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Oct4-Induced Reprogramming Is Required for Adult Brain Neural Stem Cell Differentiation into Midbrain Dopaminergic Neurons

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Abstract

Neural stem cells (NSCs) lose their competency to generate region-specific neuronal populations at an early stage during embryonic brain development. Here we investigated whether epigenetic modifications can reverse the regional restriction of mouse adult brain subventricular zone (SVZ) NSCs. Using a variety of chemicals that interfere with DNA methylation and histone acetylation, we showed that such epigenetic modifications increased neuronal differentiation but did not enable specific regional patterning, such as midbrain dopaminergic (DA) neuron generation. Only after Oct-4 overexpression did adult NSCs acquire a pluripotent state that allowed differentiation into midbrain DA neurons. DA neurons derived from Oct4-reprogrammed NSCs improved behavioral motor deficits in a rat model of Parkinson’s disease (PD) upon intrastriatal transplantation. Here we report for the first time the successful differentiation of SVZ adult NSCs into functional region-specific midbrain DA neurons, by means of Oct-4 induced pluripotency.

Introduction

One of the fundamental questions in the field of regenerative neuroscience is whether adult forebrain subventricular zone (SVZ) neural stem cells (NSCs) can efficiently generate neuronal phenotypes other than their native inhibitory olfactory bulb (OB) interneuron populations. Adult SVZ NSCs are primarily fated to generate non-dopaminergic (DA) gamma-aminobutyric acid (GABA)-ergic olfactory bulb (OB) interneurons [1,2,3,4]. This represents an obstacle to the development of successful therapeutic strategies for neurodegenerative diseases, since region-specific phenotypes are warranted for the generation of clinically relevant neurons by mobilization of endogenous neural precursor cells (NPCs) after degeneration or lesion.

With respect of cell therapy for Parkinson’s disease (PD), several pieces of evidence now demonstrate the importance of the midbrain DA neuronal subtype as a determinant of the functional impact of cell-based strategies in animal models of PD [5,6]. The critical challenge is to generate neuronal populations with the phenotypic and molecular properties of midbrain DA neurons in order to achieve proper striatal reinnervation. However, there is still no evidence of the successful manipulation of adult SVZ NSCs toward a midbrain DA neuronal identity suitable for such clinical regenerative purposes. In vitro, midbrain DA neurons have only been efficiently derived from early fetal ventral midbrain and embryonic stem cells (ESCs) from preimplanted blastocysts of embryos [7]. On the contrary, adult SVZ NSCs are more restricted in their capacity to generate neuronal subtypes with a specific regional identity [1,2,8]. In vivo, different strategies have been tested in order to promote the proliferation of endogenous SVZ NPCs, their migration toward the lesioned striatum, and their differentiation into midbrain DA neurons [9,10,11,12]. However, there is no evidence that such strategies promote the generation of functional midbrain DA neurons that integrate into the nigrostriatal DA system [13,14,15].

During development, adult SVZ NSCs lose their competency for neuronal region-specific patterning and therefore acquire a restricted temporal and regional specification [16]. Epigenetic modifications such as histone acetylation and DNA methylation play an important role in regulating such fate determination [17]. Importantly, DNA methylation and histone acetylation state closely correlates with NSC multipotency both in vivo and in vitro [18].

Here, we sought to investigate whether chromatin-modifying agents (such as histone deacetylase inhibitors and demethylating agents) can regulate the capacity of adult SVZ NSCs to differentiate into region-specific neuronal subtypes such as midbrain DA neurons. We found that chromatin-modifying agents increase neuronal differentiation of adult SVZ NSCs without altering their capacity to differentiate into region-specific neuronal phenotypes. Only by Oct4-induced reprogramming could adult SVZ NSCs re-acquire the competency to differentiate into multiple neuronal lineages.


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Materials and Methods

Animals

4–8 week old C57BL/6 mice (Charles River Laboratories) and B6;129-Gt(Rosa26Sor [+ ] tm1(rTA2M2)Jae; Cola1tm12[cre loxP/cre] Jae/J (rTA/Oct4) mice (The Jackson Laboratory) were used in the study. Female Sprague-Dawley rats with unilateral 6-OHDA lesions were obtained from Taconic. All animal procedures were performed in accordance with the guidelines of the National Institutes of Health and were approved by the Institutional Animal Care and Use Committee (IACUC) at McLean Hospital, Harvard Medical School (#09-3/2-6 approved on 09/16/10).

