



Age-Dependent Maturation of Toll-Like Receptor-Mediated Cytokine Responses in Gambian Infants

Citation

Burl, Sarah, John Townend, Jainaba Njie-Jobe, Momodou Cox, Uche J. Adetifa, Ebrima Touray, Victoria J. Philbin, et al. 2011. Age-dependent maturation of toll-like receptor-mediated cytokine responses in Gambian infants. PLoS ONE 6(4): e18185.

Published Version

doi:10.1371/journal.pone.0018185

Permanent link

<http://nrs.harvard.edu/urn-3:HUL.InstRepos:5266853>

Terms of Use

This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at <http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA>

Share Your Story

The Harvard community has made this article openly available.
Please share how this access benefits you. [Submit a story](#).

[Accessibility](#)

Age-Dependent Maturation of Toll-Like Receptor-Mediated Cytokine Responses in Gambian Infants

Sarah Burl^{1,2*}, John Townend³, Jainaba Njie-Jobe¹, Momodou Cox¹, Uche J. Adetifa¹, Ebrima Touray¹, Victoria J. Philbin⁴, Christy Mancuso⁴, Beate Kampmann^{1,2}, Hilton Whittle¹, Assan Jaye¹, Katie L. Flanagan^{1,9}, Ofer Levy^{4,9}

1 Infant Immunology, Medical Research Council (UK) The Gambia, Fajara, The Gambia, **2** Department of Paediatrics, Imperial College London, Paddington, London, United Kingdom, **3** Statistics Department, Medical Research Council (UK) The Gambia, Fajara, The Gambia, **4** Division of Infectious Diseases, Department of Medicine, Children's Hospital Boston and Harvard Medical School, Boston, Massachusetts, United States of America

Abstract

The global burden of neonatal and infant mortality due to infection is staggering, particularly in resource-poor settings. Early childhood vaccination is one of the major interventions that can reduce this burden, but there are specific limitations to inducing effective immunity in early life, including impaired neonatal leukocyte production of Th1-polarizing cytokines to many stimuli. Characterizing the ontogeny of Toll-like receptor (TLR)-mediated innate immune responses in infants may shed light on susceptibility to infection in this vulnerable age group, and provide insights into TLR agonists as candidate adjuvants for improved neonatal vaccines. As little is known about the leukocyte responses of infants in resource-poor settings, we characterized production of Th1-, Th2-, and anti-inflammatory- cytokines in response to agonists of TLRs 1-9 in whole blood from 120 Gambian infants ranging from newborns (cord blood) to 12 months of age. Most of the TLR agonists induced TNF α , IL-1 β , IL-6, and IL-10 in cord blood. The greatest TNF α responses were observed for TLR4, -5, and -8 agonists, the highest being the thiazoloquinoline CLO75 (TLR7/8) that also uniquely induced cord blood IFN γ production. For most agonists, TLR-mediated TNF α and IFN γ responses increased from birth to 1 month of age. TLR8 agonists also induced the greatest production of the Th1-polarizing cytokines TNF α and IFN γ throughout the first year of life, although the relative responses to the single TLR8 agonist and the combined TLR7/8 agonist changed with age. In contrast, IL-1 β , IL-6, and IL-10 responses to most agonists were robust at birth and remained stable through 12 months of age. These observations provide fresh insights into the ontogeny of innate immunity in African children, and may inform development of age-specific adjuvanted vaccine formulations important for global health.

Citation: Burl S, Townend J, Njie-Jobe J, Cox M, Adetifa UJ, et al. (2011) Age-Dependent Maturation of Toll-Like Receptor-Mediated Cytokine Responses in Gambian Infants. PLoS ONE 6(4): e18185. doi:10.1371/journal.pone.0018185

Editor: Nick Gay, University of Cambridge, United Kingdom

Received: November 25, 2010; **Accepted:** February 22, 2011; **Published:** April 13, 2011

Copyright: © 2011 Burl et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by the Medical Research Council (MRC, UK, www.mrc.ac.uk), The Gambia and by National Institutes of Health (NIH) RO1 AI067353-01A1 and Bill & Melinda Gates Foundation Grand Challenge Explorations and Global Health awards (to OL). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The laboratory of OL receives sponsored research support from VentiRx Pharmaceuticals, which manufactures certain TLR8 agonists although distinct from those studied in this manuscript. This does not alter the authors' adherence to all the PLoS ONE policies on sharing data and materials. There are no other known competing interests.

* E-mail: s.burl@imperial.ac.uk

⁹ These authors contributed equally to this work.

Introduction

The greatest burden of morbidity and mortality from infectious diseases occurs in children under 5 years of age, with the highest rates occurring in resource-poor countries. This urgent global issue is the target of United Nations Millennium Goal 4 to reduce under-five child mortality by two-thirds by 2015 [1]. One of the most effective measures to prevent infection is vaccination early in life, in particular at birth as it is the most reliable point of healthcare contact [2,3]. However, young children often do not respond to vaccines as efficiently as adults as a result of distinct features of their immune systems [4–8].

Among the distinct features of neonatal immune system is a diminished ability of monocytes and antigen-presenting cells (APCs) to generate Th1-polarizing signals in response to most stimuli, including reduced production of TNF α and interferon gamma (IFN γ) important for host defence against intracellular

pathogens and for generation of adaptive immune responses [9–11]. However, the impairment is stimulus-specific and some stimuli are able to effectively activate neonatal APCs [7,10,12]. In this context, characterizing the ontogeny of human immune responses including the contribution of innate immunity, provides insight into susceptibility of newborns and infants to infection, assesses immune polarization that may affect risks of allergy and atopy [13], and may inform development of age-specific adjuvanted vaccines [7].

Modifying the innate response may increase the strength of adaptive responses required for inducing efficient immunological memory in infants. Until recently, the only approved adjuvant component in vaccines was alum. However, the discovery that microbial products activate host cells via pattern recognition receptors such as Toll-like receptors (TLRs) and enhance adaptive immune responses by triggering cytokine production and dendritic cell maturation, has led to TLR agonists being developed as

vaccine adjuvants [14–18]. The TLR4 agonist monophosphoryl lipid A (MPL) has been combined with alum (AS04) in hepatitis B and human papilloma vaccines [19–21]. Agonists of several other TLRs, including TLR3, 7, 8 and 9, are in clinical development as vaccine adjuvants targeting mycobacterial, parasitic and diseases [22] [7,23–27]. Of note, several existing vaccines trigger TLRs: *Bacillus Calmette Guerin* (BCG), the most widely used vaccine against TB with an established safety and efficacy profile activates TLR2, -4 and -8 [28–30] and the meningococcal outer membrane protein complex used to adjuvant the Haemophilus conjugate vaccine, was subsequently shown to be a TLR2 agonist [31]. These examples suggest that in certain contexts TLR agonists can be safe and effective as vaccine adjuvants and highlight the importance of characterizing ontogeny of TLR-mediated responses in target populations, including newborns and infants.

Age-dependent changes in TLR function have been demonstrated in humans by natural deficiencies in TLR signaling molecules including a deficiency in IRAK-4 (a kinase involved in the TLR signaling cascade) [32,33] and MyD88 (a TLR adaptor molecule) [11] that can result in life-threatening infections in infancy and childhood. Of note, infections decrease in IRAK-4-deficient children >8 years of age, suggesting a greater dependency early in life on TLR signaling for protection against infection. Although newborns demonstrate similar basal expression of monocyte TLRs at birth [34], neonatal blood mononuclear cell responses to TLR agonists are distinct from those of adults, with impairment of Th1 cytokine responses (e.g., TNF α , IFN γ) but similar or greater production of cytokines with Th2 (IL-6, IL-1 β), Th17 (IL-23) and anti-inflammatory (IL-10) activity [4,34–42]. By contrast, cord blood responses to TLR8 agonists, including imidazoquinoline compounds and single stranded RNAs (ssRNAs), induced robust adult-like TNF α responses raising the possibility that TLR8 agonists may serve as effective vaccine adjuvants for newborns and young infants [34,43].

Because infant peripheral blood is more difficult to obtain than cord blood, relatively less is known about TLR function during infancy. The few infant studies available have been conducted in resource-rich settings and indicate that there is a marked increase in TLR-mediated Th1 cytokine production within the first 6 months of life [13,37,44,45]. To our knowledge, there are no published studies of TLR function in newborns and infants from resource-poor countries and none from the African continent. Such data may be of considerable importance as these populations are at high risk of infection, and may demonstrate distinct innate response profiles compared to other populations with lesser burdens of disease. For example, infants from resource-poor countries often respond quite differently to immune stimuli including to BCG vaccination [46,47]. In addition to providing insight into age-dependent development of TLR-mediated innate immunity, characterizing the ontogeny of TLR responses in an African country may also inform development of adjuvanted vaccines for high-risk newborns and infants. We therefore characterised the production of Th1, Th2, and anti-inflammatory cytokines in response to agonists of TLRs 1–9 in whole blood cultures derived from Gambian infants during the first year of life. We found that cord blood Th1-polarising cytokine responses were generally impaired, that TLR8 agonists gave the strongest Th1-cytokine responses at birth and that by 1 month of age the pro-inflammatory responses to most TLR agonists were increased, with predominance of responses to TLR4, -5 and -8 agonists that changed across the first 12 months of life.

