A unique B2 B cell subset in the intestine

Yasuyo Shimomura,1,2,7 Atsuhiro Ogawa,1,7 Mayumi Kawada,3,7 Ken Sugimoto,1,2,7 Emiko Mizoguchi,3,4,7 Hai-Ning Shi,4,5,7 Shiv Pillai,4,6,7 Atul K. Bhan,1,2,4,7 and Atsushi Mizoguchi1,2,4,7

1 Experimental Pathology Unit, 2 Center for Systems Biology, 3 Division of Gastroenterology, 4 Center for the Study of Inflammatory Bowel Disease, Massachusetts General Hospital, Boston, MA 02114
3 Mucosal Immunology Laboratory, 4 MGH Cancer Center, Massachusetts General Hospital, Charlestown, MA 02129
7 Harvard Medical School, Boston, MA 02116

Over 80% of the body’s activated B cells are located in mucosal sites, including the intestine. The intestine contains IgM+ B cells, but these cells have not been characterized phenotypically or in terms of their developmental origins. We describe a previously unidentified and unique subset of immunoglobulin M+ B cells that present with an AA4.1−CD21−CD23− major histocompatibility complex class II bright surface phenotype and are characterized by a low frequency of somatic hypermutation and the potential ability to produce interleukin-12p70. This B cell subset resides within the normal mucosa of the large intestine and expands in response to inflammation. Some of these intestinal B cells originate from the AA4.1+ immature B2 cell pool in the steady state and are also recruited from the recirculating naive B cell pool in the context of intestinal inflammation. They develop in an antigen-independent and BAFF-dependent manner in the absence of T cell help. Expansion of these cells can be induced in the absence of the spleen and gut-associated lymphoid tissues. These results describe the existence of an alternative pathway of B cell maturation in the periphery that gives rise to a tissue-specific B cell subset.

Abbreviations used: BAFF-R, receptor of B cell activation factor from the TNF family; Btk, Bruton’s tyrosine kinase; DSS, dextran sodium sulfate; FDC, follicular dendritic cell; GALT, gut-associated lymphoid tissue; LT, lymphotoxin; MLN, mesenteric lymph node; PP, Peyer’s patch; SHM, somatic hypermutation.

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plasma cell development. Approximately half of all IgA+ plasma cells in the intestine may originate from peritoneal B1 cells, whereas the remainder may be either derived from activated B cells that migrate from organized lymphoid tissues such as PPs, or originate from naive B cells through a distinct pathway for gut-primed B cells (17–20, 23–28). In addition to plasma cells, IgM+ B cells also exist within the normal intestine and significantly increase within large intestinal mucosal tissues during inflammatory responses (29–32). However, the phenotypic and developmental properties of IgM+ B cells in the intestine have not yet been fully explored.

We describe a previously unidentified, intestine-specific, IgM+ B cell subset that is characterized by an AA4.1+CD21−CD23+MHCII bright phenotype, by a low frequency of somatic hypermutation (SHM), and by the potential ability to produce IL-12p70 after exposure to microbes. These cells appear to exist in a unique preactivated state, as indicated by the very high expression levels of MHC class II on these cells. This intestinal IgM+ B cell subset may originate from AA4.1+ immature transitional B cells in the steady state, and may be significantly increased in the inflamed intestine after recruitment from the recirculating naive B cell pool.

RESULTS

Identification of a unique AA4.1−CD21−CD23−IgM+ B cell subset in the large intestine

IgM+ B cells have been described in the large intestine, and these cells expand significantly and form B cell clusters in certain disease states involving intestinal inflammation (29–32). We herein demonstrate that the majority of IgM+ B cells in the large intestine of WT mice exhibit a unique surface phenotype that is distinct from that of recirculating splenic and mesenteric LN (MLN) B cells. In contrast to splenic, MLN, and PP B cells, the majority of intestinal IgM+ B cells are CD21−CD23−IgD low (Fig. 1 A). Because a significant increase in intestinal B cells has been well documented in several colitis models (29–32), we next examined the phenotype of IgM+ B cells from the inflamed intestine of three colitis models, including TCRαKO mice (Th2 cell–dependent chronic colitis), IL-10 KO mice (Th1 cell–dependent chronic colitis), and dextran sodium sulfate (DSS)–induced acute colitis (29, 32, 33). As previously demonstrated (29, 32), a significant increase in IgM+ B cells was observed in the inflamed intestine of TCRαKO mice and IL-10 KO mice (Fig. 1 B). Interestingly, we found that in the DSS colitis model intestinal B cell expansion was specifically induced during the recovery phase (after termination of DSS treatment), but not during the acute phase when DSS was being administered. In comparison to day 0 (normal mucosa), the proportion as well as the absolute numbers of IgM+ cells in the large intestine were slightly decreased at day 4 (acute phase), but increased significantly at day 8 (during the recovery phase) of DSS colitis (Fig. 1, B and C). Like normal intestinal IgM+ B cells, the majority of intestinal IgM+ B cells in these colitis models were also of the CD21−CD23−IgD low phenotype (Fig. 1 D). In addition, intestinal B cells from normal and inflamed mucosa were CD24 intermediate−CD62L−CD1d+ and did not express B1 cell markers (CD43, CD11b, and CD5; Fig. 1 E and Fig. S1). The intestinal B cells proliferated in vitro in the response to BCR ligation, as well as LPS stimulation (Fig. 1 F). These findings define a previously unidentified, unique B cell subset that exists in normal and inflamed large intestine.

