Absolute Enrichment: Gene Set Enrichment Analysis for Homeostatic Systems

The Harvard community has made this article openly available. Please share how this access benefits you. Your story matters.

Citation

Published Version
doi://10.1093/nar/gkl766

Accessed
August 7, 2018 8:21:30 PM EDT

Citable Link
http://nrs.harvard.edu/urn-3:HUL.InstRepos:8123170

Terms of Use
This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA

(Article begins on next page)
The GSEA algorithm is easily generalized to any procedure that results in a ranking of genes in an expression experiment. Although Mootha (1) implemented a 'signal-to-noise ratio' for his first implementation, other statistics more relevant to particular studies can be used in implementing the GSEA. For example, in comparisons of two groups that are a matched time series, the paired t-test can be the more appropriate test, and the paired t-statistic is used to rank the genes in such systems. The use of the paired t-statistic (or any other statistic) is driven by the nature of the experiments and is merely the ordering metric and does not address the challenge of identifying bidirectionally perturbed groups of genes. The method of AE, as described below, does not however depend on any particular metric (for the same reasons that GSEA is metric independent).

Gene set enrichment is most often used when one-gene-at-a-time analysis reveals no significant differential expression but when a set of genes might. In this paper, we describe the Absolute Enrichment (analysis) or AE which takes both up and down regulations into account (using the absolute signal-to-noise ratio or absolute SNR as the ranking metric) as well as the standard GSEA for comparison.

MATERIALS AND METHODS

We will first describe Mootha’s (1) original approach to the implementation of the GSEA. Next, we will describe our AE approach using the absolute SNR as the ranking metric.

The analyses presented in this paper were conducted on datasets that were generated from RNA hybridizations to HG-U133A Affymetrix human genechips. Thus, we created gene sets for this genechip.

Standard GSEA using the signal-to-noise ratio SNR as the ranking metric

Given an expression dataset that has an ‘affected’ group and a ‘control’ group, one starts by taking (for each gene or probeset) the average of all the controls and the average for all the affecteds. The difference between the average values for each gene is calculated. This difference is then divided by the standard deviation of the expression values to obtain the signal-to-noise ratio (SNR) for each gene. The SNR values are then ranked from highest to lowest, and the gene sets are analyzed based on this ranking.
of the affected and the average value of the control group is taken and then divided by the sum of the standard deviations of each group to give us the signal-to-noise ratio statistic. The largest positive number then is the most upregulated. GSEA as described by Mootha (1) only looks at perturbation in one direction (e.g. upregulation) and the genes are ordered by signal-to-noise ratio (giving us an ordering based on upregulation). Each row contains the data pertaining to a single probeset or gene and thus each row is reordered based on the ranking metric (SNR in Mootha’s algorithm).

Next the Kolmogorov Smirnov statistic is calculated according to the following equations:

\[
X_N = \frac{G}{\sqrt{N - G}}
\]

if the gene is not part of the gene set, and

\[
X = \sqrt{\frac{N - G}{G}}
\]

if the gene is part of the gene set, where \(G\) is the number of genes in the gene set and \(N\) is the total number of genes in the dataset.

Next a running sum is obtained on the (reordered) dataset. The maximum of the running sum is the value of the enrichment score.

Essentially the enrichment score is a measure of the ‘enrichment’ of the gene set at the top of the list of (the reordered) genes. The highest enriched gene set then is the most significantly differentially expressed gene set in the system under study.

This gene set is then tested for significance. If the columns (pertaining to sample conditions belonging to either control or affected) are randomly shuffled over the course of 1000 permutations, the number of times each gene set comes to the top divided by 1000 gives us an estimate of the \(P\)-value (1). If the \(P\)-value is higher than a predefined value (e.g. >0.05) then that gene set is not significantly differentially expressed in the analysis.
The ranking metric in the GSEA and other metrics (e.g. the $t$-statistic)

The ranking metric used in the GSEA is not fixed to a certain type. Mootha et al. (1) averaged each data class or condition and took the difference (and then divided by the sum of the standard deviations of each data class) to obtain the SNR. Mootha’s (1) data are replicates at the same time point. There are no pairwise comparisons to be made. In the case of a time series where we have a paired control and affected sample at each time point, the paired $t$-statistic may be more appropriate. The AE just like the GSEA is independent of ranking metric—either the SNR or the paired $t$-statistic or essentially any metric that discriminates between the ‘affected’ and ‘control’ classes can be used.

