encompass an ever-larger fraction of all daughter cells, the
corresponding percent of switching mother cells asymptoti-
cally approaches the marginal switch distribution of Figure
3A (reproduced in black), which represents the limit of no
genealogical relationship. As in the marginal switch case
above, we are careful to weigh each of these mother-daughter
pairs according to how likely we were to experimentally
observe them.

To measure the underlying rates governing this process, we
examined the possible switching events diagrammed in
Figure 3A. In this simplified view, we assume cell pairs can
either switch together into the ON state at a rate $c(t)$,
or independently of one another at a rate $r/C_0$. In this way,
the total switch rate for any given cell sums to $r$ at all times,
as required by the marginal switch distribution. We assume that
the correlations decay with a rate $c(t) \sim r/C_1 e^{-t/\tau_c}$, which is
reminiscent of an Ornstein-Uhlenbeck process (Figure 3A)
[16,28]. The fixed delay of 20 min is included to account for
slow chromophore (YFP) maturation as observed in our data
daughters that switch within the first 20 min after cell
division have mothers that always switch). This model
includes two free parameters: $r$, the overall switch rate, and
$\tau_c$, the characteristic time for the correlation to decay. A
global least-squares fit to both curves (Figure 3B, red and blue
curves) simultaneously yields ($r = [7.0 \pm 0.5] \cdot 10^{-4}$ min$^{-1}$
$= 0.12 \pm 0.01$ gen$^{-1}$) and ($\tau_c = 197 \pm 54$ min). This
correlation rate is quite similar to the average cell doubling
time of 177 min (Text S2 and Figure S1), and similar
connections between doubling time and decorrelation have
been found in other protein regulatory networks [28].

Correlations of Switching Times between Cell Pairs Vary
by Genealogical Relationship

The above analysis suggests that when cell pairs do switch,
they will do so in synchrony. To demonstrate that this is

Figure 2. A Genealogical Switching History

We designate the first cell in each movie cell 1 and sequential daughters of that cell 1–1, 1–2, 1–3. These daughter cells bud in turn, giving rise to cells 1–1–1, 1–1–2, 1–2–1, etc.

(A) As in Figure 1, an initially OFF cell grows into a variegated micro-colony. Beginning at 600 min, or 4 generations, several cells fluoresce almost simultaneously. This includes the mother-daughter pairs (1,1–2) and (1–1–1,1–1–1–1). Conspicuously, cell 1–1 does not switch for the duration of our observation, even though its mother, daughter, and closest sibling all do.

(B) The family tree for colony in (A). Black lines indicate cells in the OFF state, whereas pink lines represent cells after they have switched to the ON state.

(C) Fluorescent time courses for mother cell 1 and her daughter 1–2, showing each as they switch into the ON state. The marginal switch times $\tau_1$ and $\tau_{1–2}$ run from cell birth until the beginning of the increase in fluorescence and do not depend on any other cells. The period labeled $\tau_{1–1–2}$ runs between the birth of cell 1–2 and the fluorescence of cell 1 and is an example of a conditional switch time.

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The cumulative percentage of cells that have switched is plotted against their marginal switch time. The black squares represent 251 switching cells, and the blue line is an exponential fit. The cyan dashed line is a result of our stochastic simulation (see Figure S). Error bars are derived from a bootstrap analysis. The fits are consistent with the idea that a constant-rate process may underlie the network. The inset shows ways that mother-daughter pairs may switch, either dependently via the center route or independently of one another via the outer routes.

(b) Gray circles describe the likelihood that a mother cell has switched given that its daughter cell is known to have switched before this time. The solid red line describes a two-parameter least-squares fit simultaneously to both curves with parameters described in the inset and main text. The dashed dark red line shows the fit resulting from the stochastic simulation. Black squares and blue lines are reproduced from (A) for comparison. doi:10.1371/journal.pbio.0050239.g003

Figure 3. Single-Cell Fate

(A) The cumulative percentage of cells that have switched is plotted against their marginal switch time. The black squares represent 251 switching cells, and the blue line is an exponential fit. The cyan dashed line is a result of our stochastic simulation (see Figure S). Error bars are derived from a bootstrap analysis. The fits are consistent with the idea that a constant-rate process may underlie the network. The inset shows ways that mother-daughter pairs may switch, either dependently via the center route or independently of one another via the outer routes.