Intestinal B cells are derived from AA4.1+ immature and transitional B cells

Intestinal B cells with their unique phenotype could either represent a naive or an activated B cell population. These cells could develop in the intestine in situ from uncommitted lymphoid progenitors, or they may be derived from newly formed B cells that not only seed the spleen but also home to the intestine, or they could represent a population that differentiates from recirculating follicular phenotype B cells or from B1 B cells. The rapid expansion of intestine-specific CD21−CD23− B cells in the DSS colitis model provided us an opportunity to more closely examine how these cells expand either by proliferation of existing cells or by enhanced recruitment or development from progenitors. Because intestinal B cells share some phenotypic features with T1 B cells, but the latter also express AA4.1, a marker for recently generated B cells, (34), we next examined AA4.1 expression on intestinal B cells. Less than 10% of intestinal B cells expressed AA4.1 (Fig. 2 A). Unlike AA4.1+ intestinal B cells, the AA4.1− intestinal B cells consisted of two subsets, T1-like CD23− and T2-like CD23+ B cells (Fig. 2 A). To examine whether AA4.1−CD21−CD23− intestinal B cells could potentially mature from the AA4.1+ immature B cells, in vivo BrdU pulse labeling was performed as previously described (35). In the AA4.1+ IgM+ population from the normal large intestine of WT mice, BrdU+ B220+ cells were observed on both day 2 and 6 after the initial BrdU injection (Fig. 2 B, top). In contrast, BrdU+ B220+ cells were not detectable at day 2 in the AA4.1+ IgM+ population, but became clearly detectable at day 6 (Fig. 2 B, bottom). Similar findings were also observed in the inflamed intestine after DSS treatment (Fig. S2). To confirm whether AA4.1+ immature B cells actually possess the ability to differentiate into intestinal IgM+ B cells, we purified AA4.1+ B220+ B cells from the spleen of GFP transgenic mice, and then transferred them (5 × 10^6) intravenously into WT mice, with or without induction of DSS colitis. GFP+ IgM+ B cells were detected in the colon and spleen of both recipient mouse groups (Fig. 2 C, right). The reconstituted GFP+ IgM+ B cells in the spleen expressed CD21 and CD23 (Fig. 2 D). In contrast, the GFP+ IgM+ B cells reconstituted in the colon did not express CD21 and CD23 (Fig. 2 D). Naïve follicular B cells, which mature from AA4.1+ immature B cells, have the ability to recirculate in search of antigens (36). Therefore, we next examined the involvement of recirculating naïve B cells in the development of intestinal B cells. To do so, we purified IgD low naïve recirculating B cells from the spleen of GFP transgenic mice and transferred them (5 × 10^6) intravenously into WT mice with or without induction of DSS colitis. 4 d after cell transfer, donor-derived
GFP<sup>+</sup> IgM<sup>+</sup> B cells were detected in the spleen of both recipient mouse groups (Fig. 2 E, left). In contrast, GFP<sup>+</sup> IgM<sup>+</sup> B cells were detectable in the inflamed colon (DSS-treated), but not in normal colon (Fig. 2 E, right). In addition, the reconstituted GFP<sup>+</sup> IgM<sup>+</sup> B cells in the spleen had a CD21<sup>+</sup> CD23<sup>+</sup> phenotype, whereas the GFP<sup>+</sup> IgM<sup>+</sup> B cells in the colon were CD21<sup>−</sup> CD23<sup>−</sup> (Fig. 2 F). These data, together with Fig. 2 (B and C) and Fig. S2, support the view that AA4.1<sup>+</sup> intestinal B cells directly or indirectly mature from AA4.1<sup>+</sup> immature B cells in the steady state and that intestinal inflammatory stimuli induce the recruitment of IgD<sup>high</sup> naive recirculating B cells to the intestine, resulting in the significant increase in IgM<sup>+</sup> B cells within the inflamed intestine.