To be able to generate enough permutations to run the P-test (for significance) without generating repeats, we require a relatively large number of affected and control samples. Eight samples (or more) in each class are usually enough to meet this criteria.

AE

Rather than ordering the genes by SNR, a value that goes from negative to positive, we simply take the absolute value thereby lumping together up- and down-regulated components of the homeostatic system. The datasets analyzed in this paper used this absolute value statistic in implementing the AE.

RESULTS

Applications of the AE

Datasets available at GEO datasets (4,5) at the NCBI website were analyzed using our AE analysis. One particularly illuminating dataset (using the HG-U133A genechip) measured gene expression in uterine fibroids (6). On running the AE analysis on this dataset, we found that the gene set c7_U133_probes from Mootha’s original gene set grouping was topmost ranked (see Figure 2 for the expression pattern of this gene set). This gene set, however, was not the top ranked gene set in either the up-regulated analysis or the down-regulated analysis. The top ranked gene set in the standard GSEA analysis was the OXPHOS gene set.

As in Mootha’s work (1), the SNR metric was used to rank the genes (however, absolute values were taken). The c7 gene set is number three in the down-regulated analysis, number five in the up-regulated (or standard GSEA) analysis, and then is top ranked in the AE analysis. The SNRs for the genes in this gene set are given in Figure 2.

The c7 gene set was then entered into EASE (a utility for calculating overrepresentation of Gene Ontology (GO) annotations in lists of genes) (7,8), (http://david.niaid.nih.gov/david/ease.htm) to see which GO categories are enriched. The top ranked ‘biological process’ GO category is ‘cell adhesion’, the top ranked ‘molecular function’ GO category is ‘extracellular matrix structural constituent’ and the top ranked ‘cellular component’ GO category is ‘extracellular matrix’. Table 1 lists the top ranked GO categories pertaining to each of these GO categories separately (the aggregate rank obtained through EASE is given in the third column).

Nine other datasets were studied and four of these showed a different gene set appearing as the top ranked AE gene set compared with either the up- or down-regulated analyses. These results are presented in Table 2.

DISCUSSION

The AE may bring important insights to gene expression datasets

The AE can identify important gene sets that may not be identified through either the up- or down-regulated analyses. For example, in the uterine fibroid dataset, the c7 gene set was top ranked. This gene set was then analyzed through EASE. This gene set was found to be enriched for all three GO categories pertaining to the Extracellular matrix (Table 1 lists the rankings of the three GO categories derived from this gene set through the use of EASE).

According to (9) ‘Uterine fibroids (correctly called leiomyomas or myomas) are benign myometrial neoplasms enriched in extracellular matrix (ECM)’. This paper lends support to the use of the AE, because it shows us that the gene set that has obtained top rank in the AE is made up of components that others have shown to be important in that system.

To further validate the c7 group of (possibly) coregulated genes, we studied the up- and down-regulated components

Figure 2. SNR scores from the uterine fibroid paper. The right graph is an absolute value plot (to show more clearly the symmetry in the way the gene set responded to enrichment).
of this gene set to see if the positive or negative expression values in the leiomyomas versus control have previously been shown to have similar signaling either in leiomyomas of the uterus or in other similar and analogous pathologies.

To perform the above analysis, all the genes from this gene set were run through EASE. The highest rated gene set genes were then sub-extracted (pertaining to ECM). These were then segregated into positive and negative ones (with

### Table 1. Top 24 GO categories (separately ranked by the GO categories: Biological process, molecular function and cellular component) for the top ranked gene set in the AE analysis from the uterine fibroid dataset analyzed through EASE