(b) Gray circles describe the likelihood that a mother cell has switched given that its daughter cell is known to have switched before this time. The solid red line describes a two-parameter least-squares fit simultaneously to both curves with parameters described in the inset and main text. The dashed dark red line shows the fit resulting from the stochastic simulation. Black squares and blue lines are reproduced from (A) for comparison. doi:10.1371/journal.pbio.0050239.g003

Indeed the case, we turned our focus to the further subset of cell pairs where both cells are observed to switch during the experiment (and therefore ignoring cases where only one cell in a pair switches). More specifically, we concentrated on three cell relationships: mothers with daughters (henceforth M-D), grandmothers with granddaughters (GM-GD), and older siblings with younger siblings (S1-S2). Instead of marginal switching times, which are measured relative to each individual cell’s time of birth, we chose instead to compute the switch times of both cells relative to the moment when their two respective branches of the family tree first broke apart. Put another way, this quantifies the amount of time between a switching event and the last moment that switching events are measured relative to the same point for both cells. For M-D pairs, the time we use is simply the birth of the daughter; for GM-GD pairs, however, it is the birth of the intervening daughter; and for S1-S2 pairs, it is the older sibling’s birth. Formally we define the conditional switch time, $\tau_{MD}$, as the time elapsed between the fluorescing of cell X and the birth of cell Y. When X and Y both refer to the same cell, we recover the marginal switch time (i.e., $\tau_{MD} = \tau_X$).

Comparing M-D conditional switch times (Figure 4A), we observe nearly synchronous switching that extends at least 300 min and yields a correlation coefficient of $p_{MD} = 0.87$ ($p < 10^{-45}$). GM-GD and S1-S2 pairs (Figure 4B and 4C) show somewhat lower correlation coefficients of $p_{GMGD} = 0.74$ ($p < 10^{-9}$) and $p_{SS} = 0.60$ ($p < 10^{-7}$), respectively, although the total coefficient for all data combined remains a robust $p_{TOT} = 0.8$ ($p < 10^{-6}$). The strength and duration of these correlations are surprising and were not found in bacterial [16,25] and mammalian [38] studies, except in context of morphological traits [39]. Like the marginal switch data, these scatter plots should be viewed in the context of finite experimental viewing times, giving weights to points that are inversely proportional to the number of experimental opportunities to have seen them (Text S3, Figures S6 and S7).

Memory of Switching Persists for Several Generations

One dynamic measure for the randomness associated with the distribution is the average square difference of switch times for pairs of cells with comparable mean switch times (Figure 4D, blue curve). This curve rises rapidly at first, but at longer times it flattens out. This flattening is likely due, at least in part, to the limited duration of our experiments (on average 920 min), which constrains the scatter distribution to reside in the box shown in Figure 4A–4C.

To understand what this means, it is helpful to compare our results to those obtained using a stochastic Poisson model [40], where closely related cells are assumed to switch independently of one another and with constant probability in time (Text S3, Figures S2 and S8). To compare directly with our data, we ran the simulation for the same duration as our experiment and included all cell-pair relationships, giving the more complicated curve shown in Figure 4D (red curve).

The ratio of the data’s mean square variation to that of the Poisson simulation (Figure 4E, green curve) is a measure for how correlated cells remain after a given period of time has passed. Points below a value of one (Figure 4E, dashed line) represent correlated switching behavior, whereas points above it would signify anticorrelated behavior. For over 600 min, the distribution remains distinctly sub-Poissonian. Only for the longest measured times are there indications that the cells switch independently of their history, and even this is with large uncertainty. Put another way, pairs of cells often remain on approximately the same trajectory for several cell divisions, even though cell growth has diluted many of the relevant proteins to a fraction of their original level.