Primary neural stem cell cultures and differentiation

NSC cultures were established from the lateral ventricular walls of 4- to 8-week-old female mice, as described [19]. Briefly, the SVZ was isolated [20] and digested in 0.1% trypsin-EDTA. Cells were plated at a density of 20 cells/µl in growth medium, consisting of mouse NeuroCult NSC basal medium, mouse NeuroCult NSC proliferation supplements, 2 µg/ml heparin, penicillin [100 U/ml]/streptomycin [100 U/ml], EGF (20 ng/ml) and bFGF (10 ng/ml) (all from StemCell Technologies). To analyse cell proliferation, wild-type (n = 3) or rTA/Oct4 (n = 3) primary NSCs were plated at 8000 cells/cm² in neurosphere growth conditions. After 3–4 days, neurospheres were harvested, mechanically dissociated, counted and re-plated under the same culture conditions. For in vitro neurosphere formation assay, NSCs from wild-type (n = 5) and rTA/Oct4 mice (n = 5) were plated in 24-well polyornithine coated plates at a density of 8000 cells/cm² in growth medium with or without doxycycline (DOX) (2 µg/ml; Sigma). Number of neurospheres (diameter ≥100 µm) was counted 7 days after plating [21].

To induce differentiation, cells were manually dissociated and plated on glass coverslips coated with 10 µg/ml poly-L-ornithine and 1 µg/ml laminin at density of ~5×10³ cells/cm². Cells were first expanded and patterned for 4 days after plating in N2 medium supplemented with FGF2 (10 ng/ml), Shh (500 ng/ml), and FGF8 (100 ng/ml) (all from R&D Systems). After 4 days, cells were subsequently differentiated in N2 medium containing ascorbic acid (AA) (200 µM; Sigma) for 10–14 days. In some experiments, NSCs were treated with VPA (1 mM, EMD), TSA (100 nM, EMD), AZA (1 µM, Sigma), BIX-01294 (1 µM, Sigma) or 100 nM TSA/500 nM AZA for 72 hours and then replated in differentiation medium containing 1 mM VPA, 100 nM TSA, 1 µM AZA, 1 µM BIX-01294 or 100 nM TSA/500 nM AZA. After an additional 48 hours, chemicals were withdrawn and differentiation was carried out as described above. Untreated NSCs and dimethyl sulfoxide (DMSO)-treated NSCs were used as control. Three independent experiments were run at least in triplicate.

Bisulfite sequencing

DNA was isolated with a QIAGEN DNeasy kit. Purified genomic DNA was denatured and converted with an Epitect kit QIAGEN. 500 ng of DNA was then bisulfite converted and purified. Bisulfite sequencing was carried out by Genpathway, Inc. Bisulfite conversion specific primers were designed using MethPrimer. PCR products were cloned into the TOPO-TA vector (Invitrogen), transformed into bacteria and plated on agarose plates. Colonies were picked, inserts were PCR amplified using M13 forward and reverse primers and products were visualized on a 1% agarose gel. Amplified PCR products were sequenced using M13 reverse primers. The following primers were used: bisulfite PCR-forward (GTAATGGTTTTGTTTTTGTTTTTTTTTTTTTTT), bisulfite PCR-reverse (CCACCCTTCAACCTTAACCTCTTAAAC).

Generation of NSC-derived iPSCs

NSCs derived from 4-week-old rTA-Oct4 transgenic mice were seeded at a density of 5×10³ cells per 6-well plate in growth medium with DOX (1–2 µg/ml). Forty-eight hours later, cells were further subcultured on irradiated MEFs (GlobalStem) in ESC medium (DMEM supplemented with 15% FBS, nonessential amino acids, L-glutamine/streptomycin, β-mercaptoethanol, and 1000 U/ml LIF) with DOX (2 µg/ml). In some conditions, VPA (0.5 mM) was added to ESC medium for 7–10 days. ES-like colonies were mechanically isolated, and individual cells were dissociated and subsequently replated onto MEFs.

Quantitative Real-Time PCR

Total RNA was extracted with an RNeasy kit (QIAGEN) as described [22]. The expression level of each gene was normalized to endogenous β-actin. Fold-change in gene expression was calculated using ΔΔCt methodology [23]. All the results are from three technical replicates of three independent experiments. Primer sequences are available on request.