**Intestinal B cells exhibit a unique activated phenotype**

Because some intestinal B cells originate from the recirculating naive B cell pool, particularly under inflammatory conditions, we next tested whether the intestinal B cells phenotypically resemble naive or activated B cells. The expression of several activation markers (16) was analyzed. Interestingly, the IgM<sup>+</sup>...
intestinal B cells were characterized by remarkably high expression levels of MHC class II in comparison to splenic B cells (Fig. 3 A). The expression levels of MHC class II were comparable on B cells from normal and inflamed intestine (Fig. 3 A). In addition, a slight increase in the expression of CD69 was observed on the intestinal B cells in comparison to splenic B cells (Fig. 3 A). In contrast, no significant up-regulation of Fas, PNA, CD80, or CD86 was found on the intestinal B cells. Therefore, it is possible that intestinal B cells represent a unique activated B cell population that is MHCII bright, CD69 intermediate, Fas low, PNA low, CD80 high, and CD86 low. To elucidate whether the phenotype of intestinal B cells represents a response to an intestinal pathogen, and to determine if these cells can be further activated by pathogens, we examined intestinal B cells after infection with *Citrobacter rodentium*. This infection did not induce the expression of Fas or PNA on intestinal B cells (Fig. 3 B). These data suggest that the MHCII bright intestinal IgM + B cell population is not elicited by a known intestinal pathogen, and these cells may represent a unique preactivated B cell population.

**Functional property of IgM** + B cells in the large intestine**

Because the intestine is a major source of IgA-plasma cells (17–20, 23–25), the differentiation of activated intestinal B cells into IgA plasma cells was next investigated. Interestingly, although IgM + B cells were markedly increased in the inflamed mucosa during the recovery phase of DSS colitis, no significant increase in the proportion of B220 IgA + or CD138 + plasma cells was observed at this site (Fig. 4 A). ELISPOT assay also showed that the absolute number of IgA-secreting cells among total intestinal cells decreased during the recovery phase of DSS colitis (day 8; Fig. 4, B and C). In addition, expression of activation-induced deaminase an enzyme required for class switch recombination and SHM (37, 38) was slightly lower in the IgM + B cells of the large intestine when compared with splenic IgM + B cells (Fig. 4 D). To analyze the frequency of SHM, genomic DNA was isolated from purified IgM + B cells from the large intestine and the spleen of WT mice and subjected to PCR/DNA sequence analysis using a primer set that amplifies rearranged V_{H}4-J_{H}5 light chain segments of genomic DNA (39). There was no significant restriction of V_{H}4 subfamily usages between intestinal and splenic IgM + B cells. Interestingly, the frequency of SHM within the V_{H}4 segments was significantly lower in intestinal IgM + B cells compared with splenic IgM + B cells (Fig. 4 D). These findings suggest that IgM + B cells do not appear to contribute to IgA-producing plasma cells in the large intestine, and they are characterized by the low frequency of SHM.

Several recent studies have suggested that, like T cells, B cells also possess the ability to produce several kinds of cytokines.
from the large intestine, but not the spleen, of WT mice produce a large amount of IL-12p70 in response to CpG (Fig. 4, G and H).

Antigen-independent polyclonal expansion of intestinal CD21⁻CD23⁻ B cells

The intestine is constitutively exposed to a wide spectrum of enteric microorganisms (19). Therefore, we examined the clonality of B cells in the large intestine by analyzing the VDJ region of IgM heavy chains (VHJ558 and VHQ52) to test whether some specific enteric (microbial) antigens cause the expansion of intestinal B cells. There was no restricted diversity of the BCR repertoire of VHJ558 or VHQ52 within the B cells from the normal intestine (Fig. S3 and Fig. S4). Interestingly, a similar polyclonal pattern was also observed in B cells from the inflamed intestine of DSS colitis (Fig. S3 and Fig. S4). These findings suggest that the intestinal IgM⁺ B cells using VHJ558 and VHQ52, which together account for 65% of the BCR repertoire (44), expand polyclonally.

To further test the antigen specificity of intestinal B cell development, we generated IghelMD4 Tg x RAG1 KO mice; these mice contain only a monoclonal B cell population that expresses a restricted BCR specific for hen egg lysozyme. IgM⁺ B cells were still detected in the intestines of these mice in the absence of any inflammatory stimulus, and a marked expansion of these B cells occurred in the inflamed mucosa during the recovery phase from DSS-induced colitis (Fig. 5 A). As seen in WT mice, the expanded IgM⁺ B cells formed clusters within the inflamed mucosa (Fig. 5 B). In addition, the majority of IgM⁺ B cells were of the CD21⁻CD23⁻ phenotype (Fig. 5 C). These findings suggest that antigen/BCR interactions may not be essential for intestinal CD21⁻CD23⁻ B cell development. It however cannot be ruled out that some self-antigen that cross-reacts with hen egg lysozyme exists. To further explore the role of antigens and the BCR, we investigated KO mice deficient in Bruton’s tyrosine kinase (Btk) or Lyn, both of which play a crucial role in the BCR signaling cascade (45–47). Interestingly, CD21⁻CD23⁻ IgM⁺ B cells were detectable at normal levels in the intestines of these mice in the absence of DSS (Fig. 5 D), and they were significantly increased within the inflamed intestine during the recovery phase of DSS colitis (Fig. 5 E).

