Effect of Environmental Tobacco Smoke on Levels of Urinary Hormone Markers

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We investigated the effect of ETS exposure on the levels of urinary reproductive hormones among healthy newly married nonsmoking women in China who intended to conceive and had stopped contraception.
study and by the institutional review board of the Harvard School of Public Health.

Detailed description of field data collection can be found elsewhere (Wang et al. 2003). Briefly, the eligibility criteria for the field enrollment were as follows: a) full-time employed women workers, b) newly married, c) 20–34 years of age, and d) had obtained permission to have a child. All the women were nulliparous. Women were excluded if a) they were already pregnant before enrollment, b) they had tried unsuccessfully to get pregnant for at least 1 year in the past, and c) they planned to quit/change jobs or to move out of the city over the 1-year course of follow-up. After obtaining informed consent, the interviewer administered a baseline questionnaire, which included information on contraceptive use, reproductive history, sociodemographic characteristics, alcohol use, and environmental and occupational exposures. Beginning from the date of stopping use of contraceptive methods, each woman kept a daily diary to record sexual intercourse, vaginal bleeding, medication, and medical conditions and collected a daily first-morning void urine specimen for hormone assay. Daily diary information and urine specimens were collected for up to 12 months or until a pregnancy was clinically confirmed.

The study was conducted in Anqing Textile Mill during 1997 to 2000. The women were recruited at the local Maternal and Child Health Care Center. Of the total 1,006 newly married women, 35 women were ineligible, 10 women refused, and 961 women were enrolled; 99% of them did not smoke.

Hormone assays were performed for 387 women who had provided sufficient diary and urine samples and who did not smoke. A total of 574 women were excluded from the current analysis for the following reasons: 95 women continued to use contraceptives; 121 women declined diary or urine collection; 78 women became pregnant because of contraceptive failure; 53 did not begin recording diaries and collecting daily urine samples immediately after stopping contraception; 8 were lost to follow-up; 7 had menstrual irregularity at baseline; the others did not have adequate diary and urine samples.

Laboratory assays of urinary PdG, E$_1$C, and hCG. Urine specimens were stored in our field central laboratory at ~20°C. Urinary PdG and E$_1$C levels were measured by enzyme-based immunoassays (Munro et al. 1991). This method was very sensitive and stable. The minimum detection levels for PdG and E$_1$C were 3 ng/mL and 0.096 ng/mL, respectively, and the coefficients of variation measured from the repeated standards were 4.3 and 5.1% respectively. Urinary human chorionic gonadotropin (hCG) levels were analyzed by the immunoradiometric assay (O’Connor et al. 1988). Urine creatinine levels were measured according to the method of Jaffe (Husdan and Rapoport 1968). All PdG, E$_1$C, and hCG values were normalized to creatinine values to adjust for urine concentration. All the urine specimens from each woman were analyzed and tested during a single run and were assayed in duplicates. Discrepancies of more than 3-fold between duplicate assays were presumed to result from technical error, and the assay was repeated. The geometric mean of the replicates has been used to summarize the results for each sample.

Statistical analysis. The central focus of our analysis was to examine the independent association between ETS exposure and urinary PdG and E$_1$C profiles among eligible menstrual cycles. This required characterization of following key variables: a) ETS exposure status; b) ovulation status and the day of ovulation in each cycle; c) conception status in each ovulatory cycle; and d) modeling ETS exposure in relation to menstrual-day-specific urinary PdG and E$_1$C levels.

ETS exposure status. As part of the daily diary, information on daily exposure to ETS was obtained. Two specific questions were asked: a) Was there anyone who smoked around you at home yesterday? b) Was there anyone who smoked around you at your workplace yesterday? If the woman answered “yes” to either of the two questions, we coded the day as having ETS exposure. For a specific cycle, we counted the number of days in the cycle and calculated the percentage of days with ETS exposure. If it was greater than zero, we coded the cycle as having ETS exposure.

Day of ovulation. We used two independent methods to determine the day of ovulation. We first used a previously published E$_1$C:PdG ratio algorithm (Baird et al. 1995). Briefly, this algorithm scans 5-day sequences and looks for a 5-day sequence in which the ratio value for the first day is the highest of the five, and the ratio value for each of the last 2 days are ≤ 40% of the first-day value. The second day in this sequence was designated the day of ovulation or called day of luteal transmission (DLT) for that cycle. This algorithm had several limitations: a) The cut-point of a 60% decrease in E$_1$C:PdG ratios in the preceding 2 days was derived from previously published data and was subject to the sensitivity of laboratory methods for detecting PdG and E$_1$C. b) If there were missing data around ovulation, the algorithm may fail to detect the DLT; c) multiple DLTs may be detected in one cycle.