Materials and Methods

Study Design

This cross-sectional study was approved by the Joint Gambia Government/Medical Research Council Ethics Committee and the London School of Hygiene and Tropical Medicine Ethics Committee. Between June and November 2009, 120 Gambian infants, up to 12 months of age, were recruited at the Sukuta Health Centre (a peri-urban (area surrounding an urban town with characteristics of a rural setting) population 30 minutes from the coast of The Gambia). Infants were recruited into 8 age groups: birth and 1-, 2-, 3-, 4-, 6-, 9- or 12- months of age (or +2 weeks of age stated; n = 15 per group). For every study subject, written informed consent of a parent/guardian was obtained. Children were excluded if they had any signs of intercurrent infection. Neonates were also excluded if they had a low birth weight (< 2.5 kg) or were a twin. Each child within an age group had received comparable vaccines according to the Gambian Extended Programme of Immunisation (EPI) schedule, and had not received any vaccine within the previous 7 days. Infants were also excluded if their weight was outside the target range for age stated on the Infant Welfare Card. Ten millilitres of umbilical cord blood (collected before delivery of the placenta) and 3 mL of infant venous blood was collected into tubes containing heparin (sodium salt from porcine intestinal mucosa, Sigma-Aldrich, Poole, UK) at 7.5 U/mL blood and transferred to the laboratories within 6 hrs of collection. Blood was acquired prior to administration of any EPI vaccine that may have been indicated on the day of recruitment.

Cell culture conditions

From each blood sample, 100 μ L of undiluted whole blood (per condition) was cultured overnight (18–24 hours) without stimulation (negative control), with phorbol 12-myristate 13-acetate (PMA; 0.1 μ g/mL, Sigma)/ionomycin (calcium salt, from *Streptomyces globatus*, 1 μ g/mL, Sigma) (positive control) and with each of the 9 TLR agonists (Table S1). TLR agonists were purchased from InvivoGen (San Diego, CA, US) and included: Pam3CSK4 (synthetic tripalmitoylated lipopeptide that mimics bacterial lipoproteins; TLR1/2, 1 μ g/mL), Poly (I:C) (a synthetic analog of double-stranded RNA (dsRNA) associated with viral infection; TLR3, 100 μ g/mL), LPS (*Escherichia coli* K12 Lipopolysaccharide; TLR4, 1 μ g/mL), flagellin (from *Salmonella typhimurium*; TLR5, 10 μ g/mL), FSL-1 (synthetic lipoprotein of *Mycoplasma salivarium*; TLR2/6, 10 μ g/mL), ssRNA40 (20-mer phosphothioate protected single-stranded RNA oligonucleotide; TLR8, 10 μ g/mL), CL075 (thiazoloquinolone derivative; TLR8, 10 μ g/mL), GardiquimodTM (an imidazoquinoline amine analogue to guanosine; TLR7, 10 μ g/mL), ODN M362 (synthetic type C unmethylated-CpG dinucleotide-containing oligonucleotide; TLR9, 1 μ M) (Table S1). TLR concentrations were selected based on those that induced maximal cytokine responses from previous studies [34,43] and from preliminary studies in 9 months old Gambian infants (unpublished data). After overnight culture, 100 μ L of ice-cold serum-free RPMI media was added to the wells and centrifuged at 1,500 rpm for 10 mins. 120 μ L of supernatant was collected and stored at –20°C for subsequent analysis. For RNA studies, 300 μ L of whole blood were cultured in each of four conditions (unstimulated, ssRNA, GardiquimodTM or CLO75) for 4 hours then added to 860 μ L PaxGene lysing reagent (Pre-AnalytiX, Becton Dickinson, France) prior to storage at –70°C ahead of RNA extraction.

Cytokine measurement

Supernatants were thawed and centrifuged at 1500 rpm for 5 mins to pellet any precipitation in the sample. Concentrations of TNF α , IL-1 β , IL-6, IFN γ and IL-10 (Th1 cytokine kit, Bio-Rad, Hercules, California, US) were measured for all samples. The bead array assays were conducted according to the manufacturer's instructions (Bioplex Reagent kit, Bio-Rad) using 50 μ L of serial (1:4) dilutions of the standard in RPMI medium containing 10% human serum and 50 μ L of samples. Standard curve outliers were eliminated by identifying samples where the coefficient of variance (CV) was greater than 10% and observed/expected \times 100 (obs/exp^{*100}) was outside the range of 100 \pm 20. Cytokine concentrations below the level of detection (out of range (OOR) <) were assigned a value of half the lowest value recorded in that assay. Similarly, all samples with values above the level of detection (OOR >) were assigned a value of twice the largest value recorded in that assay.

RNA extraction

Samples were thawed and left at room temperature (15 – 25°C) >4 hours for complete lysis prior to centrifugation for 10 mins at 5000 rpm. RNA was extracted according to manufacturer's instructions with modification for the small blood volumes (PaxGene Blood RNA kit, QIAGEN, Germany). Briefly, the pellet was resuspended in 500 μ L of RNase-free water and centrifuged again at 5,000 rpm for 10 mins. The pellet was then resuspended in 360 μ L of BR1 buffer (PaxGene Blood RNA kit). The samples were further processed according to the manufacturer's instructions and eluted in a final volume of 80 μ L of Tris/EDTA (TE)-based elution buffer (BR5). RNA was then purified further using the RNeasy Mini Elute Clean Up kit (QIAGEN, Germany) and resuspended in 12 μ L of BR5. The RNA yield was measured on the Nanodrop (Thermo Scientific, Wilmington, Delaware, US) and purity was based on absorption values at 280 nm (protein) and 230 nm (detection of contaminating organics/proteins) using the following criteria: 260 nm/280 nm >1.8 and 260 nm/230 nm >1.8.

SA Bioscience Microarrays

0.1 μ g total RNA was reverse transcribed to cDNA using RT² First Strand Kit (C-03) (SA Biosciences, Frederick, MD, US) according to the manufacturer's instructions and diluted with RNase-free water into a final volume of 111 μ L. A customized 6-gene real time PCR array (SABiosciences, Frederick, MD) was employed, containing primers specific for TNF, IL-6, IL-10, IL12A, IL12B and IFNG and using the RT-SYBR Green/ROX qPCR Master Mix according to the manufacturer's instructions (SABiosciences, Frederick, MD) using approximately 1 ng of original RNA per gene. mRNA expression was calculated as a copy number (Ct) value and normalized using multiple housekeeping genes (B2M, RPL13A, GAPDH, Δ C_t). Fold change (Δ Δ C_t) in gene expression was used to display the data and calculated between the unstimulated control cells in comparison to TLR agonist-stimulated cells applying the following equation: if X >1, X, -1/X, if X = Δ C_t therefore all values <0 represented down-regulated genes and all values >0 represented up-regulated genes. Δ C_t values were used for all statistical analysis.

Statistical analysis

Net cytokine responses were calculated by subtracting the cytokine concentrations in the medium control wells from those in supernatants derived from samples stimulated with TLR agonists. For each TLR agonist/cytokine combination the resulting datasets of net responses (all ages combined) were dichotomised into \geq

median value, or < median value. Due to a number of responses above the level of detection, comparisons of responses at difference ages were made by comparing the proportions of values above and below the median using Fisher's exact test. Tests for a trend with age were carried out using logistic regression with above or below the median as the response variable and age as the independent variable. Comparisons between the response to medium and the gross responses to each of the TLR agonists in cord blood were made using Sign tests which are robust to the influence of out of range values, in accordance with the paired nature of the data. Analyses that compared two time points or two ages were done using a non-parametric Mann U Whitney test and correlations were calculated on log-transformed data using a Pearson correlation. Data were analysed using Stata version 11.0 (StataCorp, Texas, US) and GraphPad Prism version 5.01 (GraphPad Software Inc., US). Real time PCR analysis used software from SA Biosciences as stated above. Box and whisker plots demonstrate inter-quartile ranges indicated by boxes, median values by horizontal bars, and 10 and 90 percentiles indicated by whiskers. P-values <0.05 were taken to indicate statistical significance.

Results

Reactivity to TLR4, 5 and 8 agonists predominate at birth

The positive control PMA/ionomycin induced cytokine responses that were largely independent of age (TNF α p = 0.603, IL-6 p = 0.758, IL-1 β p = 0.257, IFN γ p = 0.167). The lone exception was IL-10, whose production diminished with increasing age (p = 0.005; data not shown).

Compared to the unstimulated control, there was a significant induction of TNF α , IL-6, IL-1 β and IL-10 production in cord blood to all the TLR agonists studied with the exception of FSL-1 (TLR2/6) that did not induce a significant TNF α response (Table 1).

Of note, the efficacy with which TLR agonists induced these cytokines, differed both between agonists and with age. Agonists of TLR4, -5, and, in particular TLR8 (including the combined TLR7/8 agonist) induced the highest levels of most cytokines, with agonists of TLR1/2, -3, -2/6, -7 and -9 eliciting a reduced cytokine response (Table 1 and Figure 1A). The combined TLR7/8 agonist (the thiazoloquinolone CL075) induced the highest levels of TNF α compared to agonists of TLR4 (p = 0.0202) and TLR5 (<0.0001). Moreover CL075 (TLR 7/8) was the only TLR agonist that induced IFN γ in cord blood compared to the unstimulated control (p = 0.002, Table 1 and Figure 1B). In agreement with previous studies [43], the TLR8 agonists (ssRNA and CLO75) induced significantly more cytokine production in cord blood cultures compared to the TLR7 agonist (GardiquimodTM) (Figure 2).