Stochastic Model

To examine our results at a microscopic level, we constructed a simple model that allows us to probe how the rich correlated switching dynamics arise from a simple...
regulatory network. Specifically, we asked whether the stochastic fluctuations of a single regulatory protein in our system could simultaneously explain the observed Poisson switching behavior that is expected for randomly selected individuals and subsequent long–timescale correlations. One key protein, Gal80p, functions to regulate the expression of all other genes in the network (Text S1). When it is present in the nucleus, Gal80p binds in a highly cooperative manner to the transcription factor Gal4p and represses the expression of Gal2p, Gal3p, and YFP (Volfson et al., for example, assume a Hill number of 8 between Gal4p and transcription at the GAL1 promoter [33]).

Such high levels of cooperativity frequently give rise to steep transfer functions, which can result in switch-like behavior. This means that even a small decrease in the concentration of Gal80p can cause the transcription rate of downstream genes to increase dramatically from a very small basal rate to a large maximal rate. Once the downstream protein, Gal3p, begins to be produced, it will lead to sequestering of Gal80p to the cytoplasm, completing the feedback loop and causing the cell to completely switch from the OFF to the ON state.

We constructed a simple model that captures the essential properties of this process. In our cells, Gal80p is present in very low numbers, and we therefore account for the effects of stochastic production and degradation for this protein. Protein bursting invariably increases noise levels by amplifying rare events such as changes in promoter activation or mRNA creation and destruction [18,22,41]. We assumed that the burst-size distribution was exponential in shape with a mean consistent with the results of Bar-Even et al., who found an average of 1,200 proteins per burst [30]. We further assumed that the decay rate of the protein is dominated by dilution and therefore set by the division time of the cell. Finally, we included in our model a nonzero chromophore maturation time of 20 min, as observed in our data. To account for the cooperativity between Gal80p and Gal4p, we assumed that when Gal80p levels drop below a threshold

Figure 4. Cell-Pair Behavior
The conditional switch times for closely related cells are compared. (A) The daughter switch time is compared to the mother switch times for 141 cell pairs. For times extending past 350 min (about two cell divisions), a strong correlation in times is observed. The other cell pair relationships, shown again in (B, C), are shadowed in grey. (B, C) The more distant relationships of GM-GD (n = 55) continue to show significant correlation, while the S1-S2 relationships (n = 74) shows somewhat less. The notable asymmetry of the S1-S2 distribution reflects the tendency for older siblings to sometimes switch before the younger sibling is even born. (D) In blue, the mean squared difference of the switch times from the combined relationships in (A–C), binned according to their average switch time. In red, a computer-generated Poisson simulation sets a bound for switching correlation in the limit of correlation tends to zero. The mean cell doubling time is labeled \( t_{\text{doub}} \).

(E) Dark green squares show the ratio of the two curves in (D), demonstrating the persistence of a correlation for at least hundreds of minutes after cell division. In purple, the predicted fit from our stochastic simulation after fitting to the curves in Figure 3.

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value a cell rapidly activates gene expression and enters the ON fluorescent state.

In total, the model has only three parameters: (1) mean number of Gal80p molecules present per cell, (2) the switching threshold, and (3) the Gal80p burst size estimated from literature. We estimated the first two of these parameters by fitting the model to the marginal and conditional switching distributions shown in Figure 3B. Once the theoretical switching rates were fit to the experimental data, we asked if the model explained the highly correlated switching times observed between related cells. Without any additional fitting parameters, we predicted the mother (\(t_{MD}\)) and daughter (\(t_{DMD}\)) conditional switching times (Figure 5F, brown squares) as well as their mean squared deviation \(\langle (t_1 - t_2)^2 \rangle\) (Figure 4E, purple diamonds). These predictions matched remarkably well with the experimental data (Figure 4E, green boxes; Figure 5F, gray circles). The model therefore predicts that related cells will remain highly correlated in their switching times even though switching events seem to occur in a Poisson manner. A robustness analysis (Text S3, Figure S9) suggested a narrow range of possible values with an optimum centered around (average, threshold): (2,400 proteins, 670 proteins).