Western-Blot

Cells were lysed in NE-PER Nuclear and Cytoplasmic Extraction Kit (Pierce) according to the manufacturer’s protocol. Protein extract (20 µg) was run on a 4%–15% gradient gel (Biorad). The following primary antibodies were used: Oct-4 (1:200, Santa Cruz, H-134), β-TubIII (1:1000, Abcam), GFAP (1:1000, Millipore), GAPDH (1:5000, Millipore).

In vitro differentiation of NPC-derived iPSCs

Cells were harvested by trypsinization and transferred to bacterial culture dishes in ESC medium without LIF. After 3 days, aggregated cells were plated onto gelatin-coated tissue culture dishes and incubated for another 7 days. Differentiated EBs were fixed with 4% paraformaldehyde.

In vitro differentiation of NSC-derived iPSCs into DA neurons

NPC-derived iPSCs were differentiated into DA neurons using a previously described protocol [7], with some modifications. iPSCs (stage 1) were dissociated and plated in bacterial dishes. EBs were allowed 3 days to form in DMEM containing defined 10% FBS, L-glutamine (2 mM), 1× NEAA, HEPES (10 mM), β-mercaptoethanol (1 mM), penicillin (100 U/ml)/streptomycin (100 µg/ml) (stage 2). EBs were allowed 1 day to attach to tissue culture dishes, and neuronal precursors were then selected by incubation in DMEM F-12 medium containing apo-transferrin (50 g/ml), insulin (5 g/ml), sodium selenite (30 nM), fibronectin (250 ng/ml), penicillin (100 U/ml)/streptomycin (100 µg/ml) for 9–10 days (stage 3). Cells were subsequently dissociated by 0.05% trypsin, and neuronal precursors were expanded and patterned for 4 days after plating onto polyornithine/laminin-coated plates at a density of 75,000 cells/cm² in N2 medium supplemented with laminin (1 mg/ml), FGF-2 (10 ng/ml), Shh (500 ng/ml) and murine FGF8 (100 ng/ml) (stage 4). The cells were subsequently differentiated in N2 medium containing AA (200 µM) for 10–14 days (stage 5).

Flow Cytometry and Cell Sorting

Cell sorting was performed as previously described [24]. Briefly, cells were harvested at stage 5 (day 7–10), and stained with

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primary mouse IgM anti-SSEA-1 antibody (0.4 μg/ml; DSHB) for 20 minutes. Cells were then washed, incubated with Alexa Fluor 488 fluorescent secondary antibody. FACS sorting was performed using a FACSAria cell sorter and FACSDiva software (BD Biosciences). A 100 μm nozzle, sheath pressure of 20–25 psi, and an acquisition rate of 1,000–2,000 events per second were used as the standard conditions. Isotype-matched control antibodies were used to set the gate. Annexin V Apoptosis Detection Kit (BD Pharmingen) was used according to the manufacturer’s protocol.

Transplantation into 6-Hydroxydopamine-Lesioned Rats and Analysis of Drug-Induced Rotational Behaviour

NSC-derived iPSCs, sorted for SSEA1 negative expression, were resuspended in HBSS containing 1 μg/ml GDNF (Sigma-Aldrich) at a density of 5 × 10^5 cells/μl. 6-OHDA lesioned rats were grafted into the lesioned striatum with 4 μl of cell suspension as 2 deposits of 100,000 cells using the following coordinates from bregma: site 1: AP +0.4; ML -3; DV -5.0; site 2: AP -0.5; ML -3.6; DV -5.0. Cells were engrafted at a rate of 0.3 μl per minute. Immunosuppression, anesthesia, transplantation, and analgesia were performed as previously described [25]. Rotational asymmetry of 6-OHDA-lesioned rats was analysed after i.p. injection of amphetamine (4 mg/kg; 90 min) or s.c. injection of apomorphine (0.1 mg/kg; 40 min) two weeks before transplantation and at 4, 6 and 8 weeks after transplantation. Functionally lesioned rats were randomly assigned to 2 groups, one control group (n = 6) and one group for the transplantation of differentiated NSC-iPSCs (n = 5). The animals were sacrificed 8 weeks post-transplantation. Brains were removed, postfixed in 4% paraformaldehyde, equilibrated in 30% sucrose, and sectioned on a freezing microtome in 40-μm coronal slices.