TLR-mediated IL-6 and IL-1 β production is high from birth, while TNF α and IFN γ production peak at 1 month

Each cytokine studied demonstrated a distinct age-dependent profile in response to each TLR agonist during the first year of life. An assessment of effect with age for each cytokine response to each agonist revealed that TNF α responses to FSL-1 (TLR2/6; p <0.001), LPS (TLR4; p = 0.009), Flagellin (TLR5; p = 0.001), GardiquimodTM (TLR7; p <0.001), ssRNA (TLR8; p <0.001), and CL075 (TLR7/8; p <0.001) all demonstrated age-dependent effects (Table S2A). Graphic representation of these data suggested this effect was due to a predominant increase in TNF α production between cord blood and 1 month of age in response to most agonists (Figure 3). This impression was confirmed by repeating

Table 1. TLR agonists induce cytokine production in cord blood of Gambian newborns.

	Unstim. Control	Pam. (TLR1/2)	PolyI:C (TLR3)	LPS (TLR4)	Flag. (TLR5)	FSL-1 (TLR2/6)	Guard. (TLR7)	ssRNA (TLR8)	CL075 (TLR7/8)	ODN (TLR9)
TNFα	1.309	3.98*	6.34*	1,117.17***	133.52***	4.29	12.13*	5,003.68***	5,180.04***	10.74**
IL-6	245.9	7,161.75***	1,831.22**	22,529.47***	25,209.28***	8,811.02***	5,390.73***	14,412.48**	21,262.00***	3,802.29**
IL-1β	3.11	36.42**	15.31**	1,293.61***	1,522.90***	30.17**	59.92**	3,069.16***	2,557.96***	36.92**
IL-10	10.36	151.20***	35.05**	966.90***	1,145.50***	145.06***	156.36***	745.06***	1,647.61***	33.06**
IFNγ	3.335	3.34	2.83	6.37	6.38	3.24	3.34	23.36	59.17**	3.34

100 μ l cord whole blood was cultured overnight with each of the TLR agonists and cytokine production (pg/mL) in supernatants was measured as described in the Materials and Methods. Concentrations are absolute values in pg/mL. Comparisons between unstimulated and stimulated values, n = 12–15; *p = 0.05 – 0.019, **p = 0.01 – 0.001, ***p < 0.001.

doi:10.1371/journal.pone.0018185.t001

the analysis between ages 1 to 12 months (excluding the cord blood) (Table S2B) and comparing the individual responses between birth and 1 month of age (Figure 4). After the initial increase in TNF α responses by 1 month, TNF α responses to most agonists were stable from 1 month to 12 months of age (Table S2B and Figure 3).

Responses to the TLR8 agonist (ssRNA) and the combined TLR7/8 agonist (CLO75) were notable exceptions to this pattern. The initial increase in reactivity to CLO75 from birth to 1 month of age (p = 0.005), was then followed by a decline from 1 to 12 months of age (trend analysis p < 0.001, from a median at 1 month of 13,852 pg/mL, to a median at 12 months of 2,771 pg/mL), though nevertheless remained higher than those for TLR2, -3, -2/6 and -9 (Table S2 and Figure 3). In contrast, TNF α responses to ssRNA (TLR8) did not reveal a significant trend from 1 to 12 months of age (Figure 3). The relatively low TNF α response to Pam3CSK4 (TLR1/2), Poly (I:C) (TLR3) and ODN M362 (TLR9) at birth failed to increase in any age group to 12 months, in fact the TLR9-mediated TNF α response significantly decreased from the 1 month to 12 month age groups (trend analysis p = 0.008, Table S2 and Figure 3).

While TLR-mediated IFN γ production at birth was very limited (Figure 1 and Table 1), most TLR agonists elicited greater IFN γ production by 1 month of age: Pam3CSK4 (TLR1/2; p = 0.003 (Figure 5A), Poly I:C (TLR3; p < 0.001), LPS (TLR4; p < 0.001), Flagellin (TLR5; p < 0.001), FSL-1 (TLR6/2; p = 0.002), GardiquimodTM (TLR7; p < 0.001; Figure 5B), ssRNA (TLR8; p = 0.017; Figure 5C), CL075 (TLR7/8; p < 0.001; Figure 5D), ODN M362 (TLR9; p = 0.066; Figure 4 and Table S2). TLR-mediated IFN γ production then remained stable from 1 to 12 months of age, with the exception of responses to the TLR1/2 agonist that decreased with age (trend analysis; p = 0.003, Figure 5A).

All the TLR agonists studied induced high levels of IL-6 at birth (Table 1) and generally TLR-mediated IL-6 production remained high for the first year of life (Table S2). Similarly, IL-1 β and IL-10 production was high at birth in response to TLR4, -5, and -8 agonists and remained high for the first year of life with the exception of the combined TLR7/8 agonist (CLO75) that induced less IL-1 β with age (Table S2). In accordance with prior reports of an early life bias towards a high ratio of IL-6/TNF α production [40,41], in our cohort, ratios of TLR-mediated IL-6/TLR-mediated TNF α were initially high at birth (cord) and then decreased with age, reflecting a relatively constant and robust IL-6 production and an increase in TLR-mediated TNF α by 1 month of age (data not shown).

TLR8 agonists elicit distinct early age-dependent Th1 responses

In accordance with a prior cord blood study [43], TLR8 agonists induced greater Th1-polarising cytokine responses in early life compared to agonists of other TLRs. CLO75 (TLR7/8) and ssRNA (TLR8) differed with respect to the dynamics across age of cytokine responses. By one month of age, CLO75 induced higher concentrations of TNF α than ssRNA (p = 0.006), but by 2 months of age the two agonists induced similar concentrations (p = 0.735) which remained constant up to 12 months (Figure 6A). IFN γ responses to these two agonists were similar from birth up to 6 months of age, from which point ssRNA induced greater levels of IFN γ than CLO75 through 12 months of age (p = 0.001) (Figure 6B). In contrast, CLO75-induced IL-10 production exceeded that induced by ssRNA at all ages (p < 0.05, data not shown).

TLR8-mediated IFN γ protein production correlates with gene transcription

As the ability to induce IFN γ is of particular importance to host defence and generation of adaptive immunity, we further characterised TLR-mediated IFN γ induction in blood samples from 5 individuals for which longitudinal samples were available from birth, 1 and 12 months of age. In addition, we also measured *IL-12A* and *IL-12B* that encode components of IL-12p70, a cytokine that is important to IFN γ production [48,49] and to Th1 adaptive immune responses but can be challenging to detect at the protein level. Total RNA was purified from blood samples that were stimulated for 4 hours with GardiquimodTM (TLR7), ssRNA (TLR8) and CLO75 (TLR7/8) and mRNA levels measured by real time PCR. Cytokine mRNAs were significantly up-regulated at each time point for each TLR agonist-stimulated condition compared to the unstimulated control (data not shown).

In accordance with the protein data, cytokine mRNA expression patterns appeared similar with lower responses to GardiquimodTM (TLR7) compared to ssRNA (TLR8) and CLO75 (TLR7/8) for most cytokines at each age (Figure 7) except *IL-12A* expression that appeared to be expressed at similar levels in response to each of the agonists at a given age (Figure 7). In addition, the pattern of protein concentrations of IL-10 demonstrated similar expression patterns to *IL-10* mRNA with greater responses to CLO75 than to ssRNA for all ages (Figure 7).

Correlations between mRNA expression levels and protein levels for the same cytokines were analysed as pooled ages. In accord with greater IFN γ response to TLR8 versus TLR7 agonists, significant correlations were noted between TLR

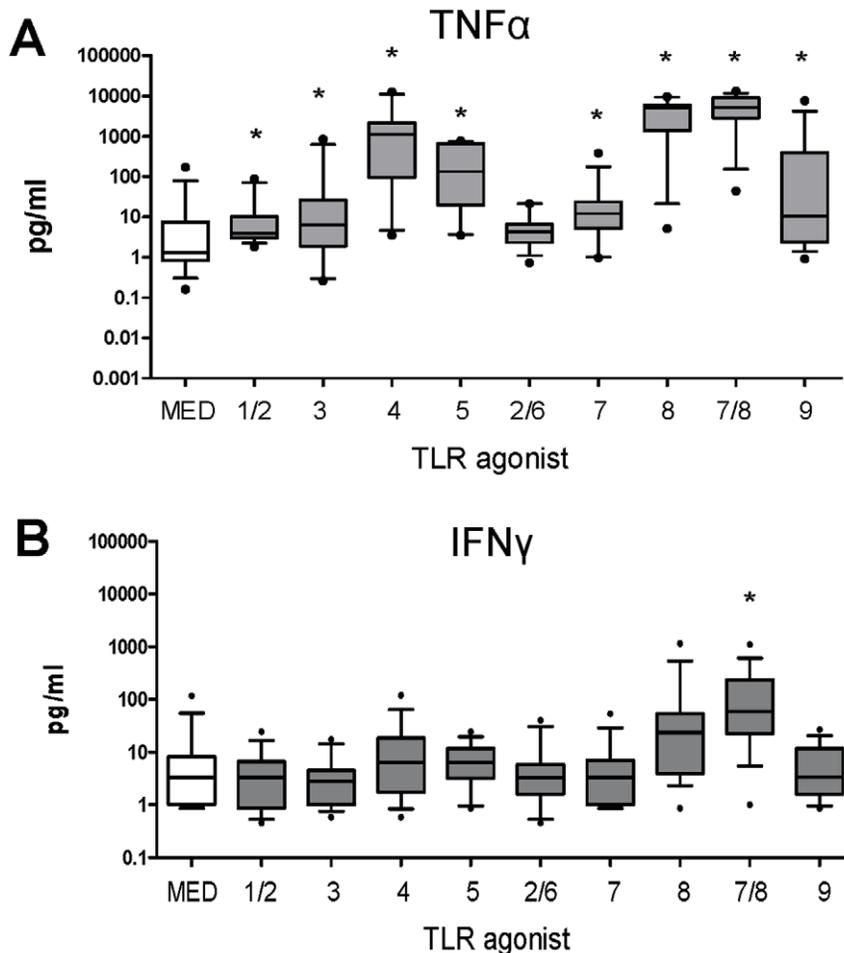


Figure 1. TLR-mediated cytokine production in cord blood. 100 μ l of whole cord blood was cultured overnight with each of the TLR agonists and supernatants recovered for measurement of (A) TNF α and (B) IFN γ as described in Materials and Methods (pg/mL) and presented as box and whisker plots illustrating 10 and 90 percentile error bars, $n=12-15$. Comparisons between cytokine levels in the stimulated samples and the unstimulated samples were analysed using a paired Sign test at 5% significance (* represent significant differences). doi:10.1371/journal.pone.0018185.g001

