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Moderate caloric restriction initiated in rodents during adulthood sustains function of the female reproductive axis into advanced chronological age

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Summary

Age-related ovarian failure in women heralds the transition into postmenopausal life, which is characterized by a loss of fertility and increased risk for cardiovascular disease, osteoporosis and cognitive dysfunction. Unfortunately, there are no options available for delaying loss of ovarian function with age in humans. Rodent studies have shown that caloric restriction (CR) can extend female fertile lifespan; however, much of this work initiated CR at weaning, which causes stunted adolescent growth and a delayed onset of sexual maturation. Herein we tested in mice if CR initiated in adulthood could delay reproductive aging. After 4 months of CR, the ovarian follicle reserve was doubled compared to ad libitum (AL)-fed age-matched controls, which in mating trials exhibited a loss of fertility by 15.5 months of age. In CR females returned to AL feeding at 15.5 months of age, approximately one-half remained fertile for 6 additional months and one-third continued to deliver offspring through 23 months of age. Notably, fecundity of CR-then-AL-fed females and postnatal offspring survival rates were dramatically improved compared with aging AL-fed controls. For example, between 10 and 23 months of age, only 22% of the 54 offspring delivered by AL-fed females survived. In contrast, over 73% of the 94 pups delivered by 15.5- to 23-month-old CR-then-AL-fed mice survived without any overt complications. These data indicate that in mice adult-onset CR maintains function of the female reproductive axis into advanced age and dramatically improves postnatal survival of offspring delivered by aged females.

Key words: aging; caloric restriction; fecundity; fertility; menopause; oocyte; ovary; reproduction.

Introduction

Compared to other organ systems, the female reproductive axis ages exceptionally early in life, culminating in a cessation of normal ovarian function that in humans heralds the onset of menopause at around 50 years of age (Richardson et al., 1987; Faddy et al., 1992). A progressive decline and ultimately exhaustion of the ovarian oocyte-containing follicle reserve is the prime determinant of the age of onset of menopause (Richardson et al., 1987; Faddy et al., 1992; Tilly, 2001; Charleston et al., 2007; Hansen et al., 2008), a period in life associated with a loss of fertility and the emergence of a diverse spectrum of health issues resulting from an absence of cyclic ovarian function (Buckler, 2005). Furthermore, concomitant with reduced follicle numbers, the quality of the remaining oocytes in females generally declines with advancing age, leading to extremely poor success rates of natural and assisted fertility attempts (Navot et al., 1991; van Kooij et al., 1996). This latter outcome is an increasingly problematic issue as more and more women, especially in Western societies, are postponing childbearing to older ages (Ventura, 1989; Ventura et al., 2004; Hamilton et al., 2007).

While laboratory rodent models do not exhibit menses and, thus, do not undergo a true menopause like humans, female mice do exhibit an age-related decline in their ovarian follicle reserve, leading to a state of natural infertility approximately halfway through their chronological lifespan (Gosden et al., 1983; Perez et al., 1999; Wu et al., 2005). Furthermore, as a consequence of the loss of cyclic ovarian function, aging female mice exhibit many of the same adverse physiological changes observed in postmenopausal women, and have thus been extensively utilized as model for understanding the impact of ovarian failure on the aging female body. For example, past studies have shown that transplantation of young adult mouse ovaries into aging female mice delays the decline in reproductive potential and increases overall life expectancy of the recipient (Cargill et al., 2003). More recently, it was reported that minimizing follicle loss and prolonging ovarian function in female mice through targeted inactivation of the pro-apoptotic Bax gene (Perez et al., 1999) extends fertile lifespan and minimizes many age-related health problems, including bone and muscle loss, excess fat deposition, alopecia, cataracts, deafness, increased anxiety and selective attention deficit (Perez et al., 2007). Despite the promising nature of these findings in mice, strategies for delaying ovarian aging or the timing of menopause in women do not currently exist.