To cross-validate the day of ovulation identified by the above method, we also applied a two-piecewise regression model for daily PdG levels to identify the day when PdG started to rise (PdG rising point). This method is based on the fact that PdG remains at a lower level during follicular phase and rises after ovulation. For each cycle, we assumed that log(PdG) values follow a normal distribution with constant mean and variance before ovulation. After ovulation, we modeled the log(PdG) with a normal distribution with a quadratic mean function and constant variance. For the day j of cycle i, we modeled the log(PdG) value as:

$$\log (PdG_{ij}) = \beta_0 + \beta_1 (j-\delta) + \beta_2 (j-\delta)^2 \quad \text{if } j \leq \delta$$

$$\log (PdG_{ij}) = \beta_3 + \beta_4 (j-\delta) + \beta_5 (j-\delta)^2 \quad \text{if } j > \delta$$

where day $\delta + 1$ was assigned as the day of ovulation. A “best fit” (maximum $R^2$) algorithm was applied to identify the turning point $\delta$. We compared the ovulation day identified by each method. In the subsequent analyses, we only included cycles in which the ovulation days identified by the different methods were within ±3 days. To be consistent, we used the ovulation day derived from the PdG rising point in the subsequent analysis.

Figure 1 illustrates the sampling frame and steps we took to determine the day of ovulation. Of the total 1,484 cycles from the 387 women, 804 cycles were selected for urinary hormone analysis. We oversampled the cycles with early pregnancy loss when we selected cycles. Sixteen cycles were found without ovulation by algorithms, and their anovulatory status was verified by graphic examination. After determining the day of ovulation by two different methods, we excluded those cycles for which ovulation could not be determined by one of the methods, or in which comparison of the two methods yielded a difference in ovulation day greater than 3 days. As a result, a total of 673 cycles from 371 women were included in the subsequent analysis. We compared the cycles that were included and excluded and did not find any significant difference in terms of subjects’ demographic characteristics and ETS exposure status (data not shown).

Conception status. A Bayesian model was applied to determine conception status based on daily urinary hCG values. Details can be found elsewhere (Wang et al. 2003). Briefly, urine samples from 37 control women who were not at risk for conception were collected for serving nonconception controls cycles, and urine samples from cycles ending with clinical pregnancy were used for conception control cycles. We assumed that square root of hCG values followed a normal distribution with a constant mean and variance before conception. For postconception, we modeled the square root of hCG with a normal distribution with a quadratic mean function and constant variance, with the mean and variance differing for clinical pregnancy and early pregnancy loss. We chose noninformative proper prior distributions for each parameter.
and fitted the model using Markov Chain Monte Carlo methods. This model allowed us to calculate a probability of conception for each observed cycle. We defined conception as a probability ≥ 0.9.

**ETS exposure and urinary hormone profiles.** Once the ovulation day was identified, we aligned the cycles according to the day of ovulation. The average length of follicular phase was 15.8 days with a range of 6 to 39 days and luteal phase was 15.0 days with a range of 6 to 33 days. We focused on a 20-day window starting from 9 days before ovulation to 10 days after ovulation for comparison of hormone profiles for the following reasons. Most cycles had at least 20 days in length. This ensures comparability of day-specific hormones among cycles, and each cycle has relatively equal contribution to the model parameter estimation. Because the distribution of values for urinary PdG and E\textsubscript{1}C were strongly skewed toward the upper end, we transformed daily urinary PdG and E\textsubscript{1}C to the log scale for analysis. We calculated each day mean log(PdG) and log(E\textsubscript{1}C) for ETS exposure and nonexposure cycles. By inverse transformation, we plotted the daily mean PdG and mean E\textsubscript{1}C by ETS exposure status. We further used linear regression models to examine the associations between ETS exposure and daily log(PdG) and log(E\textsubscript{1}C) levels within the defined window. Because E\textsubscript{1}C and PdG levels fluctuate during preovulatory and postovulatory phase, we adjusted the days relative to ovulation using indicator variables. The basic model was:

\[
Y_{ij} = \beta_0 + \beta_1 \times \text{ETSi} + \beta_2 \times (\text{day} = -9) + \beta_3 \times (\text{day} = -8) + \ldots + \beta_{20} \times (\text{day} = 9)
\]

\[Y_{ij} = \log(\text{PdG}) \text{ or log}(\text{E}1C)\text{ of day } j \text{ in cycle } i\]