agonist-induced mRNA expression of *IFNG* and IFN γ protein levels induced by ssRNA ($p=0.005$; Figure 8B) and CL075 ($p=0.027$; Figure 8C), but not GardiquimodTM ($p=0.075$; Figure 8A). Of note, TLR-mediated increases in *IL12A* and *IL12B* mRNA also correlated with TLR-mediated IFN γ protein production, including ssRNA-induced *IL-12A* and *IL-12B* expression ($p=0.017$, $p=0.001$ respectively) as well as CL075- and GardiquimodTM-induced *IL-12B* expression ($p=0.027$ and $p=0.035$, respectively). For the other cytokines studied, the ssRNA- and CL075-induced cytokine mRNA levels at 4 hours of stimulation did not correlate with the corresponding cytokine protein levels at 24 hours.

Levels of TLR-mediated *IL-12A* and *IL-12B* mRNA expression were age-dependent for all three TLR agonists whereas *IFNG* showed similar levels between birth, 1 and 12 months of age (Figure 9 and Figure 7). In response to CL075 (TLR7/8 agonist), expression of *IL-12A* increased from birth to 1 month of age ($p=0.008$) but subsequently decreased by 12 months of age ($p=0.008$). These mRNA patterns corresponded to a similar pattern of IFN γ protein production by the same agonist (comparing for the same subjects): birth vs. 1 month of age ($p=0.002$), and 1 month vs. 12 months of age ($p=0.018$). GardiquimodTM also demonstrated age-dependent expression of

mRNAs encoding *IL-10*, *IL-6* and *TNF* mainly due to low expression levels at birth that increased with age: birth to 1 month of age: *IL-10* ($p=0.008$), and birth to 12 months of age: *IL-10* $p=0.3095$, *IL-6* $p=0.032$, *TNF* $p=0.008$ (Figure 9 and Figure 7).

Discussion

Our study evaluated the ontogeny of TLR-mediated *in vitro* whole blood cytokine responses during the first year of life in infants from The Gambia employing agonists of TLRs 1-9 and blood samples from 8 different age groups. We found that cytokine responses of Gambian infants varied between TLR agonists, with the greatest responses to TLR8 agonists (ssRNA and CL075), substantial responses to TLR4 and TLR5 agonists, and relatively weak responses to TLR1, 2, 3, 7 and 9 agonists at all ages. Thus the ontogeny of responses is TLR-specific with distinct differences across age groups in the first 12 months of life. Most of the previous studies exploring TLR-mediated responses of newborns have focused solely on comparisons between cord blood and adults [4,34,40]. The few studies that have also examined infant responses are limited to resource-rich setting and did not include the wide range of agonists tested in our study [37,39,44,45].

We found that CL075 (TLR7/8) was the only TLR agonist to induce IFN γ at birth but that in general, cord blood pro-

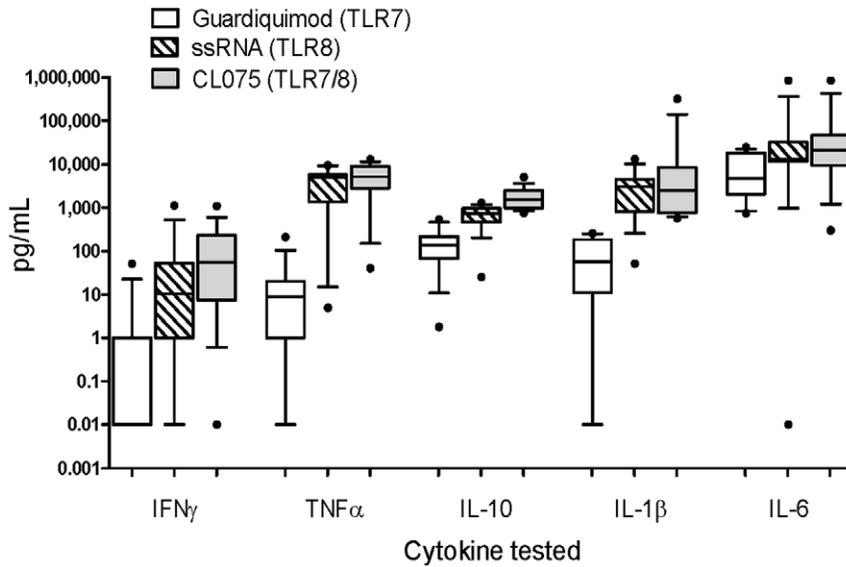


Figure 2. TLR7 and 8 agonist-induced cytokine induction in cord blood. 100 μ l of whole cord blood was cultured overnight with each of the TLR agonists and supernatants recovered for measurement of IFN γ , TNF α , IL-10, IL-1 β and IL-6 as described in Materials and Methods (pg/mL), n = 12–15. Unstimulated values are subtracted from stimulated values and presented as box and whisker plots, n = 12–15. doi:10.1371/journal.pone.0018185.g002

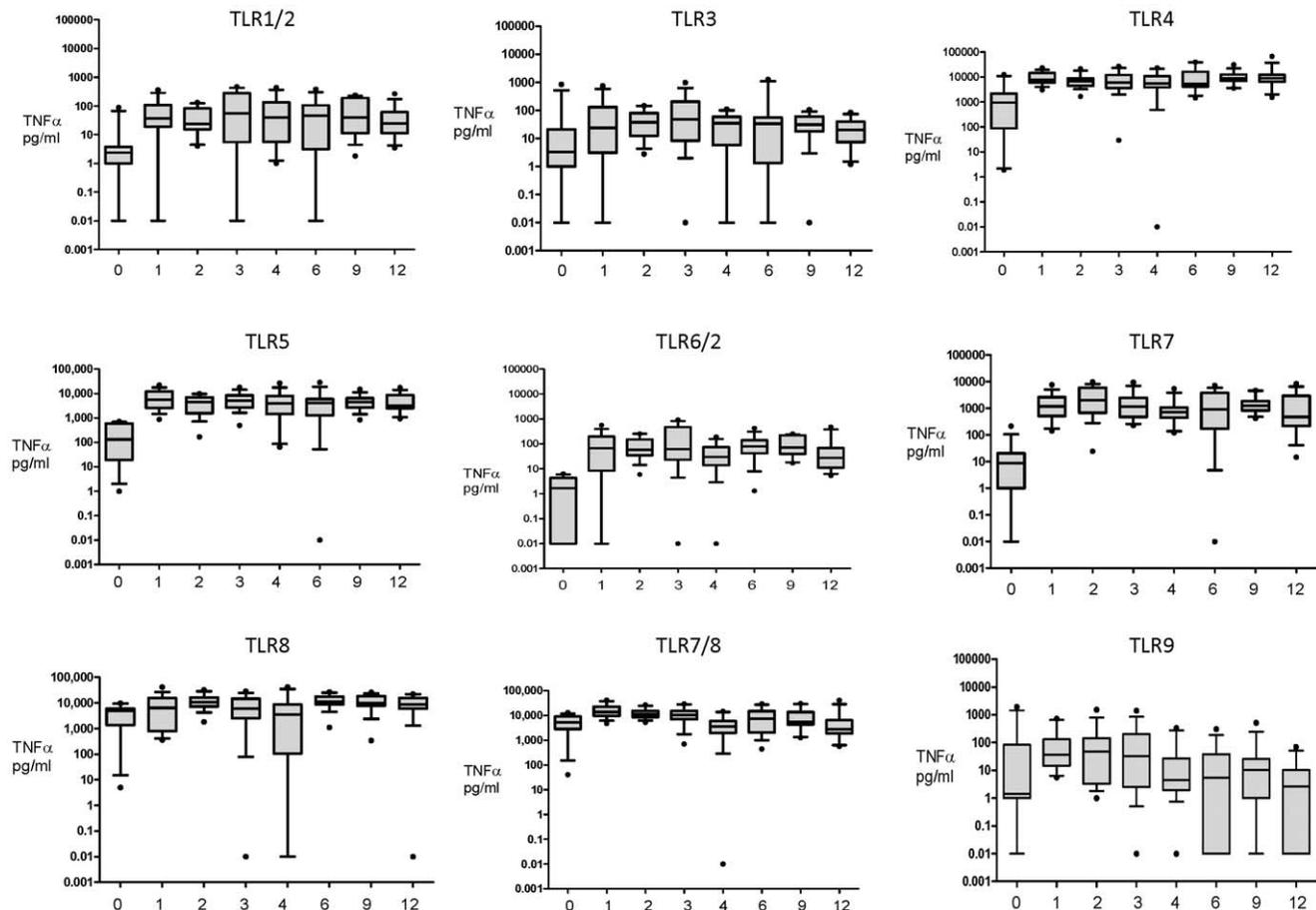


Figure 3. Net TLR-mediated TNF α responses differ between agonists and across the first year of life. 100 μ l of whole blood was cultured overnight with each of the TLR agonists at birth, 1, 2, 3, 4, 6, 9 and 12 months of age (x axis) and TNF α cytokine production (y axis) was measured as described in the Materials and Methods (pg/mL). Unstimulated values are subtracted from stimulated values and presented as box and whisker plots, n = 12–15. doi:10.1371/journal.pone.0018185.g003