T cell help is not required for intestinal CD21⁻CD23⁻ B cell development or expansion

Interestingly, expansion of intestinal B cells after an inflammatory stimulus resulted in the formation of B cell clusters with a unique pattern of T cell localization (Fig. 5 F); T cells were present around B cell clusters rather than in well-defined T cell areas, as normally seen in resident organized lymphoid tissues such as colonic patches (Fig. 5 F) (48). In addition, follicular dendritic cells (FDCs) were not detectable within the B cell clusters, whereas scattered FDCs were normally present within the follicles of organized lymphoid tissues (Fig. 5 F). Therefore, we next examined the role of T cells during intestinal B cell development by using TCRβδ

(40–43). We used real-time PCR to primarily screen B cells stimulated with ligands for pattern recognition receptors (including ligands for TLR 2, 4, 5, 7, and 9), and found that IL-12p35 mRNA expression was specifically induced in intestinal B cells after stimulation with CpG (unpublished data). Indeed, flow cytometric analysis clearly showed that IgM⁺B220⁺ B cells...
double KO (DKO) mice that have no TCRαβ or TCRγδ T cells. Just as was seen in WT mice, CD21−CD23− B cells represented the majority of IgM+ B cells in the intestine of TCRβδ DKO mice (Fig. 5 G). Interestingly, intestinal B cells were significantly increased in TCRβδ DKO mice as compared with WT mice both in the absence of inflammation and under inflammatory conditions (Fig. 5 H). Large IgM+ B cell clusters were also recognized in the inflamed intestine of TCRβδ DKO mice (Fig. 5 I). These findings suggest that T cells are not required for intestinal CD21−CD23− B cell development in a state of health and during inflammation, but may contribute to the suppression of the development or recruitment of intestinal IgM+ B cells.

Intestinal CD21−CD23− B cells develop in the absence of both the spleen and organized lymphoid tissues

The LT-αβ2 signaling cascade is required for the development of GALT (20–22). Because some intestinal IgA+ plasma cells have been shown to be derived from GALT, particularly from PP (17, 20, 23), we examined the role of GALT in CD21−CD23− B cell development by using LTα KO mice that lack GALT (20–22). Interestingly, CD21−CD23− B cells were still detectable in LTα KO mice (Fig. 6 A), and a marked expansion of these B cells was induced in the inflamed intestine after exposure to DSS (Fig. 6, B and C). IgM+ B cell clusters were still detectable in LTα KO mice (Fig. 6 D). These findings indicate that intestinal CD21−CD23− B cells develop even in the absence of GALT, which is consistent with previous reports showing that naive B cells can directly migrate into the small intestinal mucosa through a pathway different from GALT-primed B cells (24, 25).

Peripheral B cells complete their maturation through transitional stages in the spleen (1–6, 8, 34). Like recirculating follicular B cells, intestinal AA4.1−CD21−CD23− B cells also appear to mature from AA4.1+ immature B cells. Therefore, to test the role of the splenic environment during intestinal B cell development, we used splenectomized LTα KO mice in which both the spleen and GALT were absent. Like WT mice, the majority of intestinal B cells in splenectomized LTα KO mice were of the CD21−CD23− phenotype (Fig. 6 F), and a marked expansion of these B cells with cluster formation was observed during the recovery from DSS colitis (Fig. 6, C and E). These results indicate that neither the spleen nor GALT is required for the development of intestinal CD21−CD23− B cells, and that in the absence of spleen,
Figure 5. Development of intestinal CD21<sup>+</sup>CD23<sup>+</sup> B cells through BCR-independent response without T cell help. (A) Absolute number of IgM<sup>+</sup> cells in the large intestine from IgM<sup>+</sup>Btk KO mice at day 0 (normal, open bar; n = 10) and day 8 (recovery phase of DSS colitis). (B) Colonic tissues from WT mice at day 8 and from IgM<sup>+</sup>Btk KO mice at days 0 and 8 were subjected to immunohistochemical analysis for the detection of IgM<sup>+</sup> cells. Statistical significances are indicated in by ** (P < 0.001). Bars, 100 μm.

Insufficient evidence to suggest a linkage between intestinal CD21<sup>+</sup>CD23<sup>+</sup> B cells and B1 lineage cells