If a day had missing hormone data, that day was not included in the analysis. Assuming that the missing data were randomly distributed within the window, the model should be valid. The model was also adjusted for potentially important covariates, including age (linear and quadratic terms), body mass index (BMI; linear and quadratic terms), education (high/middle), shift work (yes/no), stress (low, moderate, high), noise exposure (low, middle, high), and dust exposure (low, middle, high). Because there were multiple daily PdG and E\textsubscript{1}C observations from each woman, we applied a generalized estimate equation (GEE) to adjust for intrawoman correlation with SAS procedure GENMOD (SAS Institute, Cary, NC, USA) assuming an exchangeable working correlation structure. We also explored the interaction terms of ETS exposure status and the day indicator variables and found no interactions.

**Results**

This is a young, nulliparous cohort. All the women were newly married and were attempting to conceive. Of the 371 women included, none smoke or drink alcohol. They all were full-time textile workers. Table 1 presented the characteristics of women who were included in the analyses stratified by ETS exposure status during the follow-up cycles. Women without ETS exposure were similar to those with ETS exposure in terms of age, height, weight, BMI, education, and occupational exposures.
The major occupational exposures in these women were shift work, dust, and noise.

This analysis included 371 women who contributed a total of 1,444 cycles. The average number of cycles followed was 3.8 (range, 1–16). Of the 1,444 cycles, 474 (32.8%) cycles had conception. Of the 474 conceptions, 338 conceptions reached clinical recognized pregnancy; 146 (30.8%) ended with early pregnancy losses. The median time to clinical pregnancy was three cycles.

Of the 673 cycles included in this study, 344 were nonconception cycles and 329 were conception cycles; 76 cycles did not have ETS exposure and 597 cycles had ETS exposure. Figure 2 illustrates daily mean E1C and PdG levels over the 20-day window of the menstrual cycles, stratified by conception versus nonconception cycles. Among the nonconception cycles, ETS-exposed women had a consistently lower daily urinary E1C level compared with nonexposed women. Table 2 presents the crude and adjusted associations between ETS exposure and urinary PdG and E1C stratified by conception and nonconception cycles. ETS exposure was found to be associated with a lower urinary E1C level (β = –0.43, SE = 0.08, p < 0.001 in log scale) among nonconception cycles. Among conception cycles, the association was not significant (β = –0.17, SE = 0.10, p = 0.085 in log scale). There was no significant difference in PdG level between nonexposed and exposed women regardless of conception status.

Figure 3 plots the cycle mean urinary PdG and E1C levels by the quintile of the percentage of days having ETS exposure for the cycle. The first quintile had higher mean E1C than did other quintiles among nonconception cycles, but it did not show a clear dose–response relationship.

Discussion

The potential adverse reproductive health effects of ETS exposure are of great public health concern even when the reproductive effects of exposure to ETS arc of modest magnitude, because ETS exposure is so common and widespread. We recently reported a dose–response relationship between ETS exposure and early fetal loss in the same study cohort (Venners et al. 2004). In this article we reported a significant effect of ETS exposure on urinary E1C levels during nonconception cycles but not during conception cycles. The differential effect of ETS on urinary E1C levels between conception and nonconception cycles is interesting. One possible explanation is that the E1C level was much higher during conception cycles than during nonconception cycles and we did not have enough power to detect the relatively small effect of ETS exposure among conception cycles. This is one of the few prospective studies examining the effect of ETS exposure on urinary hormone markers, and it provides new insight about potential biologic mechanism by which ETS affects reproductive outcomes and stimulates further research in this area. Further studies are needed to understand the interactions between individual susceptibility and ETS exposure on reproductive hormones.