Immunofluorescent staining and Stereological Procedures

Immunofluorescent staining was performed as previously described [24]. The following primary antibodies were used: sheep/rabbit anti-TH (1:1,000; Pel-Freez Biologicals), rabbit anti-Pitx3 (1:250, Zymed), mouse anti-En1 (1:50, DSHB), goat anti-Foxa2 (1:100, Santa Cruz), rabbit/chicken anti-BetaIII (Tuji), Covance), rabbit anti-GFAP (1:1000, Dako), mouse anti-SSEA-1 (0.4 μg/ml; DSHB), mouse anti-Oct4 (1:100, Santa Cruz Biotechnology), rabbit anti Nanog (1:100, Bethyl), mouse anti NeuN (1:1000, Chemicon), mouse anti SSEA1 (1:50, Chemicon), rabbit anti GAD67 (1:5000, Sigma), rabbit anti GABA (1:5000, Sigma), mouse anti AFP (1:100, R&D Systems), goat anti brachyury (1:100, Santa Cruz Biotechnology). Appropriate fluorescence-labeled secondary antibodies (AlexaFluor; Invitrogen) were used and nuclei were stained with Hoechst 33342 (5 μg/ml; Sigma-Aldrich). For light microscopy, biotinylated secondary antibodies (1:300, Vector Laboratories) were applied to detect anti-TH antibody, followed by incubation in streptavidin-biotin complex (Vectastain ABC Kit Elite; Vector Laboratories) for 1 h and visualized by incubation in 3,3′-diaminobenzidine (DAB; Vector Laboratories). Alkaline phosphatase (AP) staining was performed using Alkaline Phosphatase.

Detection Kit (Chemicon), according to the manufacturer’s instructions. Confocal analysis was performed using a Zeiss LSM510/Meta Station (Thornwood). Stereology was performed using Stereo Investigator image-capture equipment and software (MicroBright-Field,) and a Zeiss Axioplan I fluorescent microscope. Three coveirsips were counted for each immunostaining.

Table 1. List of reagents used in the study.

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<th>Chemical</th>
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<td>BIX-01294</td>
<td>G9a histone methyltransferase inhibitor</td>
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<td>VPA</td>
<td>HDAC inhibitor</td>
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Results

Chromatin modifying agents do not alter the differentiation potential of mouse adult SVZ NSCs

We first investigated whether epigenetic modifications induced by chromatin modifying agents can increase adult SVZ NSC responsiveness to the patterning signals that regulate midbrain DA neuron specification such as sonic hedgehog (Shh) and fibroblast growth factor 8 (FGF8). We tested the effects of a variety of chemicals that have been shown to enhance cell dedifferentiation by promoting an ESC-like state: 5-aza-2’deoxycytidine (AZA), a DNA methyltransferase (DNMT) inhibitor; BIX-01294, a G9a histone methyltransferase inhibitor; valproic acid (VPA) and trichostatin A (TSA), histone deacetylase inhibitors (HDACs) (Table 1) [26]. The non-toxic concentration of each chemical was determined by exposing adult SVZ NSCs to a range of concentrations and determining cell toxicity by Annexin V/Propidium iodide flow-cytometry analysis at 24 and 48 hours after treatment (data not shown). In order to test whether such chromatin modifying agents increase NSC competency to region-specific neuronal differentiation, NSCs were isolated from the SVZ of adult C57/BL6 mice and grown as neurospheres in the presence of epidermal growth factor (EGF) and fibroblast growth factor 2 (FGF-2). In order to limit the exposure to EGF and FGF-2 that can deregulate the spatial identity and differentiation potential of neural precursors [27], only primary neurospheres were used in these experiments.