Dietary caloric restriction (CR) without malnutrition is known to extend lifespan and minimize age-related dysfunction of many organs, including those of the female reproductive axis,
in several animal models (Masoro, 2005). In fact, a beneficial effect of CR on reproductive function in aging rodents has been documented since the early 1900s. In one of the first published studies on this topic, Osborne et al. (1917) reported that restricting food intake in young female rats resulted in sustained fertility much later in life. From this, the authors concluded that ‘the menopause has been postponed [by food restriction] long beyond the age at which it usually appears’. Ball et al. (1947) showed in mice that restricting caloric intake from weaning until 8 months of age and then allowing the females to ad libitum (AL) feed resulted in 13 times more litters over the next 4 months when compared with age-matched continuously AL-fed controls. Litter size as well as regularity of estrous cyclicity next 4 months when compared with age-matched continuously AL-fed controls. Litter size as well as regularity of estrous cyclicity next 4 months when compared with age-matched continuously AL-fed controls. Litter size as well as regularity of estrous cyclicity next 4 months when compared with age-matched continuously AL-fed controls. Litter size as well as regularity of estrous cyclicity next 4 months when compared with age-matched continuously AL-fed controls. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity next 4 months when compared with age-matched continuously AL-fed controls. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity. Litter size as well as regularity of estrous cyclicity.

Even though several historical studies have reported a beneficial effect of CR on reproductive performance in rodents, there is a large degree of variation in how these experiments were designed (e.g. the age at which CR was initiated and the duration of CR, as well as the severity of the CR relative to total caloric intake in AL-fed controls) and in what outcomes different investigators assessed over the years. Of particular importance, much of the past work that evaluated the effects of CR on long-term fertility in females was derived from studies of CR initiated at weaning, which complicates interpretation of the outcomes due to a retardation of postweaning growth to adulthood and to a delay in the onset of sexual maturation (Merry & Holehan, 1979; Holehan & Merry, 1985; Gonzales et al., 2004). Herein, we investigated in female mice the effects of moderate CR initiated in adulthood on reproductive performance with age and offspring survival.

Results

Effectiveness of the adult-onset CR protocol

Since past studies in mice have shown that one outcome of CR is an increased ovarian follicle reserve later in life (Nelson et al., 1985), we first tested the effectiveness of the adult-onset CR protocol by assessing changes in oocyte numbers. Morphometric analysis of healthy (non-atretic) follicle numbers in serially sectioned ovaries of 8-month-old mice indicated that almost twice as many quiescent primordial follicles were present in ovaries of CR females (516 ± 32 per ovary; mean ± SEM, n = 5 mice) when compared with ovaries of age-matched continuously AL-fed controls (273 ± 69 per ovary; mean ± SEM, n = 6 mice) (Fig. 1). This effect of adult-onset CR appeared to be independent of changes in primordial follicle growth activation rates or early follicle maturation, as indicated by the presence of comparable numbers of small growing follicles and no change in the ratio of resting (primordial) to total immature follicles between experimental groups (Fig. 1). However, the number of atretic immature follicles was reduced in ovaries of adult-onset CR females (468 ± 158 per ovary; mean ± SEM, n = 5 mice) when compared to age-matched continuously AL-fed controls (960 ± 112 per ovary; mean ± SEM, n = 6 mice).

The effectiveness of the adult-onset CR protocol was also evaluated by weekly monitoring of body weight, which showed that during restriction CR females weighed approximately 25% less than age-matched continuously AL-fed controls (Fig. 2). Notably, the body weight difference seen in CR females was rapidly reversed after a resumption of AL feeding at 15.5 months of age (Fig. 2).

Timing of natural reproductive failure in continuously AL-fed females

To determine a reference age bracket for the natural decline in reproductive potential of continuously AL-fed females, and thus establish a time point for the re-initiation of AL feeding in the CR cohort, fertility of continuously AL-fed females was monitored in natural mating trials. Between 10 and 12.5 months of age, 7 of 11 continuously AL-fed females became pregnant and delivered offspring (Fig. 3A). Over the following 4.5 months, however, fertility plummeted (Fig. 3B–D), with only a single pregnancy observed in the AL-fed female cohort between 15.5 months of age (Fig. 3C, D).