IgA<sup>+</sup> plasma cells in the small intestine have previously been demonstrated to originate from peritoneal B1 cells (18, 23). Therefore, we sought to determine if CD21<sup>+</sup>CD23<sup>+</sup> B cells in the large intestine might be derived from B1 B cells. To test this possibility, RAG<sup>−1</sup> KO mice were reconstituted with peritoneal CD11b<sup>+</sup> cells from WT mice, which include B1 cells and macrophages, and the mice were subsequently treated with DSS. Although CD11b<sup>+</sup> IgM<sup>+</sup> B1 cells could be recognized in the peritoneal cavity of the recipient RAG KO mice, no IgM<sup>+</sup> cells were detectable in the inflamed intestine (Fig. 7 A). We next examined IL-7 receptor α (IL-7R) KO mice in which the development of B2, but not B1, cells is impaired (49, 50) and Rac2 (guanosine triphosphatase) KO mice that show the impaired development of B1 cells (51). Only a few IgM<sup>+</sup> cells were detectable in the normal and inflamed intestine of IL-7R KO mice (Fig. 7, B and C). In addition, no B cell clusters were observed in the inflamed mucosa of IL-7R KO mice exposed to DSS (Fig. 7 D). In contrast, there were similar numbers of IgM<sup>+</sup> B cells in the normal intestine between Rac2 KO and WT mice, and significant B cell expansion was observed in the inflamed mucosa of both mouse groups after exposure to DSS (Fig. 7, B and C). These findings, together with the phenotype of intestinal B cells (CD11b<sup>+</sup>, CD5<sup>+</sup>, and CD43<sup>+</sup>) shown in Fig. 1 E, and the data from Btk KO mice, all suggest that the majority of IgM<sup>+</sup> B cells in the large intestine may originate from conventional B2 lineage cells.

Dependence of intestinal CD21<sup>+</sup>CD23<sup>+</sup> B cells on BAFF-R signaling cascade

The rapid expansion of intestinal B cells by DSS-induced intestinal inflammation provided an opportunity to closely examine the molecular events involved in intestinal B cell development and/or recruitment. We therefore performed a real-time PCR-based screening approach using purified intestinal IgM<sup>+</sup> B cells from day 0 (normal), 4 (acute colitis), and 8 (recovery phase of colitis with B cell expansion) of DSS colitis to analyze the expression of 276 molecules that have been demonstrated to play crucial roles in the genesis, differentiation, maturation, apoptosis/survival, homing and activation of B cells (1–6, 46, 47, 52, 53). We found that receptor of B cell activation factor from the TNF family (BAFF-R) is specifically up-regulated in intestinal B cells on day 8 as compared with day 4 and 0 (Fig. 8 A). In contrast, intestinal IgM<sup>+</sup> B cells did not express BCMA (B cell maturation antigen) during the course of inflammation (Fig. 8 A). BAFF-R engagement promotes processing of NF-κB2 detection of IgM<sup>+</sup> cells. Statistical significances are indicated in by ** (P < 0.001). Bars, 100 μm.
protein p100 to p52, which is dependent on NF-κB–inducing kinase (NIK) (54, 55). Recent studies indicate that activation of the BAFF-R–signaling cascade up-regulates Pim2 (a prosurvival kinase) and Bim (a BH3-only member of the Bcl-2 family) (56), and that BAFF-R signaling is negatively regulated by an adaptor protein, Act1 (57). Indeed, a significant up-regulation (56) of these mice. The data are representative of four individual recipient mice. (B–D) Intestinal inflammation was induced in WT (Day 0, n = 6), Rac2 KO (Day 0, n = 5; Day 8, n = 5) mice by oral administration of 4% DSS for 4 d. Proportion (IgM/CD3ε) at day 8 (B) and absolute number of IgMε cells in the large intestine at day 0 (normal, open bar) and day 8 (recovery, shaded bar; C) are shown. The large intestine from IL-7Rε KO mice at day 8 was subjected to immunohistochemical analysis for detection of IgMε cells (D). Presence of statistical significance (P < 0.001) compared with day 8 WT mice is indicated by **. Bars, 100 μm.

To further examine the role of the BAFF-R–signaling cascade in intestinal CD21εCD23ε B cell development, we analyzed BAFF-R–mutant (A/WySnJ) and A/J (control) mice (54–56, 58, 59). Interestingly, intestinal B cells exhibiting CD21εCD23ε phenotype were significantly reduced in BAFF-R–mutant mice as compared with control mice (Fig. 8 C). In addition, DSS–induced colitis failed to induce a significant expansion of IgMεCD21εCD23ε B cells in BAFF-R–mutant mice (Fig. 8, D and E). Indeed, B cell clusters within the inflamed mucosa were detectable in control (Fig. 8 F) but not BAFF-R–mutant mice (Fig. 8 G). In addition, significantly enhanced apoptosis of intestinal B cells was observed in BAFF-R–mutant as compared with control mice (Fig. 8 H), which is consistent with an important role of BAFF-R–signaling in B cell survival (54–59). In addition, immature AA4.1ε B cells in the large intestine were significantly reduced in BAFF-R–mutant mice as compared with control mice (Fig. 8 I). These findings suggest that the BAFF-R–associated signaling pathway plays a crucial role in intestinal B cell development and homeostasis. They are consistent with a role for BAFF-R in all B2 populations after the T1 stage and the absence of a role for BAFF-R in B1 B cells (58, 60).