Studying ETS raises unique challenges, including exposure assessment and confounding by active smoking (Seidman and Mashiach...
Several limitations need to be taken into account when interpreting the results of this study. All the women subjects were shift workers in a textile industry and were young and-nulliparous. Therefore, caution is needed before generalizing our findings to other populations. We did not have biochemical markers of ETS exposures, and we relied only on women’s self-reporting of both active and passive cigarette smoking. The sample size of non-ETS group was relatively small. The ovulation days were estimated from urinary PdG and E1C levels, and we included only the cycles in which the ovulation days identified by two methods were within ± 3 days, and this may be still subject to errors. To address this issue, we conducted sensitivity analysis by changing the inclusion criterion of the cycles in which the ovulation days identified by two methods were the same, within ± 1 day, within ± 2 days, within ± 4 or more days, and repeated the analysis respectively. The results did not change significantly (data not shown).

Furthermore, ETS exposure at different days of the menstrual cycle may have different effects, and we did not have adequate sample size to examine the potential timing effect of ETS exposure.

Our study also has the following strengths: The accurate determination of day of ovulation is critical for appropriate comparisons of hormone levels among individuals, given that hormone levels change significantly over a menstrual cycle. In this study, the day of ovulation for each menstrual cycle was cross-validated by two methods. This allowed us to align the individual menstrual cycles according to ovulation day. This was a prospective study, and the ETS exposure status was based on daily diary recording during the menstrual cycle, which eliminated potential recall biases commonly encountered in retrospective studies. In many previous studies, it was difficult to tease out the effect of active smoking versus ETS exposure when the study population contained a high proportion of women who were exposed to both active and passive smoking. In this study, all the women were nonsmokers. In addition, these women were homogeneous in terms of sociodemographic characteristics and occupation; thus, this study is less likely confounded by other environmental exposures.

The observed association between ETS exposure and urinary E1C level is biologically plausible. Cigarette smoking has been suggested to have antiestrogenic effects (Baron et al. 1990; Spangler 1999). Evidence showed that smoking reduces the risk of endometrial cancer, increases the risk of osteoporosis, and decreases the age of menopause. Our finding is also consistent with a number of studies indicating the effects of active smoking on estrogen levels. MacMahon et al. (1982) reported lower urinary levels of estrone, estradiol, and estril during the luteal phase of the cycle among the premenopausal smokers compared with non-smokers and ex-smokers and suggested that smoking might reduce luteal estrogen production. Subsequently, Mochizuki et al. (1984) reported significantly lower estrogen levels in pregnant smokers compared with pregnant nonsmokers. Basu et al. (1992) reported that smoking and oral contraceptives independently lower serum estradiol and progesterone concentrations in premenopausal women. Westhoff et al. (1996) further reported that cigarette smoking was associated with decreased midcycle and luteal-phase estradiol levels.

Other studies of smoking and reproductive hormones yielded negative results. Friedman et al. (1987), in a study of 9 postmenopausal smokers and 16 nonsmokers, found that estrone, estradiol, dihydrotestosterone, and dehydroepiandrosterone sulfate (DHEA-S) did not differ between the two groups. Key et al. (1991) further compared serum concentrations of estradiol, progesterone, and DHEA-S, and urinary excretion rates of six steroids of predominantly adrenal origin, in a large cohort of healthy premenopausal and postmenopausal female smokers and nonsmokers, and found that cigarette smoking does not affect serum estradiol. Thomas et al. (1993) from a study of 25 premenopausal cigarette smokers and 21 nonsmokers also reported no significant difference in urinary concentrations of estradiol, estrone, or estriol.

The possible explanations for these discrepancies include inadequate sample size for the negative studies, population differences, and, most important, lack of control of the timing of ovulation relative to the hormone sampling. As is well known and also seen from our data, reproductive hormones fluctuate over a menstrual cycle, and the timing of the specimen collection is likely to be a critical factor in making comparisons among groups or studies. It is conceivable that without controlling for ovulation, it would be difficult to make valid comparison in hormone levels between exposed and unexposed individuals. The selection of serum versus urinary hormone markers and laboratory methods may also account for some differences. In addition, reproductive hormones are affected by both physiologic and pathologic states, including a woman’s age, BMI, pre-versus postmenopause, ovulation versus nonovulation cycle, conception versus nonconception cycle, use of oral contraceptives, or other exogenous sex hormones.

In summary, this prospective study indicated that ETS exposure in demographically homogeneous nonsmoking women was associated with significantly decreased urinary E1C levels throughout the nonconception menstrual cycles, suggesting that the adverse reproductive effect of ETS may act in part through its antiestrogen effects.

References