Fig. 1 Impact of adult-onset caloric restriction (CR) on the ovarian follicle reserve. Number of non-atretic and atretic immature follicles per ovary in continuously ad libitum (AL)-fed or CR female mice at 8 months of age, after initiation of CR 4 months earlier. These data are the mean ± SEM of combined results from analysis of 5 (CR) or 6 (AL-fed) mice, with significant differences in mean values indicated.
and 17 months of age (Fig. 3D). Furthermore, offspring survival rates also markedly declined with advancing maternal age of the AL-fed controls (Fig. 3). It should be mentioned here that fertility of CR females while on the restriction protocol (i.e. between 10 and 15.5 months of age) was poor, with a total of 9 pregnancies achieved by the 11 CR females (7 females pregnant once, 1 female pregnant twice). Furthermore, all offspring were delivered dead or died very shortly after birth (data not shown). These outcomes in CR females during the period of food restriction were fully anticipated, however, based on results of both prior studies (Visscher et al., 1952) as well as on the logical need for increased energy demands on the pregnant dams not being met under the CR protocol.

**Adult-onset CR prolongs function of the reproductive axis in aging females**

Re-initiation of AL feeding of CR females at 15.5 months of age, which corresponds to a time when natural fertility in the continuously AL-fed controls was lost (Fig. 3), resulted in a rapid return of fertile potential in subsequent mating trials. Specifically, between 15.5 and 17 months of age, 50% of the CR-then-AL-fed females achieved a pregnancy with delivery of offspring (Fig. 4A), and many CR-then-AL-fed females retained reproductive function into more advanced ages (Fig. 4A–E). Litter size as well as offspring survival rates were also remarkably high for CR-then-AL-fed mothers at advanced ages. For example, one 20- to 21.5-month-old CR-then-AL-fed female delivered 7 offspring, of which 6 survived and developed without any apparent complications (female #5, Fig. 4D). Two of the CR-then-AL-fed females actually remained fertile past 23 months of age; however, the 5 total offspring delivered by these females did not survive for more than a few hours after birth (data not shown).

**Fertility, fecundity and offspring survival are all improved by adult-onset CR**

The data presented in Figs 3 and 4 depict the outcomes for each mouse, so that the reproductive performance of individual

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**Fig. 2** Effect of adult-onset caloric restriction (CR) on body weight. Weekly weight measurements (mean ± SEM) of continuously ad libitum (AL)-fed and CR/CR-then-AL-fed female mice between 10 and 23 months of age. Note the return of body weight in CR mice to continuously AL-fed control levels within 2 weeks after resumption of AL-feeding at 15.5 months of age (CR-then-AL-fed), highlighted by grey shading.

**Fig. 3** Natural decline in fertility of continuously ad libitum (AL)-fed control females with age. (A–D) Fertility of continuously AL-fed females between 10 and 12.5 (A), 12.5 and 14 (B), 14 and 15.5 (C), and 15.5 and 17 (D) months (M) of age (each mouse is represented by a number on the x-axis). The total number of offspring delivered and that survived for each female that became pregnant are indicated. Crosses designate mice that had to be euthanized due to severe health complications or that died of natural causes during the study period.
females over the course of the mating trial period could be visualized. However, after compiling the individual mouse data, the effects of adult-onset CR on female reproductive performance with age were even more apparent. For example, between 15.5 and 17 months of age, 50% of the CR-then-AL-fed females remained fertile whereas only 12.5% of the continuously AL-fed control females achieved a pregnancy over the same time period (Fig. 5A). Furthermore, 50% of 17- to 18.5-month-old, 40% of 18.5- to 21.5-month-old, and 30% of 21.5- to 23-month-old CR-then-AL-fed females continued to achieve pregnancies with birth of offspring. At these ages, all of the continuously AL-fed females were completely infertile (Fig. 5A).