NSCs were treated with 1 μM AZA, 1 μM BIX-01294, 1 mM VPA, 100 nM TSA, or 500 nM AZA/100 nM TSA (Table 1) for 72 hours. NSCs were then differentiated upon EGF withdrawal, using a modified protocol developed for the differentiation of mouse ESCs to midbrain DA neurons [7,24] (Fig. 1A). In this protocol, the signaling molecules Shh, FGF-2 and FGF8 induce ventral midbrain DA neuron patterning of neuronal precursors. Final differentiation to mature DA neurons is induced by ascorbic acid (AA) in the absence of growth factors. Differentiation medium was supplemented with 1 μM AZA, 1 μM BIX-01294, 1 mM VPA, 100 nM TSA, or 500 nM AZA/100 nM TSA for an additional 48 hours (Fig. 1A). AZA alone and BIX-01294 at effective concentrations were found to be too toxic when added during differentiation, thus were excluded from further studies. Treatment with HDACi (TSA, VPA) induced a significant increase of β-Tubulin III+ neurons as revealed by immunohistochemistry at day 30 (40.5±0.5%, 55.25±1.6%, 62.5±4% in untreated, TSA and...
Figure 1. Chromatin modifying agents increase neuronal differentiation of adult mouse SVZ NSCs. (A) Schematic representation of the experimental design: adult SVZ NSCs were treated with TSA, TSA/AZA or VPA for 5 days and differentiated using a modified protocol developed for the differentiation of mouse ESCs to midbrain DA neurons. (B) Quantification of β-TubIII⁺ neurons and GFAP⁺ astrocytes in untreated, TSA-, TSA/AZA- or VPA-treated cultures. Treatment with TSA and VPA increased the number of β-TubIII⁺ neurons, whereas TSA, TSA/AZA and VPA decreased the number of GFAP⁺ astrocytes. Error bars indicate SEM. Three independent experiments were performed in triplicate (** p<0.01, *** p<0.001; One-way ANOVA). (C) Immunofluorescence staining for neuron-specific class III β-Tubulin (TUJ1) and the glial-specific marker glial fibrillary acidic protein (GFAP) of untreated, TSA-, TSA/AZA- and VPA-treated cultures. Nuclei were counterstained with Hoechst. (D) Western blot analysis of differentiated
VPA treated cells; p≤0.01 and p≤0.001 compared to untreated respectively) and western blot analysis (Fig. 1B–D). Treatment with TSA, TSA/AZA and VPA decreased the number of GFAP+ astrocytes in differentiated cultures at day 30 (65±1.9%, 37.25±1.8%, 48.34±1.9% in untreated, TSA, TSA/AZA, and VPA treated cells; p≤0.001, p≤0.01 and p≤0.001 compared to untreated cells) (Fig. 1B–C). Western blot analysis confirmed the decrease of astrocytic differentiation after treatment with TSA, TSA/AZA and VPA (Figure 1D). We examined the expression of tyrosine hydroxylase (TH), the rate-limiting enzyme of DA synthesis, and other midbrain DA neuronal markers [engrailed-1 (En1), paired-like homeodomain transcription factor 3 (Pitx3), dopamine transporter (DAT), and nuclear receptor related 1 (Nurr1)] by immunocytochemistry and RT-PCR at day 16, 26 and 30. Some TH+ neurons were transiently detected at day 16 in HDACi treated cultures (data not shown). However, after the completion of the differentiation protocol (day 30), differentiated neurons did not express any midbrain DA marker. We found that differentiated neurons almost exclusively expressed GAD67, a marker for GABAergic interneurons (Fig. 1E).

Epigenetic modifiers increase the expression of pluripotency-associated genes and partial demethylation of Oct4 promoter

We then analysed the expression of pluripotency-associated genes such as Nanog, Sox2, Oct4, and Klf4 in adult SVZ NSCs at 24 and 48 hours after treatment with VPA, TSA or TSA/AZA. We found that TSA alone or TSA/AZA significantly induced the expression of Oct4 and Klf4 at 24 and 48 hours (Fig. 2A). We investigated the DNA methylation profile of Oct4 promoter. Oct-4 is the master regulator of stem cell pluripotency and differentiation [28] and the methylation of its promoter drives the conversion from primitive NSCs (pNSCs) to definitive NSCs (dNSCs) [29],
Figure 3. Short-term overexpression of Oct4 increases proliferation, self-renewal and neuronal differentiation of adult SVZ NSCs.

(A) Adult SVZ NSCs were isolated from doxycycline (DOX)-inducible Oct4 transgenic mice (rTA-Oct4) and grown as neurospheres in growth medium with EGF/FGF-2. Oct4 was induced by 2 μg/mL DOX. Fluorescence images of rTA-Oct4 NSCs cultured for 48 hours, with or without DOX, and stained with an antibody to Oct4. Percentages of Oct4+ cells are indicated. (B) DOX-dependent induction of Oct-4 protein in adult SVZ NSCs from rTA-Oct4 Regional Identity of Adult Brain Neural Stem Cells
transgenic mice were determined by Western blot analysis. Mouse embryonic stem cells (mESC) were used as control. GAPDH was used as loading control. (C) Growth curves of DOX-induced and -uninduced adult SVZ NSCs. Time points represent average values from triplicate measurements and their standard deviations (*p<0.05, Two-way ANOVA with post hoc analysis by Tukey's test). (D) Clonal analysis of SVZ adult NSCs after Oct4 induction. NSCs were grown as neurospheres and treated with DOX or left untreated. After dissociation, single cells were replated and the total number of neurospheres with a diameter > 100 μm was assessed and expressed as % over plated cells (**p<0.01, t-test). Error bars indicate SEM. (E) Schematic representation of the experimental design: SVZ adult NSCs were isolated from rTA-Oct4 transgenic mice and grown as neurospheres in differentiation medium with EGF/FGF-2, with or without DOX. Forty-eight hours after Oct4 induction, neurospheres were dissociated and replated in differentiation medium with Shh, FGF8, FGF-2, with or without DOX. DOX was withdrawn after 48 hours. Cells were finally differentiated with ascorbic acid (AA). (F–I) Immunofluorescence staining for β-TubIII (green, F–G) and GFAP (green, H–I) of adult SVZ NSCs differentiated with or without Oct4 induction. Nuclear were counterstained with Hoechst. The majority of β-TubIII+ neurons co-expressed GABAergic marker GAD67 (red) of differentiated adult SVZ NSCs upon Oct4 induction. The majority of β-TubIII+ neurons co-expressed GABA-ergic marker GAD67 (Fig. 3K). Scale bars: 50 μm (F–I); 10 μm (K).