	Pam3. (TLR1/2)	Poly I:C (TLR3)	LPS (TLR4)	Flag. (TLR5)	FSL-1 (TLR6/2)	Gard. (TLR7)	ssRNA (TLR8)	CL075 (TLR7/8)	ODN (TLR9)
IFN γ	**	***	***	***	**	***	*	***	
TNF α	*	*	*	***	***	***		**	
IL-1 β	*					*		**	
IL-10	***								
IL-6									

Figure 4. TLR agonists induce greater cytokine production in Gambian newborns at 1 month compared to at birth. 100 μ l whole blood was cultured overnight with each of the TLR agonists and cytokine production (pg/mL) in supernatants was measured as described in the Materials and Methods. Comparisons between cytokine levels at birth and 1 month of age were calculated using a non-parametric Mann Whitney test. *(light grey square) $p=0.05 - 0.019$, ** (medium grey square) $p=0.01 - 0.001$, *** (dark grey square) $p<0.001$. Pam3. = Pam₃CysSerLys₄, Gard. = Gardiquimod.
doi:10.1371/journal.pone.0018185.g004

inflammatory/Th1-polarizing cytokine responses (i.e., TNF α and IFN γ) were lower than at later age groups, in accordance with prior studies [4,34,39]. In contrast, TLR-mediated neonatal IL-6, IL-1 β and IL-10 production was higher or similar at birth compared to later age groups, suggesting that neonatal TLR-mediated responses are biased towards acute phase (IL-1 β and IL-6) and anti-inflammatory (IL-10) cytokines. We have previously shown in 5-day cultures that IL-10 and IL-6 responses to mycobacterial antigens were present at birth [50]. It has been speculated that the bias towards high IL-10 production may reflect the need to dampen potentially over-

exuberant responses to the numerous new antigens to which an infant is exposed in early life [51]. Greater IL-6 to TNF α ratios have been found in cord blood compared to adults, both in response to *in vitro* stimulation [41] and with respect to basal serum levels during the first week of life [40]. Our findings indicate a similar polarisation in our cohort with greater IL-6 to TNF α ratios at birth compared to 12 months of age. Relatively high IL-6 production in newborns likely contributes to initiation of an acute phase response at birth that may serve to clear perinatally-acquired microbes [52]. In addition, IL-6 enhances both differentiation of Th17 cells and production of IL-17,

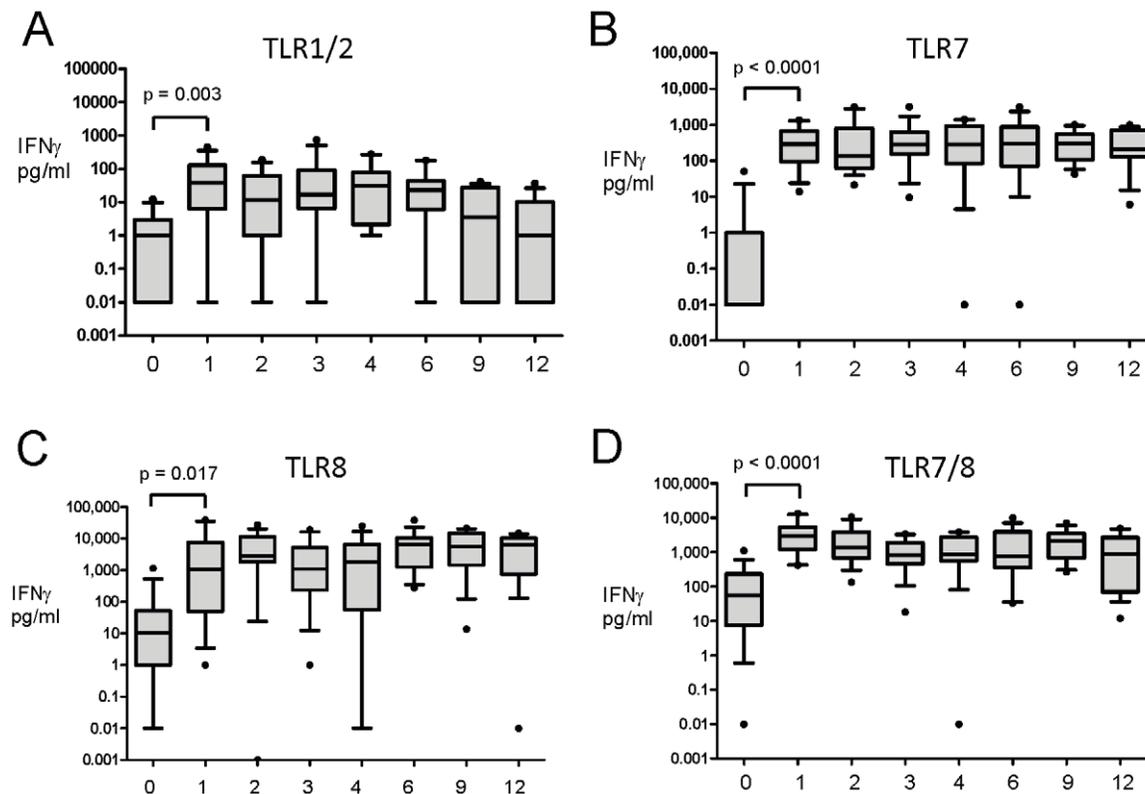


Figure 5. Net TLR-mediated IFN γ responses differ between agonists and across the first year of life. 100 μ l of whole blood was cultured overnight with (A) Pam3CSK4 (TLR1/2 agonist), (B) GardiquimodTM (TLR7 agonist), (C) ssRNA (TLR8 agonist) and (D) CL075 (TLR7/8 agonist) at birth (cord), 1, 2, 3, 4, 6, 9 and 12 months of age (x axis) and IFN γ cytokine production (y axis) was measured in supernatants (pg/mL) as described in Materials and Methods. Unstimulated values are subtracted from stimulated values and data were presented as box and whisker, $n=12-15$.
doi:10.1371/journal.pone.0018185.g005

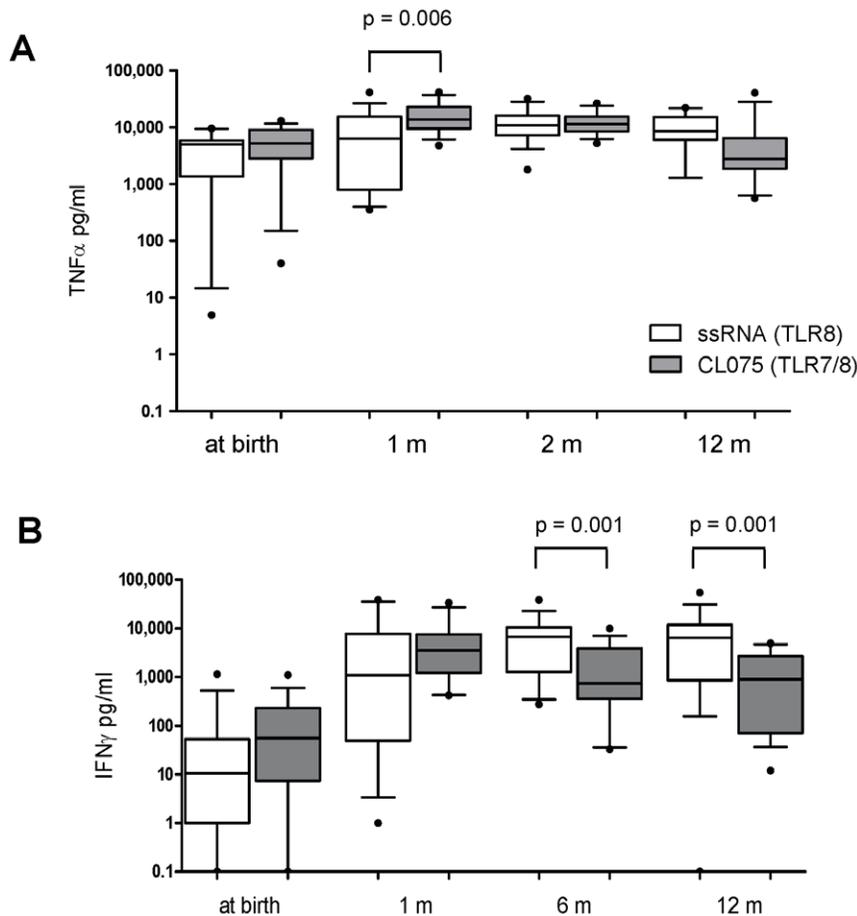


Figure 6. TLR8 agonists induce distinct patterns of TNF α and IFN γ production across the first year of life. 100 μ l of whole blood was cultured overnight with ssRNA (TLR8 agonist) and CL075 (TLR7/8) at birth, 1, 2 and 12 months of age. (A) TNF α and (B) IFN γ cytokine (pg/mL) was measured in supernatants as described in the Materials and Methods. Unstimulated values are subtracted from stimulated values and data are presented as box and whisker plots, $n = 12-15$. doi:10.1371/journal.pone.0018185.g006

and thereby may enhance neutrophil- and antimicrobial peptide-based host defence at neonatal mucosal and epithelial barriers [52–56]. That this neonatal bias towards TLR-mediated IL-6 and IL-10 responses is evident *in vivo* was recently demonstrated in a field trial of alum-adjuvanted pneumococcal conjugate vaccine [3].