Bursting events in protein production are often associated with increases of noise in protein levels [18,22]. A counterintuitive aspect of our model is that the correlation observed in cell pairs comes as a consequence of stochastic bursting. As the burst size is ratcheted up from 12 to the experimentally observed value of 1200, for example, keeping average protein level and switch rate constant, correlations begin to emerge in the cell-cell scatter plots (see Figure 5). The reason for this effect is that the periods between bursting events are dominated by dilution of proteins, a relatively low-noise process. As the burst size is increased, the time between bursts must increase commensurately, leading to long periods of correlated behavior between cells. Two cells that start with the same amount of protein will therefore dilute that protein at a similar rate and switch ON (Figure 5C, black arrows) at

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**Figure 5. Burst-Induced Correlations**

Results of our stochastic simulation.

(A–C) Fluctuations of protein concentration as a function of time are shown (gray lines). Several selected realizations are highlighted for emphasis. Proteins are created in bursts of size \(<b> = 12, 120, \text{ and } 1200\) respectively for the three panels, with the average protein level held fixed. Thresholds (solid black lines) are chosen to result in an average switching rate equal to our experimentally measured value. When protein levels drop below a threshold, that cell is considered to have switched.

(D–F) The resulting M-D scatter plots. As the burst size increases, the pattern becomes markedly more correlated. Gray circles in (F) are the experimental data reproduced from Figure 4A.
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Cells engineered in this way transition between ON and OFF states in a seemingly stochastic fashion. Cells with this genotype exhibit an extremely broad steady-state expression histogram, with fluorescence values that span more than two orders of magnitude, and the histogram has peaks on both the high and low expression limits, suggesting a bistable system with relatively infrequent transitions between the two states.

**Growth conditions.** Before imaging, cells were grown at low optical density overnight in a 30 °C shaker in synthetic dropout media with 2% raffinose as the sole carbon source. This neutral sugar is thought to neither actively repress nor induce the GAL genes [43]. We grew our cells in the absence of tetracycline, so levels of Gal80p were determined by the basal expression level of pTET02. Approximately 12 h later, cells were harvested while still in exponential phase, spun down, and resuspended in synthetic defined (SD) media. Next, cells were transferred to a chamber consisting of a thick agar pad (composed of the appropriate dropout media and 4% agarose) sandwiched between a cover glass and slide. The high agarose density constrains cells to grow largely in a two-dimensional plane.

**Microscopy.** Fluorescent and phase-contrast images of growing cells were taken at intervals of 20–35 min on 10 different days for over 100 initial progenitor cells. Image collection was performed at room temperature (22 °C) using a Nikon TE-2000E inverted microscope with an automated stage (Prior Scientific; http://www.prior.com) and a cooled back-thinned CCD camera (Micromax, Roper Scientific; http://www.roperscientific.com). Acquisition was performed with Metamorph (Universal Imaging; http://www.photomet.com).

**Supporting Information**

- **Figure S1.** Growth for Selected Cell Colony in Measurement Chamber Remains Constant for Longer than our Typical Measurement Period

- **Figure S2.** Histograms of Doubling Times as a Function of Previous Cell Divisions

- **Figure S3.** Rapid Maturation of YFP

- **Figure S4.** The Cumulative Percent of Cells That Have Switched Is Plotted against Their Marginal Switch Time

- **Figure S5.** Corrective Factors Used to Weigh Data Points According to Their Significance

- **Figure S6.** Schematic for Weighing the GM-GD Opportunity Windows and the Corresponding Window of Available Switch Times for the Example Family Tree

- **Figure S7.** Opportunity Windows for M-D, GM-GD, and S1-S2 Cell Pairs

- **Figure S8.** Overlay of Switching Patterns for all M-D, GM-GD, and S1-S2 Cell Pairs and Overlay of the Same in the Gillespie-Based Model

- **Figure S9.** Monte-Carlo Model Confidence Intervals

- **Video S1.** A Single OFF Cell Grows Over 600 min into a Small Colony of 16 Cells with Fluorescence Overlaid

- **Text S1.** Network Description

- **Text S2.** Additional Measurements

- **Text S3.** Data Analysis Methods
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Author contributions. AvO conceived and designed the experiments. BBK and QY performed the experiments. BBK, QY, and JTM analyzed the data. All authors wrote the paper.

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