**DISCUSSION**

In this study, we have described a previously unidentiﬁed, intestine–speciﬁc, IgMε B cell subset that is characterized by a unique AA4.1εCD21εCD23εMHCIImφ surface phenotype,
Figure 8. Requirement of BAFF-R–mediated signaling for intestinal CD21–/CD23– B cell development. (A) Expressions of BAFF signaling-related molecules in purified B cells from the large intestine of day 0 (open bars), 4 (gray bars), and 8 (black bars) after DSS treatment and from the spleen of day 0 (hatched bars) are shown. The results represent the means ± the SD from six individual mice in each group. *, P < 0.001 (comparison between colonic B cells at day 0 versus day 8). (B) Western blot analysis shows the expression levels of NF-κB p100 and p52 in purified B cells (>97% are IgM+) of the large intestine from 4–30 pooled WT mice at day 0 (D0, normal) and 8 (D8, recovery of DSS colitis), and of the spleen from day 0. The data are representative of two individual experiments. (C) Expressions of IgM versus CD21 or CD23 on the gated B220+ cells from normal large intestine of control and BAFF-R mutant mice are shown. (D–G) Intestinal inflammation was induced in BAFF-R mutant (A/WySnJ; Day 0, n = 5; Day 8, n = 16) and control (A/J) mice (Day 0, n = 5; Day 8, n = 10) by oral administration of 4% DSS for 4 d. (D and E) Proportion (CD3/IgM; D) and absolute numbers of IgM+ B cells (E) in the colon of control and BAFF-R mutant mice at day 8 of DSS colitis are shown. Presence of statistically significant (P < 0.001) compared with control mice at day 8 is indicated by **. (F and G) Large intestine from control mice (F) and BAFF-R mutant mice (G) at day 8 were subjected to immunohistochemical analysis for the detection of IgM+ cells. (H) Apoptosis of intestinal IgM+ B cells in control mice and BAFF-R mutant mice at day 8 of DSS colitis as judged by Annexin V staining is shown. The mean percentages of Annexin V+ cells among IgM+ cells are 36.6 ± 1.09% in control mice (n = 7) and 75.6 ± 1.64% in BAFF-R mutant mice (n = 7; P < 0.001). (I) Expression of AA4.1 versus B220 on the cells from normal intestine of control (left) and BAFF-R mutant (right) are shown. Bars, 100 μm.
low frequency of SHM, and the ability to secrete IL-12 in response to a TLR ligand. This B cell subset exists in the normal mucosa of the large intestine and expands in the inflamed mucosa of mice with acute or chronic colitis. The LTo1B2 signaling cascade is required for the development of organized lymphoid tissues (such as PP) that provide a site of origin of some IgA + plasma cells in small intestine (17, 18, 20, 23, 24). Most other small intestinal plasma cells originate from peritoneal B1 cells (17, 18, 20, 23, 24, 26–28). We herein describe an additional pathway for intestinal IgM + B cell development, in which AA4.1−CD21−CD23− B cells in the large intestine are supplied from B2 lineage cells that do not require organized lymphoid tissues for their development. Indeed, recent studies have revealed that some IgM + B cells in the small intestine are derived from the BM through a pathway distinct from that used by gut-primed B cells (24, 25). Although the phenotype of BM-derived small intestinal B cells has not been characterized in previous studies, it is possible that the AA4.1−CD21−CD23−MHCII+bright B cells in the large intestine characterized in this study may have a counterpart in the small intestine.

Recirculating B cells mature through transitional stages in the spleen, as well as the BM (2, 3, 5, 8). Our BrdU labeling and cell transfer studies suggest that intestinal AA4.1+ IgM + B cells can be derived from the AA4.1+ immature B cell pool. In addition, intestinal AA4.1+ IgM + B cells were virtually absent in BAFF-R mutant mice in which development of AA4.1+ CD23+ T2 transitional B cells is defective and T1 transitional/newly formed B cells accumulate (61–63). Therefore, it is likely that recirculating T2 B2 cells that mature from T1 cells in the spleen seed the intestinal IgM +CD21− B cell compartment in the steady state, down-regulating CD23 when they enter the intestinal milieu. Soon after they seed the intestine, they may continue to express AA4.1, which is a marker of recent maturation. In addition, CD21−CD23− B cells may accumulate in the intestine after the recruitment to the inflamed intestine of cells from the recirculating IgD+ naive B cell pool that contains both T2 cells and more mature recirculating follicular B cells. Therefore, it may be inferred that inflammatory chemokines and cytokines participate in the recruitment of recirculating B cells to the intestine during inflammation.