<table>
<thead>
<tr>
<th>Rank</th>
<th>Top groups from GO category</th>
<th>Overall rank within all 3 GO categories</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Cell adhesion</td>
<td>2</td>
</tr>
<tr>
<td>2</td>
<td>Cell communication</td>
<td>7</td>
</tr>
<tr>
<td>3</td>
<td>Morphogenesis</td>
<td>9</td>
</tr>
<tr>
<td>4</td>
<td>Development</td>
<td>12</td>
</tr>
<tr>
<td>5</td>
<td>Organogenesis</td>
<td>13</td>
</tr>
<tr>
<td>6</td>
<td>Cell growth</td>
<td>16</td>
</tr>
<tr>
<td>7</td>
<td>Regulation of cell growth</td>
<td>17</td>
</tr>
<tr>
<td>8</td>
<td>Cellular process</td>
<td>18</td>
</tr>
<tr>
<td>9</td>
<td>Skeletal development</td>
<td>19</td>
</tr>
<tr>
<td>10</td>
<td>Regulation of cellular process</td>
<td>21</td>
</tr>
<tr>
<td>11</td>
<td>Regulation of biological process</td>
<td>22</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Rank</th>
<th>Top groups from GO category</th>
<th>Overall rank within all 3 GO categories</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Extracellular matrix structural constituent</td>
<td>3</td>
</tr>
<tr>
<td>2</td>
<td>Cell adhesion molecule activity</td>
<td>6</td>
</tr>
<tr>
<td>3</td>
<td>Structural molecule activity</td>
<td>8</td>
</tr>
<tr>
<td>4</td>
<td>Extracellular matrix structural constituent conferring tensile strength</td>
<td>11</td>
</tr>
<tr>
<td>5</td>
<td>Calcium ion binding</td>
<td>15</td>
</tr>
<tr>
<td>6</td>
<td>Glycosaminoglycan binding</td>
<td>20</td>
</tr>
<tr>
<td>7</td>
<td>Insulin-like growth factor binding</td>
<td>23</td>
</tr>
</tbody>
</table>

### Table 2. AE, up and down-regulated analyses conducted on 10 datasets

<table>
<thead>
<tr>
<th>Dataset analysis</th>
<th>AE top gene set</th>
<th>Up-regulated top gene set</th>
<th>Down-regulated top gene set</th>
<th>Up&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Down&lt;sup&gt;b&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td>Uterine fibroid</td>
<td>c7_U133_probes</td>
<td>OXPHOS</td>
<td>c9_U133_probes</td>
<td>5</td>
<td>3</td>
</tr>
<tr>
<td>COPD</td>
<td>Human_mitoDB</td>
<td>MAP00040_Glutathione_metab</td>
<td>Human_mitoDB</td>
<td>116</td>
<td>1</td>
</tr>
<tr>
<td>Endothelin fibroblasts</td>
<td>c7_U133_probes</td>
<td>c7_U133_probes</td>
<td>MAP00100_Sterol_biosyn</td>
<td>1</td>
<td>17</td>
</tr>
<tr>
<td>Male female hypotal</td>
<td>c7_U133_probes</td>
<td>c34_U133_probes</td>
<td>c9_U133_probes</td>
<td>194</td>
<td>2</td>
</tr>
<tr>
<td>Lung cancer motexion</td>
<td>OXPHOS</td>
<td>Mitochondr</td>
<td>c11_U133_probes</td>
<td>3</td>
<td>6</td>
</tr>
<tr>
<td>Sarcoma and hypoxia</td>
<td>c24_U133_probes</td>
<td>c7_U133_probes</td>
<td>c24_U133_probes</td>
<td>169</td>
<td>1</td>
</tr>
<tr>
<td>Squamous lung cancer</td>
<td>Human_mitoDB</td>
<td>c7_U133_probes</td>
<td>c9_U133_probes</td>
<td>1</td>
<td>187</td>
</tr>
<tr>
<td>Tumor cell topoism.</td>
<td>Human_mitoDB</td>
<td>OXPHOS</td>
<td>c4_U133_probes</td>
<td>2</td>
<td>34</td>
</tr>
<tr>
<td>Breast cancer</td>
<td>OXPHOS</td>
<td>c2_U133_probes</td>
<td>c9_U133_probes</td>
<td>8</td>
<td>3</td>
</tr>
<tr>
<td>Bladder SMC stretch</td>
<td>c7_U133_probes</td>
<td>OXPHOS</td>
<td>c7_U133_probes</td>
<td>11</td>
<td>1</td>
</tr>
</tbody>
</table>

<sup>a</sup>Refers to the ranking of the top AE gene set on the up-regulated list.

<sup>b</sup>Refers to the ranking of the top AE gene set on the down-regulated list.
Caveat in conclusions drawn through the use of EASE about GO categories

GO only annotates \(\sim 60\%\) of all genes for *Homo sapiens* (19). This creates a bias in any analysis that uses GO. This bias may be present in the validation step that we conducted through the use of EASE since EASE makes use of GO derived categories.

**Down-regulated gene sets are just as important to study as up-regulated and absolute enrichment gene sets**

It should be noted that the traditional GSEA talks only of up-regulated gene sets. While upregulated groups of (possibly coregulated) genes are important to study, it may be equally important to look at down-regulated gene sets. When gene sets are connectable to GO biological process categories (those that come closest to defining pathways), those gene sets that are down-regulated, then talk of (possible) pathways that may be turned off in the system under study.