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Adult SVZ NSCs are reprogrammed to a pluripotent state by exogenous OCT4

We overexpressed Oct-4 in rTA-Oct4 SVZ NSC cultures by DOX and 48 hours later NSCs were plated on mouse embryonic fibroblasts (MEF) in ESC medium. In order to improve the efficiency of reprogramming, ESC medium was supplemented with 0.5 mM VPA for 7–10 days [31] (Fig. 4A). After 5–6 weeks, we were able to select mouse ESC-like colonies [from now on termed as NSCs-derived induced pluripotent stem cells (iPSCs)]. We characterized 2 NSC-derived iPSC clones (clones mm3 and mm4) (Fig. 4B). The estimated reprogramming efficiency of NSCs was 0.001%. We observed no colonies when VPA was omitted. NSC-derived iPSCs expressed alkaline phosphatases (AP), Nanog and SSEA-1, and were morphologically indistinguishable from mouse ESCs (Fig. 4 B–E). As expected, the Oct4 promoter region of NSC-derived iPSCs was found to be hypomethylated (Fig. 4F). Interestingly, the Oct4 promoter region of NSCs derived from iPSCs was hypomethylated compared to adult SVZ NSCs (Fig. 4F and 2B). These iPSC colonies could be expanded up to 25 passages. We then examined the ability of NSC-derived iPSCs to differentiate into the three germ layers by embryoid body (EB) differentiation. EBs derived from NSC-iPSCs expressed markers of the three germ layers including x-fetoprotein (AFP) (endoderm), brachyury (mesoderm), and β-TubIII (ectoderm) as determined by immunocytochemical analysis (Fig. 5 A–D).

Reprogrammed adult SVZ NSCs are successfully differentiated into midbrain DA neurons

To test whether NSC-derived iPSCs were able to generate DA neurons in vitro, we differentiated iPSCs according to the mouse ESC five-stage protocol, with some modifications (Fig. 6A) [7,24]. We induced neural differentiation by EB formation in ITSFn medium (Fig. 6C). Subsequently, the neural precursors were expanded in the presence of the growth factors Shh, FGF-2 and FGF8. Terminal differentiation was induced by growth factor withdrawal in the presence of AA (Fig. 6D). At day 32, we detected a total of 30% β-TubIII+ neurons and a total of 5% TH/β-TubIII+ neurons (Fig. 6B, E). To assess neuronal regional specification, we double-labeled TH-positive neurons with antibody against Pitx3 and En1, markers typically expressed in midbrain DA neurons (Fig. 6 G–H). Importantly, the vast majority of TH-positive neurons stained positive for these midbrain markers, suggesting their proper midbrain regional specification.
in vitro (Fig. 6 G–H). We did not find any colocalization between TH and GABA, a marker typically expressed in OB glomerular interneurons (Fig. 6F). qRT-PCR analysis further confirmed expression levels of midbrain DA markers in differentiated cultures: DAT, PITX3, En1, G-protein-activated inwardly rectifying potassium channel subunit (Girk2), vesicular monoamine transporter (VMAT), aldehyde dehydrogenase 2 (ALDH), Calbindin and Nurr1 (Fig. 6I).