Peripheral B cell maturation in the spleen and BM depends in part on BCR-linked positive selection events (2, 5, 9). Interestingly, although inflammatory conditions generally induce a restriction of BCR clonality (12), our data showed that intestinal AA4.1− IgM + B cells polyclonally expand even in the presence of intestinal inflammation (Fig. S3 and Fig. 4). In addition, B cells with a restricted BCR expanded readily in normal and inflamed intestine (Fig. 3). Furthermore, T cell help was not required for the intestinal AA4.1− IgM + B cell development. Importantly, a large number of intestinal AA4.1− IgM + B cells were still detectable even in the absence of Btk or Lyn, which are major signaling molecules involved in BCR-mediated signaling. In addition, SHM that is often induced in mature B cells after encounter with specific antigens (38) was detectable at very low frequency in the intestinal IgM + B cells. These data suggest that intestinal AA4.1− IgM + B cells that are constitutively exposed to enteric microorganisms may develop or expand through a BCR-independent pathway.

Our data demonstrated that intestinal IgM + B cells express remarkably high levels of MHC class II. In contrast, several conventional markers expressed on activated splenic B cells, were not expressed by intestinal B cells. The unique expression profile of activation markers was not altered under several intestinal inflammatory conditions, including spontaneous chronic colitis, a chemically induced acute colitis, and Citrobacter rodentium infection. Therefore, it is possible that the intestinal IgM + B cell population may represent a unique preactivated B cell population presumably caused by the constitutive exposure to enteric microorganisms in the intestine. B cells have generally been believed not to produce IL-12p70, as this cytokine was detectable only in EBV-transformed B cells but not normal systemic B cells (64). However, recent accumulating studies have identified the potential ability of B cells to produce IL-12p70 under certain inflammatory and stimulatory (e.g., combination of CpG and CD40) conditions (41–43). Interestingly, intestinal IgM + B cells showed the ability to easily produce IL-12p70 in response to a microbial product CpG. Because IL-12p70 is a well known cytokine required for safeguarding against microorganisms (65), it is possible that the intestinal IgM + B cells may contribute to preserve appropriate host–microbe interaction.

In summary, we have identified an intestine-specific AA4.1−CD21−CD23−MHCII+bright B cell subset that is derived from AA4.1+ immature B cells and from recirculating naive B cells. This maturation of these cells represents a distinct pathway of B cell development that occurs in a tissue site, rather than in the BM or the spleen.

MATERIALS AND METHODS

Mice. TCRβ−DKO, RAG−KO, IL−7RKO, KO, Rac2 KO, Lyn KO, Btk KO, IL−7R KO, IghelMD4 Tg. GFP Tg mice of C57BL/6 background, BAFF−R mutant (A/WySnJ), and control (A/J) mice were obtained from The Jackson Laboratory and maintained in specific pathogen−free facilities at Massachusetts General Hospital. IghelMD4 Tg mice were crossed with RAG1 KO mice as previously described (40) to generate a mouse strain that possesses monoclonal B cells. Splenectomy and sham operations were performed on anesthetized LTα KO mice at 3 d after birth, as previously described (35). Mice were used according to the protocol approved by the Massachusetts General Hospital Animal Care and Use Committee.

DSS-induced intestinal inflammation and C. rodentium infection. Female (10–12 wk of age) mice were administered 4% DSS (MW: 36,000–50,000, ICN Biomedicals) in drinking water for 4 d, and the DSS administration was then terminated to induce recovery from acute inflammation by changing the DSS water to normal drinking water as previously described (33). These mice were killed on day 0, 4, 8, 14, and 30 after induction of DSS-induced acute intestinal inflammation. Mice were orally inoculated with C. rodentium (strain DBS100; American Type Culture Collection), as previously described (66). Bacteria were grown overnight in Luria broth and resuspended in PBS before infecting mice (0.5 ml/mouse; ~5 × 10^8 CFU of C. rodentium). Mice were killed on day 5 and 10 after infection.

Cell preparation and B cell isolation. Large intestine and parts of small intestine containing PP were incubated with 1.5 mg/ml disperse to remove epithelial cells. The remaining tissues were then incubated with 0.8 mg/ml
collagenase for 20 min. After passing them through a glass wool column, the mucosal cells were further enriched by Percoll gradient (40/72%) centrifugation. MLNs and spleen were dispersed using 27-gauge needles, and red blood cells were removed using ACK solution. For purifying B cells, cells were incubated with PE-conjugated anti-Ig (Invitrogen) at 4°C for 30 min, and then incubated with the anti-PE microbeads at 4°C for 30 min. After washing, the labeled cells were isolated using the MACS system (Miltenyi Biotec).

**Cell transfer.** Immature AA4.1+ B220+B cells or recirculating naive IgD+ B cells were purified from the spleens of GFP transgenic mice (5–20 mice). After enrichment of B cells using the MACS system, B cell populations were purified using flow cytometry and sorting. 5 × 10^6 purified B cells were intravenously transferred into WT mice with or without DSS treatment. Peritoneal CD11b+ cells were purified from pooled WT mice (day 5) using a magnetic cell separation (MACS) system (Miltenyi Biotec). 5 × 10^6 purified cells were transferred intraperitoneally into recipient RAG1 KO mice that subsequently received DSS orally.