For most TLR agonists the major effect of age occurred within the first month of life during which time TLR-mediated TNF α and IFN γ cytokine production increased, suggesting early maturation in the ability to mount these responses. Whether this maturation in cytokine production reflects changes in the adenosine system that serves to limit production of TNF α in human newborn cord blood [41], will be the subject of future studies. The only other study that reported responses in 1 month old infants was conducted by Belderbos *et al* in Holland, which also showed increased pro-inflammatory responses (e.g., IL-12p70 and IFN α) from birth to 1 month of age in response to TLR3, -4, -7 and -9 agonists but similar levels of IL-10 production in response to TLR3, -4 and -9 [39]. That study found that loxoribine (TLR7)-induced IL-10 responses increased by 1 month of age, in accord with our observation of a borderline increase at 1 month of age ($p = 0.05$) using the TLR7 agonist GuardiquimodTM. A Belgian study by Ngyuen *et al* also found that LPS-induced cord blood production of IL-10 and IL-6 was greater than in later age groups (6–9 months and 12 months of age). Taken together, our current

study and those by Belderbos *et al* [39], and Vosters *et al* [44] indicate that for most TLR agonists, IL-10 production is largely similar from birth through to 18 months of age.

A comparison of the few studies of infant TLR function [37,39,44,45] and our current study indicates that Gambian infants have broadly similar TLR-mediated responses to those found in a Western European environment. However, Ngyuen *et al* studying a Belgian cohort found a slower postnatal increase in TNF α production in response to LPS from birth compared to our study, such that adult levels were not reached until 6 months of age [37], whereas our Gambian cohort demonstrated LPS-induced TNF α responses that peaked at 1 month of age and then remained stable to 12 months of age. Earlier maturation of LPS-induced TNF α responses in Gambian infants than in the European infants [37,39,45] may reflect more rapid polarisation to Th1 responses in a resource-poor setting, in accord with the hygiene hypothesis [13,57,58], and may also suggest that TLR4 agonists, currently used in several vaccine formulations (e.g., MPL[59]), may be useful vaccine adjuvants in early infancy.

Although TLR8 agonist-induced cord blood TNF α responses have been shown to be greater than TNF α responses to agonists of TLRs 1–7 [43], little is known regarding cytokine responses to TLR8 agonists such as ssRNA (TLR8 only) and CL075 (TLR7/8 agonist) during the first year of life. Our study confirmed the

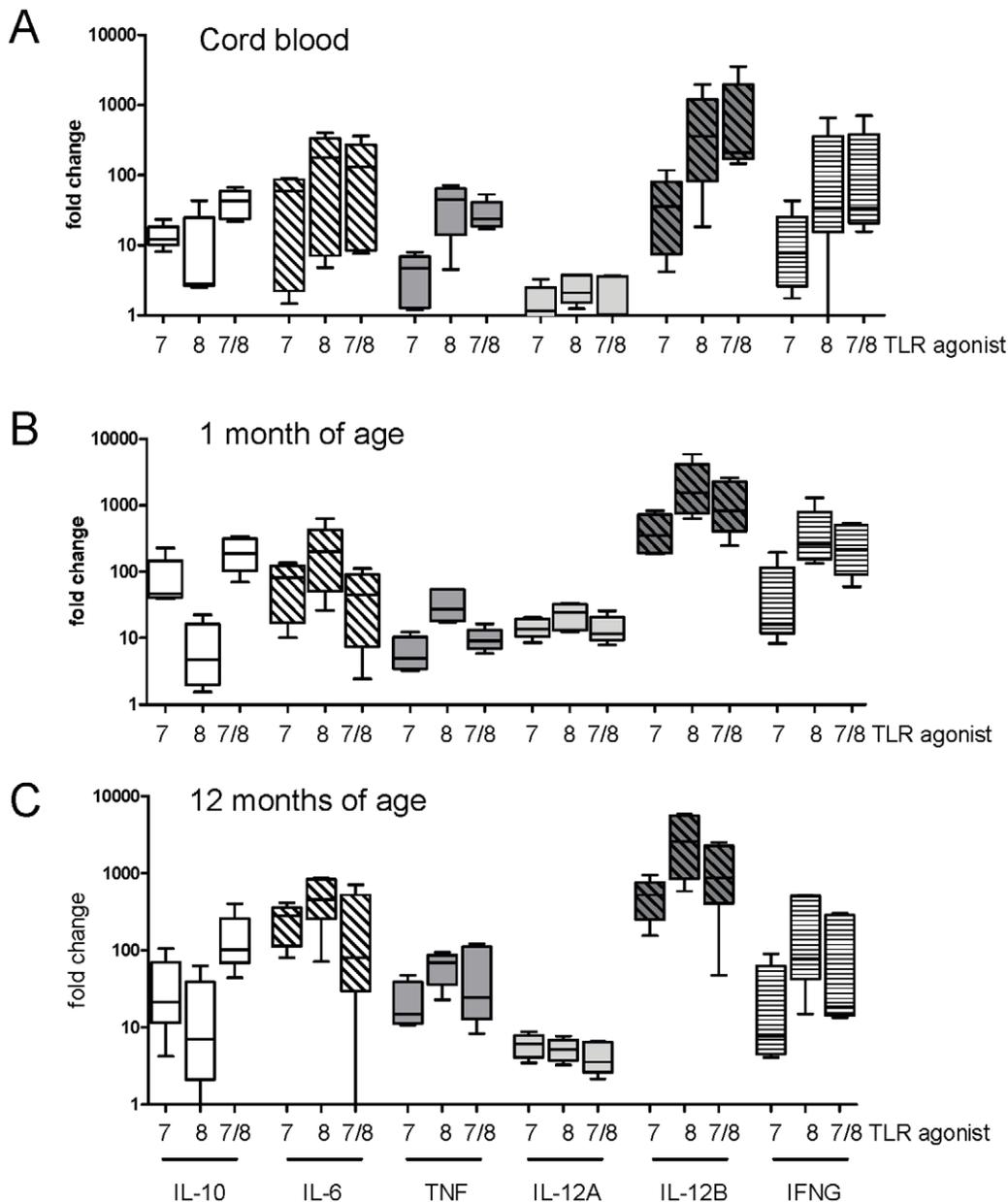


Figure 7. TLR7- and TLR8-mediated cytokine mRNA expression varies between agonists and across the first year of life. 300 μ l of whole blood was cultured for 4 hours with GardiquimodTM (TLR7), ssRNA (TLR8 agonist) and CL075 (TLR7/8) at (A) birth, (B) 1 month and (C) 12 months of age. RNA was purified and cytokine mRNA was quantified by real time PCR as described in Materials and Methods. Ct values were normalised against the housekeeping genes and compared to the unstimulated control (Δ Ct) and fold differences of Δ Ct values between unstimulated and stimulated cultures were calculated ($\Delta\Delta$ Ct). Fold changes are presented as box and whisker plots, n=5. doi:10.1371/journal.pone.0018185.g007

greater responsiveness to TLR8 agonists in cord blood and extended the finding by demonstrating that TLR8 agonists were the predominant inducers of pro-inflammatory cytokines up to 12 months of age in Gambian infants. We also found that the combined TLR7/8 agonist (CL075) induced more TNF α than the TLR7 selective agonist GardiquimodTM or the TLR8 selective agonist ssRNA up to 1 month of age. By 12 months of age however, there was a trend towards reduced TNF α production in response to the combined agonist compared to the single TLR8 agonist. Likewise, IFN γ production was similar between the two agonists up to 6 months of age after which CL075-induced IFN γ diminished relative to ssRNA-induced IFN γ and remained lower

up to 12 months of age. It should be noted however, that in addition to differences between TLR7 and TLR8 selectivity, differences in biochemical structures between ssRNA and the low-molecular weight thiazoloquinolone, CL075 may affect TLR-independent variables (e.g., solubility, protein binding and cell penetration) that may also contribute to distinct bioactivities. The correlations between *IFNG* mRNA and IFN γ protein in response to ssRNA and CL075 indicate that the ontogeny of TLR-mediated IFN γ production is manifest at the transcriptional level. Given the importance of IL-12 to IFN γ production [49,60,61] and the observed correlations of TLR-mediated IFN γ protein with *IL12B* message, TLR8-mediated IFN γ responses may also involve