Midbrain DA neurons from reprogrammed adult SVZ NSCs are successfully grafted and reverse drug-induced behaviour in parkinsonian rats

To further analyse the DA fate potential of NSC-derived iPSCs in vivo, we explored the ability of differentiated neurons to survive, integrate and reinnervate the host striatum of 6-hydroxodopamine (6-OHDA)-lesioned rats, a rodent animal model of PD. Adult
NSC-derived iPSCs (clone mm3) were differentiated into DA neurons as described above and transplanted into the striatum of 6-OHDA-lesioned rats. Animals received a striatal graft of $2 \times 10^5$ differentiated cells. In order to avoid tumor formation, SSEA1+ cells were eliminated by cell sorting FACS prior to transplantation (Figure S1). Analysis of amphetamine-induced behaviour showed a significant reduction of ipsilateral rotations at 8 weeks after transplantation (Fig. 7G), indicating significant restoration of the 6-OHDA lesion. We found that rats transplanted with differentiated NSC-derived iPSCs showed a significantly reduced number of apomorphine-induced rotations 8 weeks after engraftment, when compared to the non-transplanted rats (Fig. 7H). Eight weeks after surgery, the animal brains were prepared for morphological analysis. TH staining showed that animals displaying improvement in the behavioural assays had grafts containing large numbers of DA neurons (Fig. 7A–D). Importantly, the transplanted neurons expressed the midbrain DA marker Pitx3 and Foxa2 (Fig. 7E–F). TH-immunoreactive fibres were found to extend into the parenchyma of the host striatum (Fig. 7B,C).

**Discussion**

NSCs acquire a regional identity already at the formation of the neural plate during embryogenesis [32,33]. Therefore, fetal and adult NSCs have passed crucial checkpoints that restrict their developmental capacity to generate region-specific neuronal subtypes.

Epigenetic modifications (i.e. histone code and DNA methylation) play a critical role in regulating adult NSC differentiation and fate determination [34]. Here we investigated whether epigenetic remodeling induced by chromatin-modifying agents can reverse adult SVZ NSC developmental restriction and enable their differentiation into region-specific neuronal subtypes such as midbrain DA neurons. Chromatin remodeling factors can improve the induction of an ESC-like state and have been extensively used to enhance dedifferentiation of somatic cells to the pluripotent state [31]. We showed that chromatin-modifying agents (TSA and TSA in combination with AZA) reactivated specific pluripotency-associated genes such as Oct4 and Klf-4 in adult SVZ NSCs. These agents induced partial demethylation of Oct4 promoter, thus promoting a dedifferentiation toward a primitive neural stage. We observed an increased neuronal differentiation, paralleled by a decreased number of astrocytes in treated NSC cultures. However, these modifications were not sufficient to reverse the resistance of NSCs to the patterning signals that regulate midbrain DA development, since the majority of differentiated neurons showed a GABAergic phenotype. Previous reports have shown the derivation of iPSCs from NSCs [35,36]. Specifically, the overexpression of Oct4 alone induces the reprogramming of NSCs obtained from mouse whole brain and fetal human telencephalon NSCs [37,38]. In the current work we show for the first time that Oct4 overexpression in association with the chromatin-modifying agent VPA induced adult SVZ NSC reprogramming into iPSCs. Such Oct4-reprogrammed NSCs were then successfully differentiated into midbrain DA neurons.

**Figure 5. NPC-derived iPSCs can be differentiated into three germ layers in vitro.** NSC-derived iPSCs were differentiated into three germ layers by embryoid body (EB) differentiation. (A) In vitro EB formation. (B–D) EBs expressed markers of the three germ layers including α-fetoprotein (AFP) (endoderm), brachyury (mesoderm), and β-TubIII (ectoderm). Scale bars: 25 µm (A–C); 50 µm (D). doi:10.1371/journal.pone.0019926.g005

Regional Identity of Adult Brain Neural Stem Cells
Importantly, we show for the first time that single-factor reprogrammed adult SVZ NSCs generated functional midbrain DA neurons that successfully improved the motor behavioural deficits in a rodent model of PD. Such functional motor behaviour recovery in this model is further significant evidence for the correct subtype of DA neurons generated from NSC-derived iPSCs.

With respect to functional cell therapy for the motor symptoms of PD, it is critical to obtain DA neurons that have the molecular properties of midbrain DA neurons [6]. Both substantia nigra (SN-A9) and ventral tegmental area (VTA-A10) DA neurons contribute to widespread and dense axonal arborization [39], and SN-A9 DA neurons are responsible for appropriate striatal reinnervation and behavioural motor recovery in rodent models of PD [5,6].