In evaluating these outcomes from a different perspective, the total number of offspring born to continuously AL-fed females steadily declined between 10 and 17 months of age (Fig. 5B). Strikingly, during an age bracket when only 2 offspring were delivered by a single continuously AL-fed control female (15.5–17 months of age), age-matched CR-then-AL-fed females delivered a total of 25 offspring (Fig. 5B). Similarly, the fecundity (number of offspring per litter) of continuously AL-fed females declined throughout the study period (Fig. 5C) and was consistently below that of age-matched CR-then-AL-fed females. In fact, after re-initiation of AL feeding, the fecundity of previously CR females immediately improved, such that 15.5- to 17-month-old and 17- to 18.5-month-old CR-then-AL-fed females delivered an average of 5.2 ± 2.3 and 6.0 ± 0.3 offspring per litter, respectively (mean ± SEM, n = 5 mice per group) (Fig. 5C). Of note, this latter outcome is close to the average litter size of AL-fed C57BL/6 female mice during their peak reproductive period (6.2 pups/litter; see http://www.informatics.jax.org/external/ festing/mouse/docs/C57BL.shtml). During the entire mating trial period (10–23 months of age), 9 of the 11 continuously AL-fed females became pregnant and delivered offspring at least once, with a total of 18 pregnancies and 54 offspring delivered. By comparison, between 10 and 23 months of age, 8 of the 11 CR (10–15.5 months)/CR-then-AL-fed (15.5–23 months) females became pregnant and delivered offspring at least once, with a total of 30 pregnancies and 133 offspring delivered. Finally, offspring survival rates were also dramatically improved by adult-onset CR. Specifically, of the 94 total pups delivered by 15.5- to 23-month-old CR-then-AL-fed females, 69 (73.4%) survived and developed normally without any apparent
abnormalities (Fig. 5D). In contrast, only 12 of the 54 total pups (22.2%) born to 10- to 17-month-old continuously AL-fed females survived (Fig. 5D; note that no pregnancies were observed in this group past 17 months of age).

In addition, some of the female offspring born to 15.5- to 21.5-month-old CR-then-AL fed females were maintained for up to 12 weeks of age (n = 18), and all exhibited normal growth into adulthood with no apparent abnormalities as assessed by gross anatomical examination after euthanasia (data not shown). Normal maturation to adulthood was further indicated by the fertile potential of female offspring (n = 3) born to 20- to 21.5-month-old CR-then-AL-fed female mice. After a single mating attempt at 10 weeks of age, these females delivered normal size litters (a total of 19 offspring from the 3 females) with no pup mortality observed (data not shown).

Discussion

Socioeconomic trends within the past few decades have resulted in more women in developed countries electing to postpone childbirth to later reproductive years. In the USA for example, since the 1970s the rate of first births for women between the ages of 30 and 34 years has increased by 375%. The rate of first births for women older than 35 is even greater, rising within the last four and a half decades by 500% or more for women aged 35–39 and 40–44 years (Ventura, 1989; Ventura et al., 2004; Hamilton et al., 2007). Perhaps even more striking is the fact that the National Center for Health Statistics (NCHS) has no records of live births for women between 45 and 49 years of age in the 1970s (Ventura, 1989), whereas approximately 7000 live births were documented by the NCHS for mothers within this age range in 2006 (Hamilton et al., 2007). Despite this continuously increasing trend to have a first child later in life, when fertility for most women is suboptimal and successful pregnancies may be difficult to achieve even with the aid of modern assisted reproductive technologies, no strategies currently exist to delay the age-related decline in fertility that most women experience in their late 30s and early 40s.