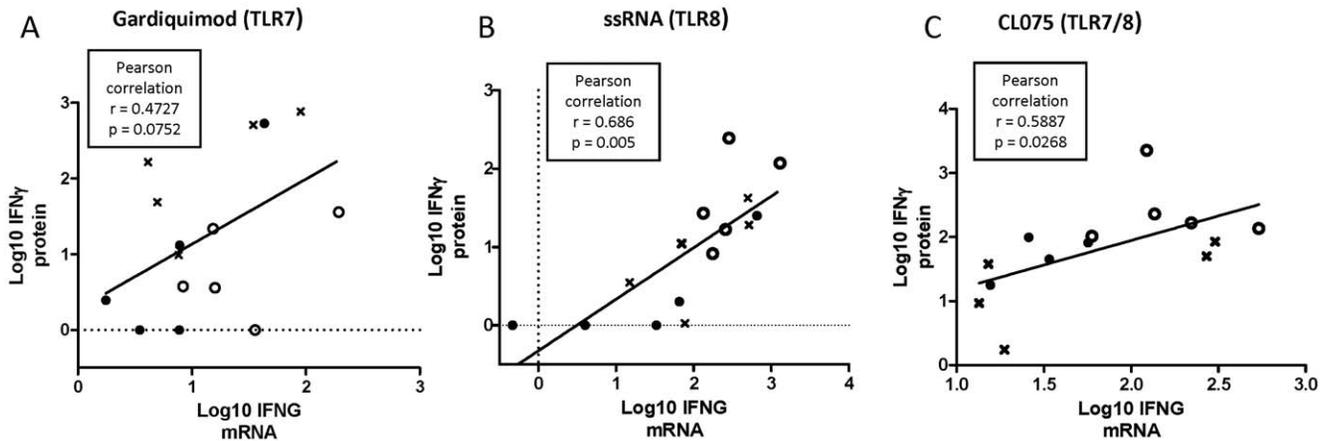


Figure 8. Correlations between ssRNA-induced cytokine mRNA and protein expression. 300 μ l of whole blood was cultured for 4 hours with (A) GardiquimodTM (TLR7), (B) ssRNA (TLR8) and (C) CL075 (TLR7/8). RNA was purified and cytokine mRNA was quantified by real time PCR as described in Materials and Methods. Comparisons between IFN γ protein and IFNG mRNA levels were calculated where stimulated values (Ct and cytokine concentrations) were divided by unstimulated values. The values were log transformed and compared using Pearson correlation test, linear regression line presented in graph, n = 5. At birth (black circles), 1 month (open circles), or 12 months (crosses) of age. doi:10.1371/journal.pone.0018185.g008

activation of *IL-12B* (that encodes for IL-12p40)[49]. However, expression of *IL-12A* (that encodes for IL-12p35) was very low at all three ages tested, in agreement with Western European studies that demonstrate deficiency in TLR-mediated IL-12p35 production by neonatal DCs, contributing to impaired neonatal IFN γ responses [62]. Analysis of TLR-mediated production of additional cytokines that may influence adaptive immune responses, including IL-4 (Th2) and IL-17 (Th17), is of importance and should be included in future studies.

Overall, characterizing the ontogeny of innate immune responses may eventually inform selection of adjuvant vaccine formulations that are tailored for certain age groups. Indeed, recent studies highlight that appropriate immunologic signatures can predict vaccine efficacy [17,63]. To the extent that TNF α and IFN γ may be markers of strong Th1-polarising responses required for induction of cell-mediated immunity, and that the whole blood *in vitro* responses measured in our study may reflect those that would pertain *in vivo*, our data suggest that vaccine adjuvants based on TLR4, -5 or -8 agonists, may be particularly effective in protecting newborns and infants against pathogens requiring cell-mediated immunity.

	Gard. (TLR7)	ssRNA (TLR8)	CL075 (TLR7/8)
IL-12A			
IL-12B			
IFNG			
IL-6			
IL-10			
TNF			

Figure 9. Age-dependent effects of cytokine transcription in response to TLR agonists. 300 μ l whole blood was cultured for 4 hours with GardiquimodTM (TLR7 agonist), ssRNA (TLR8 agonist) and CL075 (TLR7/8 agonist) at birth (cord), 1 and 12 months of age and *IL-12A*, *IL-12B*, *IFNG*, *IL-6*, *IL-10* and *TNF* cytokine mRNA gene transcription was measured. Ct values were normalised against the housekeeping genes and compared to the unstimulated control (Δ Ct). Comparisons of responses from birth, 1 and 12 months of age were made using the Kruskal Wallis test. Grey squares represent significant effects with age, $p > 0.05$. Gard. = Gardiquimod. doi:10.1371/journal.pone.0018185.g009

Supporting Information

Table S1 Summary of the TLR agonists used in the study. (DOC)

Table S2 Analysis for age-dependent evolution of cytokine responses to TLR agonists. 100 μ l of whole blood was cultured overnight with Pam3CSK4 (TLR1/2), Poly (I:C) (TLR3), LPS (TLR4), Flagellin (TLR), FSL-1 (TLR6/2), GardiquimodTM (TLR7), ssRNA (TLR8) and CL075 (TLR7/8) or ODN M362 (TLR9) at birth (cord), 1, 2, 3, 4, 6, 9 and 12 months of age. IFN γ , TNF α , IL-1 β , IL-10 and IL-6 cytokine concentrations (pg/mL) were measured in supernatants as described in Materials and Methods. Comparisons of responses were made (A) from birth to 12 months of age and (B) from 1 to 12 months of age by comparing the proportions of values above and below the median using Fisher's exact test. All grey squares represent significant effects with age ($p < 0.05$) while dark grey squares indicate the effects with age corresponding to a significant decline in cytokine production from 1 to 12 months of age using trend analysis. (DOC)

Acknowledgments

We would like to thank the field workers and nurses at the MRC clinic in Sukuta, and Sally Savage and her staff at the Sukuta Government Health Centre. We also acknowledge the support of the mothers and newborns without which this study could not take place. OL acknowledges the mentorship of Drs. Michael Wessels, Richard Malley, Christopher Wilson

and Gary Fleisher. We thank the Gambian government for their support, in particular the National EPI programme.

Author Contributions

Conceived and designed the experiments: SB KLF OL AJ HW VJP. Performed the experiments: SB JNJ MC CM. Analyzed the data: SB JT

OL. Contributed reagents/materials/analysis tools: SB JT KLF OL. Wrote the paper: SB OL KLF BK. Acquired field data and ran field team: SB UJA ET. Contributed to review and editing of manuscript: SB JT JNJ MC UJA ET VJP CM BK HW AJ KLF OL. Interpretation of the data: SB OL KLF BK.