However, no studies have reported successful in vitro differentiation of adult SVZ NSCs into midbrain DA neurons. When grown as neurospheres and differentiated according to the most dedicated differentiation culture protocols, fetal and adult NSCs only generate a small number of TH⁺ neurons [40]. Furthermore, such differentiated TH⁺ neurons do not express markers of midbrain DA neurons [41,42], which is the required cell type for grafting and appropriate striatal reinnervation in PD. In vivo, mobilization of SVZ endogenous precursor cells has been envisaged as a promising alternative to cell transplantation for the treatment of neurodegenerative diseases including PD [43]. Therefore, different approaches have been tested to promote the differentiation of adult SVZ NSCs to midbrain DA neurons and endogenous NPC proliferation in order to achieve tissue repair.
and functional recovery in several animal models of neurodegenerative diseases [9,10,11,12,44]. However, there is no evidence that such strategies promote the generation of functional midbrain DA neurons that integrate into the injured or naïve nigrostriatal DA system [13,14,15].

Adult NSCs are resistant to the signaling molecules (Shh, FGF8) that control the development of midbrain and hindbrain [45]. The precise mechanism by which such restriction is controlled and maintained in adult SVZ NSCs is still poorly understood. During mouse development, the earliest NSCs can be isolated starting at embryonic day 5.5 (E5.5) [32]. These pNSCs retain ESC characteristics such as high Oct-4 expression and in vitro LIF-dependence [46]. pNSCs are highly responsive to regionalization cues, allowing the efficient generation of region-specific neuronal subtypes. LIF-dependent NSCs with similar antigenic and functional properties can also be isolated from E5.5–7.5 mouse embryos [47]. Between embryonic day 7.5 (E7.5) and E8.5, germ cell nuclear factor (GNFC)-mediated Oct4 promoter methylation drives the conversion from pNSCs to dNSCs [29]. In the developing neuroectoderm and NSC cultures, such transition from pNSCs to dNSCs restricts the potential of dNSCs to form non-neural cell types and their capacity to generate region-specific neuron populations.

Interestingly, we found that Oct-4 overexpression, without reprogramming to an ESC state, increased NSC self-renewal and long-term proliferation in the presence of EGF and FGF-2. In addition, we showed that Oct4 overexpression increases neuronal differentiation of NSC cultures in the presence of Shh, FGF-2 and FGF8. However, our data indicate that such short-term Oct-4 overexpression alone does not confer competence for adult SVZ NSC midbrain regionalization. In contrast, long-term overexpression (30–40 days) of Oct-4 in adult SVZ NSCs grown on MEFs in ESC medium with LIF induced complete dedifferentiation to a pluripotent state. Only after Oct-4 induced reprogramming to a pluripotent state were adult SVZ NSCs successfully patterned to midbrain DA neurons.
In summary, we show that epigenetic modifications are not sufficient to reverse the resistance of SVZ NSCs to the patterning signals that regulate midbrain regional specification. Such modifications do not promote the dedifferentiation of adult SVZ dNSCs toward a primitive neural stage. The current work describes for the first time the reprogramming of adult SVZ NSCs by means of Oct4 overexpression and provides the first evidence that functional midbrain DA neurons can be derived from Oct4-reprogrammed adult SVZ NSCs. These results indicate that the complex and precise regulatory processes responsible for the regional specification of adult NSCs are irreversible, and further de-differentiation steps are required to regain the competency to generate region-specific neuronal phenotypes. These findings have major fundamental scientific and practical implications for regenerative neuroscience.

Supporting Information

Figure S1 Elimination of SSEA1+ cells from NSC-derived iPSC neuronal cultures by cell sorting. (A–C)

References

15. de Chevigny A, Cooper O, Vinuela A, Reske-Nielsen C, Lagace DC, et al. (2008) Fate mapping and lineage analyses demonstrate the production of a large number of striatal neuronoblasts after transforming growth factor alpha and noggin striatal infusions into the dopamine-depleted striatum. Stem Cells 26: 2349–2360.

Neuronal NSC-derived iPSC cultures after sorting based on SSEA1 expression. After sorting, cells were replated onto tissue culture dishes in N2 medium with AA. Three days after sorting, SSEA1− sorted cells displayed mostly neuronal morphology, whereas the SSEA1+ sorted cells exhibited an undifferentiated ES cell morphology. (D–F) Immunofluorescence images of neuronal cultures 3 days after sorting stained for β-TubIII (green), TH (red) and SSEA1 (blue). Scale bars: 50 μm (A–F).

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