**Phenotypic analysis.** Cells were isolated from the large intestine (excluding the cecum), MLN, spleen, PP, and BM as previously described (33). The mAbs used were FITC-, PE-, PerCP-, APC-, or biotin (followed by incubation with PerCP-streptavidin)-conjugated anti-CD1 (B2), -CD3e (145-2C11), -CD5 (53–7.3), -CD9 (KMC8), -CD11b (M1/70), -CD21 (7G6), -CD23 (B3B4), -CD24 (M1/69), -CD43 (S7), -CD47 (H1.2F3), -B220 (RA3-6B2), -CD62L (53–7.3), -CD9 (KMC8), -CD11b (M1/70), -CD21 (7G6), -CD23 (B3B4), -CD24 (M1/69), -CD43 (S7), -CD47 (H1.2F3), -B220 (RA3-6B2), -CD62L (hen egg lysozyme-14), -CD138 (282–2), -IgD (11-26C2a), -Fas (Jo2; BD Biosciences), -IgA (SouthernBiotech), -IgG1 (SouthernBiotech), -IgD, -FDC.

**Intracellular cytokine staining.** Spleen and large intestinal cells were stimulated either with or without 1 μM CpG (InvivoGen) for 17 h. GolgiStop (BD Biosciences) was added during the last 7 h. The cells were first stained with FITC-conjugated anti-B220 (RA3-6B2) and APC-conjugated anti-IgM antibody, and then they were fixed and permeabilized with Cytofix/Cytoperm solution. The cells were then stained intracellularly with PE-conjugated anti-IgM+ cells were performed as previously described (35). Immunohistochemical staining was performed using the avidin–biotin complex method, as previously described (67). The mAbs used were anti-CD3e, -IgM, -IgD, and -FDC.

**Ig secretion and proliferation of B cells.** Cells were subjected to ELISPOT assay for the detection of IgA-secreting cells as previously described (32). For proliferation assay, B cells were enriched by negative sorting for depletion of CD3+, Gr1-, DX5-, and CD11c-expressing cells, as previously described (40). The enriched B cells (5 × 10^6) were incubated with 50 μg/ml F(ab′)2 goat anti-IgM (Jackson ImmunoResearch Laboratories) or LPS (5 μg/ml, InvivoGen) and cultured for 48 h, as described previously (34). After pulsing with 1 μCi of [3H] thymidine for 18 h, uptake was assessed by scintillation counting.

**Analysis of SHM.** SHM analysis was performed as previously described (39). Genomic DNA was prepared from purified IgM+ B cells using a DNasey Tissue kit (Qiagen) and amplified using a V4 light chain primer set (V4w-68F, 5′-GATTITTCAGCTCTCCGTAATGATGCC-3′ and Jwseq-3R, 5′-TGATAATGAGCCCTCTCCTCAT-3′) and Platinum Taq DNA polymerase High Fidelity (Invitrogen). Amplified products were cloned into pcRC2.1-TOPO vector (Invitrogen) and subjected to DNA sequence analysis.

**Quantitative real-time PCR.** After RNA purification from the purified B cells using QiA shredder and RNeasy Mini kit (Qiagen), the first strand of the cDNA was synthesized by SuperScript (Invitrogen) according to the manufacturer’s instruction. Real-time PCR was performed as previously described (33). cDNA was initially amplified by using SYBR green PCR core reagent kit (Stratagene) with a primer set specific for β-actin. The real-time PCR reaction was performed by MX3000p QPCR machine (Stratagene), and the cDNA concentration was normalized at the CT value of β-actin. The normalized cDNA samples were subsequently subjected to real-time PCR with Rhox dye as a reference dye for the measurement of expression levels of specific molecules.

**BCR repertoire.** For DNA sequence of CDJ R3 within IgM heavy chain, cDNA from purified B cells were amplified using VHJ558 or VπQ52 and Cμ primer set, as previously described (68). The amplified products were cloned using TOPO-TA cloning system. Positive clones that were randomly picked up were subjected to DNA sequence.

**Detection of NFκB.** After B cells were positively purified from spleen and colon of pooled mice (4–30 mice) under MACS system, purified IgM+ cell (2 × 10^6) B cells were immediately lysed with RIPA buffer. 50 μg of protein was subjected to immunoblot using rabbit polyclonal anti-p52 (k-27; Santa Cruz Biotechnology, Inc.) and anti–rabbit HRP (Thermo Fisher Scientific), as previously described (55).

**Online supplemental material.** Fig. S1 shows the expression of B1 cell markers on colonic IgM+ B cells. Fig. S2 shows the BIR3-containing incorporated AA4.1+ and AA4.1+ IgM+ cells under inflammatory condition. Fig. S3 shows BCR repertoire (VHJ558) of colonic B cells in steady state and under inflammatory condition. Fig. S4 shows the BCR repertoire (VHQS2) of colonic B cells in steady state and under inflammatory condition. The online version of this article is available at http://www.jem.org/cgi/content/full/jem.20071572/DC1.

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