References

- UNICEF (2009) Reducing Child Mortality: Millennium Development Goals. UNICEF.
- Demirjian A, Levy O (2008) Safety and efficacy of neonatal vaccination. *Eur J Immunol*.
- van den Biggelaar AH, Richmond PC, Pomat WS, Phuanukoonnon S, Nadal-Sims MA, et al. (2009) Neonatal pneumococcal conjugate vaccine immunization primes T cells for preferential Th2 cytokine expression: a randomized controlled trial in Papua New Guinea. *Vaccine* 27: 1340–1347.
- Kollmann TR, Crabtree J, Rein-Weston A, Blimkie D, Thommai F, et al. (2009) Neonatal Innate TLR-Mediated Responses Are Distinct from Those of Adults. *J Immunol* 183: 7150–7160.
- Siegrist CA (2001) Neonatal and early life vaccinology. *Vaccine* 19: 3331–3346.
- Upham JW, Rate A, Rowe J, Kusel M, Sly PD, et al. (2006) Dendritic cell immaturity during infancy restricts the capacity to express vaccine-specific T-cell memory. *Infect Immun* 74: 1106–1112.
- Philbin VJ, Levy O (2009) Developmental biology of the innate immune response: implications for neonatal and infant vaccine development. *Pediatr Res* 65: 98R–105R.
- Prabhudas M, Adkins B, Gans H, King C, Levy O, et al. Challenges in infant immunity: implications for responses to infection and vaccines. *Nat Immunol* 12: 189–194.
- Marodi L (2006) Innate cellular immune responses in newborns. *Clin Immunol* 118: 137–144.
- Willems F, Vollstedt S, Suter M (2009) Phenotype and function of neonatal DC. *Eur J Immunol* 39: 26–35.
- Yan SR, Qing G, Byers DM, Stadnyk AW, Al-Hertani W, et al. (2004) Role of MyD88 in diminished tumor necrosis factor alpha production by newborn mononuclear cells in response to lipopolysaccharide. *Infect Immun* 72: 1223–1229.
- Marchant A, Goetghebuer T, Ota MO, Wolfe I, Ceesay SJ, et al. (1999) Newborns develop a Th1-type immune response to *Mycobacterium bovis* bacillus Calmette-Guerin vaccination. *J Immunol* 163: 2249–2255.
- Belderbos M, Levy O, Bont L (2009) Neonatal innate immunity in allergy development. *Curr Opin Pediatr* 21: 762–769.
- Vasilakos JP, Smith RM, Gibson SJ, Lindh JM, Pederson LK, et al. (2000) Adjuvant activities of immune response modifier R-848: comparison with CpG ODN. *Cell Immunol* 204: 64–74.
- Wu JJ, Huang DB, Tying SK (2004) Resiquimod: a new immune response modifier with potential as a vaccine adjuvant for Th1 immune responses. *Antiviral Res* 64: 79–83.
- Wille-Reece U, Wu CY, Flynn BJ, Kedl RM, Seder RA (2005) Immunization with HIV-1 Gag protein conjugated to a TLR7/8 agonist results in the generation of HIV-1 Gag-specific Th1 and CD8+ T cell responses. *J Immunol* 174: 7676–7683.
- Pulendran B, Li S, Nakaya HI (2010) Systems vaccinology. *Immunity* 33: 516–529.
- Katsenelson N, Kanswal S, Puig M, Mostowski H, Verthelyi D, et al. (2007) Synthetic CpG oligodeoxynucleotides augment BAFF- and APRIL-mediated immunoglobulin secretion. *Eur J Immunol* 37: 1785–1795.
- Didierlaurent AM, Morel S, Lockman L, Giannini SL, Bisteau M, et al. (2009) AS04, an aluminum salt- and TLR4 agonist-based adjuvant system, induces a transient localized innate immune response leading to enhanced adaptive immunity. *J Immunol* 183: 6186–6197.
- Petaja T, Keranen H, Karppa T, Kawa A, Lantela S, et al. (2009) Immunogenicity and safety of human papillomavirus (HPV)-16/18 AS04-adjuvanted vaccine in healthy boys aged 10–18 years. *J Adolesc Health* 44: 33–40.
- Boland G, Beran J, Lievens M, Sasadeusz J, Denticio P, et al. (2004) Safety and immunogenicity profile of an experimental hepatitis B vaccine adjuvanted with AS04. *Vaccine* 23: 316–320.
- Nicholls EF, Madera L, Hancock RE (2010) Immunomodulators as adjuvants for vaccines and antimicrobial therapy. *Ann N Y Acad Sci* Early viewing online.
- Aponte JJ, Aide P, Renom M, Mandomando I, Bassat Q, et al. (2007) Safety of the RTS,S/AS02D candidate malaria vaccine in infants living in a highly endemic area of Mozambique: a double blind randomised controlled phase I/IIb trial. *Lancet* 370: 1543–1551.
- Bejon P, Lusingu J, Olotu A, Leach A, Lievens M, et al. (2008) Efficacy of RTS,S/AS01E vaccine against malaria in children 5 to 17 months of age. *N Engl J Med* 359: 2521–2532.
- Abdulla S, Oberholzer R, Juma O, Kubhoja S, Machera F, et al. (2008) Safety and immunogenicity of RTS,S/AS02D malaria vaccine in infants. *N Engl J Med* 359: 2533–2544.
- Coffman RL, Sher A, Seder RA (2010) Vaccine adjuvants: putting innate immunity to work. *Immunity* 33: 492–503.
- Dillon S, Agrawal S, Banerjee K, Letterio J, Denning TL, et al. (2006) Yeast zymosan, a stimulus for TLR2 and dectin-1, induces regulatory antigen-presenting cells and immunological tolerance. *J Clin Invest* 116: 916–928.
- Davila S, Hibberd ML, Hari Dass R, Wong HE, Sahiratmadja E, et al. (2008) Genetic association and expression studies indicate a role of toll-like receptor 8 in pulmonary tuberculosis. *PLoS Genet* 4: e1000218.
- Uehori J, Matsumoto M, Tsuji S, Akazawa T, Takeuchi O, et al. (2003) Simultaneous blocking of human Toll-like receptors 2 and 4 suppresses myeloid dendritic cell activation induced by *Mycobacterium bovis* bacillus Calmette-Guerin peptidoglycan. *Infect Immun* 71: 4238–4249.
- Tsuji S, Matsumoto M, Takeuchi O, Akira S, Azuma I, et al. (2000) Maturation of human dendritic cells by cell wall skeleton of *Mycobacterium bovis* bacillus Calmette-Guerin: involvement of toll-like receptors. *Infect Immun* 68: 6883–6890.
- Latz E, Franko J, Golenbock DT, Schreiber JR (2004) Haemophilus influenzae type b-outer membrane protein complex glycoconjugate vaccine induces cytokine production by engaging human toll-like receptor 2 (TLR2) and requires the presence of TLR2 for optimal immunogenicity. *J Immunol* 172: 2431–2438.
- Picard C, von Bernuth H, Ghandil P, Chrabieh M, Levy O, et al. (2010) Clinical features and outcome of patients with IRAK-4 and MyD88 deficiency. *Medicine* in press.
- Ku CL, von Bernuth H, Picard C, Zhang SY, Chang HH, et al. (2007) Selective predisposition to bacterial infections in IRAK-4-deficient children: IRAK-4-dependent TLRs are otherwise redundant in protective immunity. *J Exp Med* 204: 2407–2422.
- Levy O, Zarembek KA, Roy RM, Cywes C, Godowski PJ, et al. (2004) Selective impairment of TLR-mediated innate immunity in human newborns: neonatal blood plasma reduces monocyte TNF-alpha induction by bacterial lipopeptides, lipopolysaccharide, and imiquimod, but preserves the response to R-848. *J Immunol* 173: 4627–4634.
- Goriely S, Goldman M (2007) From tolerance to autoimmunity: is there a risk in early life vaccination? *J Comp Pathol* 137(Suppl 1): S57–61.
- De Wit D, Tontonoz S, Orlislaegers V, Goriely S, Boutriaux M, et al. (2003) Impaired responses to toll-like receptor 4 and toll-like receptor 3 ligands in human cord blood. *J Autoimmun* 21: 277–281.
- Nguyen M, Leuridan E, Zhang T, De Wit D, Willems F, et al. (2010) Acquisition of adult-like TLR4 and TLR9 responses during the first year of life. *PLoS One* 5: e10407.
- Schultz C, Rott C, Temming P, Schlenke P, Moller JC, et al. (2002) Enhanced interleukin-6 and interleukin-8 synthesis in term and preterm infants. *Pediatr Res* 51: 317–322.
- Belderbos ME, van Bleek GM, Levy O, Blanken MO, Houben ML, et al. (2009) Skewed pattern of Toll-like receptor 4-mediated cytokine production in human neonatal blood: Low LPS-induced IL-12p70 and high IL-10 persist throughout the first month of life. *Clin Immunol*.
- Angelone DF, Wessels MR, Coughlin M, Suter EE, Valentini P, et al. (2006) Innate immunity of the human newborn is polarized toward a high ratio of IL-6/TNF-alpha production in vitro and in vivo. *Pediatr Res* 60: 205–209.
- Levy O, Coughlin M, Cronstein BN, Roy RM, Desai A, et al. (2006) The adenosine system selectively inhibits TLR-mediated TNF-alpha production in the human newborn. *J Immunol* 177: 1956–1966.
- Vanden Eijnden S, Goriely S, De Wit D, Goldman M, Willems F (2006) Preferential production of the IL-12(p40)/IL-23(p19) heterodimer by dendritic cells from human newborns. *Eur J Immunol* 36: 21–26.
- Levy O, Suter EE, Miller RL, Wessels MR (2006) Unique efficacy of Toll-like receptor 8 agonists in activating human neonatal antigen-presenting cells. *Blood* 108: 1284–1290.
- Vosters O, Lombard C, Andre F, Sana G, Sokal EM, et al. (2010) The interferon-alpha and interleukin-10 responses in neonates differ from adults, and their production remains partial throughout the first 18 months of life. *Clin Exp Immunol*.
- Yerkovich ST, Wikstrom ME, Suriyaarachchi D, Prescott SL, Upham JW, et al. (2007) Postnatal development of monocyte cytokine responses to bacterial lipopolysaccharide. *Pediatr Res* 62: 547–552.
- Lalor MK, Ben-Smith A, Gorak-Stolinska P, Weir RE, Floyd S, et al. (2009) Population Differences in Immune Responses to Bacille Calmette-Guerin Vaccination in Infancy. *J Infect Dis*.
- van den Biggelaar AH, Prescott SL, Roponen M, Nadal-Sims MA, Devitt CJ, et al. (2009) Neonatal innate cytokine responses to BCG controlling T-cell development vary between populations. *J Allergy Clin Immunol*.

48. Jacobson NG, Szabo SJ, Guler ML, Gorham JD, Murphy KM (1995) Regulation of interleukin-12 signal transduction during T helper phenotype development. *Res Immunol* 146: 446–456.
49. Jacobson NG, Szabo SJ, Weber-Nordt RM, Zhong Z, Schreiber RD, et al. (1995) Interleukin 12 signaling in T helper type 1 (Th1) cells involves tyrosine phosphorylation of signal transducer and activator of transcription (Stat)3 and Stat4. *J Exp Med* 181: 1755–1762.
50. Burl S, Adetifa UJ, Cox M, Touray E, Ota MO, et al. (2010) Delaying bacillus Calmette-Guerin vaccination from birth to 4 1/2 months of age reduces postvaccination Th1 and IL-17 responses but leads to comparable mycobacterial responses at 9 months of age. *J Immunol* 185: 2620–2628.
51. Madura Larsen J, Benn CS, Fillie Y, van der Kleij D, Aaby P, et al. (2007) BCG stimulated dendritic cells induce an interleukin-10 producing T-cell population with no T helper 1 or T helper 2 bias in vitro. *Immunology* 121: 276–282.
52. Levy O (2007) Innate immunity of the newborn: basic mechanisms and clinical correlates. *Nat Rev Immunol* 7: 379–390.
53. Kimura A, Kishimoto T IL-6: regulator of Treg/Th17 balance. *Eur J Immunol* 40: 1830–1835.
54. Peck A, Mellins ED (2009) Precarious balance: Th17 cells in host defense. *Infect Immun* 78: 32–38.
55. Dorschner RA, Lin KH, Murakami M, Gallo RL (2003) Neonatal skin in mice and humans expresses increased levels of antimicrobial peptides: innate immunity during development of the adaptive response. *Pediatr Res* 53: 566–572.
56. Bettelli E, Carrier Y, Gao W, Korn T, Strom TB, et al. (2006) Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature* 441: 235–238.
57. Strachan DP (1989) Hay fever, hygiene, and household size. *Bmj* 299: 1259–1260.
58. Rook GA, Hamelmann E, Brunet LR (2007) Mycobacteria and allergies. *Immunobiology* 212: 461–473.
59. Casella CR, Mitchell TC (2008) Putting endotoxin to work for us: monophosphoryl lipid A as a safe and effective vaccine adjuvant. *Cell Mol Life Sci* 65: 3231–3240.
60. Gately MK, Renzetti LM, Magram J, Stern AS, Adorini L, et al. (1998) The interleukin-12/interleukin-12-receptor system: role in normal and pathologic immune responses. *Annu Rev Immunol* 16: 495–521.
61. Robinson DS, O'Garra A (2002) Further checkpoints in Th1 development. *Immunity* 16: 755–758.
62. Goriely S, Vincart B, Stordeur P, Vekemans J, Willems F, et al. (2001) Deficient IL-12(p35) gene expression by dendritic cells derived from neonatal monocytes. *J Immunol* 166: 2141–2146.
63. Pulendran B (2009) Learning immunology from the yellow fever vaccine: innate immunity to systems vaccinology. *Nat Rev Immunol* 9: 741–747.