In rodent models, the ability of reduced food intake to postpone age-related infertility in females has been documented for nearly a century (Osborne et al., 1917; Ball et al., 1947; Vischer et al., 1952; Berg, 1960). Although the collective results of these studies are promising, it is important to emphasize that in these earlier studies the CR protocol was often severe and was initiated at weaning. Thus, as normal postnatal growth and development were compromised, the prolongation of female fertile lifespan resulting from CR was attributed to a stunting of overall growth and to a delayed onset of normal sexual maturation (Ball et al., 1947).
effectively maintain overall well-being and increase lifespan of adult-onset CR protocol that has recently been shown to easily occur in adult female rodents (Johnson et al., 1997; Ball et al., 1947), our studies of aging female mice subjected to an adult-onset CR protocol argue against the idea that CR extends female fertile lifespan by simply stunting adolescent growth and delaying the age at which sexual maturation is reached. In fact, the outcomes obtained herein following the initiation of CR as late as 2 months after attainment of sexual maturity were just as dramatic, if not even more so, as those reported earlier for CR initiated at weaning. For example, over an 8-month period at ages when reproductive capacity had already ceased entirely in continuously AL-fed control females (15.5–23 months of age), many adult-onset CR-then-AL-fed females achieved pregnancies and delivered several litters (up to 5 pregnancies/female) at a rate that was comparable to the fertility observed for continuously AL-fed C57BL/6 female mice during prime reproductive life (for additional details, refer to http://www.informatics.jax.org/external/festing/mouse/docs/C57BL.shtml). It is currently unclear, however, why some of the CR females remained infertile after the resumption of AL feeding at 15.5 months of age, but this may simply reflect individual variation in response to the CR protocol. Nevertheless, these results indicate that a delay of female reproductive senescence can be achieved using a procedure in adults that does not cause a delay in the timing of sexual maturation (i.e. CR initiated at weaning: Ball et al., 1947; Holehan & Merry, 1985; Gonzales et al., 2004) or require the use of genetic manipulation (e.g. Bax gene knockout: Perez et al., 1999, 2007).

Another observation made herein is that the ability of adult-onset CR to sustain function of the female reproductive axis with age appears to be mediated, at least in part, via maintenance of the ovarian follicle reserve. Although a systematic assessment of follicle dynamics throughout the entire study period (4–23 months of age) was not performed, the increased number of primordial follicles detected in aging females 4 months after the initiation of CR may reflect a decrease in follicle loss via atresia, reduced rates of primordial follicle growth activation or increased primordial follicle renewal, the latter of which has recently been shown to occur in adult female rodents (Johnson et al., 2004, 2005; Kerr et al., 2006; Bukovsky et al., 2007; Lee et al., 2007; Tilly & Rueda, 2008). Irrespective of how the follicle reserve is maintained by adult-onset CR, the quality of the oocytes remaining in the ovaries of the CR females does not appear to be compromised by advanced maternal age. In fact, our data show that both litter size and postnatal offspring survival rates were remarkably high for 15.5- to 23-month-old CR-then-AL-fed females, especially when compared with the same outcomes in considerably younger (10- to 15.5-month-old) continuously AL-fed controls. These findings offer compelling evidence that age-related deterioration of egg quality, which is widely believed to be a principal driving force behind poor pregnancy success rates in aging females (Navot et al., 1991), can be overcome by a dietary modification initiated in adult life. This conclusion aligns well with recent data from studies of domestic farm animals, which indicate that oocyte and embryo quality can be improved by a short-term reduction in feed intake (Lozano et al., 2003; Freret et al., 2006).