It would be important to know which groups of genes are turned off just as it is important to know those that are turned on. For example, the turning off of tumor suppressor genes has profound effects on the cell phenotype as it leads to the development of cancer, and this fact is probably just as important as the turning on of oncogenes in the cell. We can think of an example outside biology that can also illustrate the importance of looking at downregulation. When the brakes of an automobile are working (and therefore on), the automobile can be safely driven. When the brakes are not working (and therefore off and not able to function), the automobile is highly dangerous to drive and is prone to accidents. Taking this analogy further to see how homeostatic mechanisms can be captured, we see that if the automobile is running at very high speeds it could be that this is a normal condition (fast driving). However, when we combine this with absent brakes, we obtain the homeostatic picture where the automobile is running fast because another component that controls or suppresses its function is now defective and ‘downregulated’.

**Gene sets are not necessarily pathways and GO gene categories should not be mixed**

At the GO website, it is stated that ‘A biological process is a recognized series of events or molecular functions. A biological process is not equivalent to a pathway, although some GO terms do describe pathways.’ Thus, although some GO biological process categories may be deemed as pathways, most GO categories are much simpler. Thus, we should not necessarily draw pathway conclusions from the use of GO categories (although some may pertain to pathways).

**Absolute enrichment is not asymmetric because of the concurrent use of down regulations along with up regulation**

It may seem that up- and down-regulated data are asymmetric. It may seem that there is no theoretical limit to up regulation, while there may seem to be a limit to down regulation since we cannot go below zero in gene expression. However, this is not necessarily the case. Up and down regulations are defined relative to another condition. If we take up regulation to be the affected minus the control while taking down regulation to be control minus affected, we see that there does not need to be a limit to the level of expression in the control. Thus, the definition of up and down regulated is arbitrary depending on whether we subtract control from the affected or vice versa.

We further checked the uterine fibroid dataset for any asymmetry in the control versus the affected along numerous metrics such as largest absolute expression in the control and affected, largest absolute difference value between affected and control and vice versa, and largest signal-to-noise ratio between affected and control and vice versa. Along each of these metrics, we found consistent symmetry between the down-regulated expression values and up-regulated expression values. Thus, the AE statistic does not suffer from any type of asymmetry because of the use of the down-regulated limb in our analysis.

The AE statistic is usually not more likely to extract gene sets from the middle of the ranked list (by enrichment score) of gene sets in either the up-regulated or the down-regulated rankings.

We also checked to see if the AE statistic is likely to pick up those gene sets that are not near the top of either the up- or down-regulated gene set rankings. The results of our analyses on nine additional datasets (and also the results from the uterine fibroid dataset) are shown in Table 2. We note that the AE statistic often captures gene sets that are typically near the top or bottom of either the up- or down-regulated gene set lists (204 gene sets were run) but sometimes far enough away from the ends to be missed.

We note that even if the top ranked AE gene sets are far from the up- or down-regulated lists, this is not necessarily a sign of any error since it could just be that in the system under study most gene sets that were defined turned out to be heavily either up or down regulated and those that were both up and down regulated therefore found themselves in the center.

Further, we also note that the AE is more likely to capture higher levels of differential expression precisely because it makes up- and down-regulated components of the gene expression symmetric thus capturing more extremely differentially expressed genes (depending on the gene sets) vis-à-vis just up- or down-regulated analyses which may be more likely to capture genes from the middle of the reordered lists of genes (reordered by the ranking metric such as the absolute SNR).

Although we found a four out of nine or 44\% chance of discovering new gene sets in our AE analysis that were not top ranked in either the up- or down-regulated analyses, we feel that this number itself may be reflective of how gene sets themselves have traditionally been defined. Many of the ‘already-made’ gene sets that we obtained from the literature and the ones that we constructed ourselves were created from clustering techniques applied to gene expression datasets that made use of solely either up- or down-regulated gene groups. Thus, if the gene sets themselves are created from such solely up- or solely down-regulated clusters, then they may tend to only capture those aspects of other datasets to which they are applied in analyses. We suspect that if gene sets are created using bidirectional clustering techniques then we expect the AE to capture these gene sets more often.
This line of thought also creates a justification for the use of bidirectional clustering in capturing homeostatic systems in analyzing gene expression datasets.

ACKNOWLEDGEMENTS

Funding to pay the Open Access publication charges for this article was provided by Children’s Hospital Informatics Program. Dr. Kohane is partially supported by National Institutes of Health National Center for Biomedical Computing grant 5U54LM008748-02.

Conflict of interest statement. None declared.

REFERENCES