In summary, we have shown in mice that moderate CR initiated in adult life can dramatically extend function of the female reproductive axis into advanced age, with significant benefits noted not just in the females placed under CR but also in the survival rates of the offspring conceived by aged CR females once returned to an AL diet. Although these data suggest that there may be ways to safely extend reproductive function in aging females without negatively affecting adolescent growth or the timing of sexual maturation, it remains unknown if aging humans and nonhuman primates would respond similarly. Nevertheless, we are encouraged by an increasing body of evidence indicating that CR produces many of the same physiological, metabolic and hormonal adaptations in aging nonhuman primates (Roth et al., 2000, 2004; Bodkin et al., 2003; Ingram et al., 2006) and humans (Walford et al., 2002) that are observed in rodents. Moreover, in light of recent data from mice demonstrating the beneficial effects of prolonged ovarian function on the female body with age (Perez et al., 2007), sustaining the ovarian follicle pool by adult-onset CR may, irrespective of the fertility issues discussed herein, play a significant role in improving the overall quality of life of aging females normally compromised by endocrine changes associated with the loss of cyclic ovarian function. Thus, even if direct application of adult-onset CR protocols in humans is not feasible for the purpose of sustaining function of the female reproductive axis with age for fertility or health reasons, ongoing efforts to develop clinically relevant small molecule CR mimetics (Ingram et al., 2004, 2007; Baur & Sinclair, 2006; Austad, 2007; Chen & Guarente, 2007) may be of considerable interest to this important aspect of the biology of aging.

Experimental procedures

Animals

Virgin C57BL/6 female mice were obtained from the National Institute on Aging (NIA, Bethesda, MD, USA). Wild-type C57BL/6 adult male mice were obtained from Jackson Laboratories (Bar Harbor, ME, USA). All animal husbandry and experimental procedures were reviewed and approved by the institutional animal care and use committee of Massachusetts General Hospital.

Feeding regimen

For these experiments, we used an adult-onset CR protocol developed by the NIA for their Biomarkers of Aging study (Turturro et al., 1999), in which CR is initiated at 14 weeks of age at 10%
restriction, increased to 25% restriction at 15 weeks, and to 40% restriction at 16 weeks where it is maintained until the resumption (if so desired) of AL feeding at some point later in life (see http://www.nia.nih.gov/ResearchInformation/ScientificResources/AgedRodentColoniesHandbook/CaloricRestrictedColony.htm). Each female was housed individually in a conventional (non-ventilated) cage. The CR females were fed once daily between 06:00 and 09:00 hours with 2.7 g (4.33 Kcal g–1) of NIH-31/ventilated) cage. The CR females were fed once daily between weekly to assure proper effectiveness of the feeding regimen.

Mice were weighed access to NIH31 standard open-formula food. Water was provided CR protocol was continued until the females reached 15.5 months of age, at which time the previously CR females were allowed AL access to NIH31 standard open-formula food. Water was provided AL for all animals during the study period. Mice were weighed weekly to assure proper effectiveness of the feeding regimen.

Fertility testing
Mating trials were initiated at 10 months of age for all mice. For mating attempts during CR, a male mouse of proven fertility was housed overnight in a cage with a female and removed the following morning, so that the female could be fed her dietary food ration. The AL and CR females were handled identically. Upon return of CR females to AL feeding at 15.5 months of age, male mice were left with the females until obvious signs of pregnancy were observed, and then removed until the next mating attempt. Males were randomly rotated among the cages during the mating trials. The total number of offspring delivered per litter and the number of offspring delivered that were viable and survived were recorded separately for each pregnancy. Offspring that did not survive were either found dead at birth or died very shortly after delivery. All viable offspring were allowed to remain with the dam until weaning (day 21 post-partum), at which time the offspring were removed from the cages to allow for a subsequent mating attempt with the dam. For all offspring that survived, no overt anatomical or health complications were observed (data not shown).

Ovarian follicle counts
Oocyte-containing follicles were scored by histomorphometric evaluation as originally described (Jones & Krohn, 1961), with slight modifications detailed previously (Tilly, 2003; Skaznik-Wikel et al., 2007). Each ovary was given a numerical code so that all counts were conducted without knowledge of sample identity. Slides were then decoded and the total number of healthy and atretic immature follicles per ovary was calculated. Accuracy and reproducibility of the counts were independently confirmed by having a different observer reassess follicle numbers in randomly selected slides without knowledge of sample identity.

Data presentation and analysis
Graphs depict results from each individual mouse or combined data obtained from independently replicated experiments (mean ± SEM). Where appropriate, the combined data were analyzed by a one-way analysis of variance followed by two-tailed t-test for statistical comparisons between mean values.

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