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Actin disassembly by coflin, coronin, and Aip1 occurs in bursts and is inhibited by barbed-end cappers

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Introduction

Actin filaments turn over rapidly in cells, and their assembly through barbed end growth is thought to generate protrusive force. Rapid assembly must be balanced by equally rapid disassembly to replenish the monomer pool and allow remodeling of cell shape. Disassembly is arguably the least understood step in the actin dynamics cycle (Pollard and Borisy, 2003). We need to elucidate its mechanism to understand the morphogenesis of dynamic actin assemblies and answer the fundamental question of how polymerization dynamics transduce the energy of ATP hydrolysis into mechanical work. Physiological disassembly mechanisms must fulfill two requirements: (1) fast kinetics and (2) disassembly in a cytoplasm that contains high concentrations of polymerizable monomers. We here image the disassembly of single actin filaments by coflin, coronin, and actin-interacting protein 1, a purified protein system that reconstitutes rapid, monomer-insensitive disassembly (Brieher, W.M., H.Y. Kueh, B.A. Ballif, and T.J. Mitchison. 2006. J. Cell Biol. 175:315–324). In this three-component system, filaments disassemble in abrupt bursts that initiate preferentially, but not exclusively, from both filament ends. Bursting disassembly generates unstable reaction intermediates with lowered affinity for CapZ at barbed ends. CapZ and cytochalasin D (CytoD), a barbed-end capping drug, strongly inhibit bursting disassembly. CytoD also inhibits actin disassembly in mammalian cells, whereas latrunculin B, a monomer sequestering drug, does not. We propose that bursts of disassembly arise from cooperative separation of the two filament strands near an end. The differential effects of drugs in cells argue for physiological relevance of this new disassembly pathway and potentially explain discordant results previously found with these drugs.
Although our previous work began to define roles for each factor in the purified protein system (Brieher et al., 2006), it did not determine how the full purified system promotes disassembly on a single-filament level. We address this question here.

**Results**

**Actin filaments disassemble in bursts**
We imaged the disassembly of single, fluorescently labeled actin filaments catalyzed by the three-component purified protein system consisting of coflin, coronin, and Aip1 (Brieher et al., 2006). Although our previous work began to define roles for each factor in the purified protein system (Brieher et al., 2006), it did not determine how the full purified system promotes disassembly on a single-filament level. We address this question here.

**Figure 1.** Actin filaments disassemble in bursts in coflin, coronin, and Aip1. (A) Time-lapse wide-field epifluorescence images of fluorescently labeled actin filaments in the presence of 2 μM coflin, 1 μM coronin, 200 nM Aip1, 5 μM of actin monomer, and 2 mM ATP. Filaments shorten and disappear from the field of view. Bar, 3 μm. (B) Successive time-lapse images showing a single actin filament (f1) over time, along with kymographs drawn along the contours of representative filaments (f1–f4). The red lines on the image of f1 at t = 0 denote the contour on which the kymograph was drawn. Time is given on the x axis of the kymograph, whereas the position along the filament contour is given on the y axis. Mean integration time for a single image was 400 ms for f1–f3 and 16 ms for f4. Triangles denote endwise bursting (f1–f3); yellow triangles denote initial burst (f1–f3), red triangles denote successive proximal bursts (f1 and f2), and green triangle denotes a successive distal burst (f3). Same-side bursts occurred more frequently (78%) than opposite-side bursts (22%; P < 0.001, one-tailed z test). The square denotes internal disassembly event counted as a severing event (f3). Bar, 1 μm. (C) Histogram of filament burst size. The mean burst size was 260 subunits. (D) Histogram of waiting times between successive bursts (red), fit to a single exponential (black). Single exponential fit gave characteristic decay time of τ = 14 s.
microscopy (Fig. 1 A and Video 1, available at http://www.jcb.org/cgi/content/full/jcb.200801027/DC1).

In the presence of cofilin, coronin, and Aip1, filaments shortened and disappeared from the field of view within the first 100 s (Fig. 1, A and B; and Video 1). We tracked the locations of filament ends using kymographs drawn along the contours of individual actin filaments (Fig. 1, B and C, filaments f1–f4). Kymographs showed that filaments did not shrink smoothly from an end; instead, they disassembled in infrequent bursts, with filaments abruptly losing mass from an end (Fig. 1 B, triangles). Most disassembly events occurred at filament ends, but events were occasionally initiated in the middle of the filament; these were scored as severing events (Fig. 1 B, blue square; see Fig. 2 for a detailed comparison of this reaction to a cofilin-mediated severing reaction). In an attempt to resolve molecular events catalyzed by a three-component system to the cofilin-alone reaction, we scored disassembly events under both conditions and counted them as either bursting or severing events. For scoring purposes, a severing event was one in which both daughter filaments remained attached to the surface of the coverslip for at least two frames after breakage. Such an analysis revealed that, although filaments in the three-component system underwent occasional internal disassembly events that were scored as severing events (Fig. 1 B, f3, square), bursting from ends was the predominant mode of disassembly (Fig. 2 B). In contrast, severing was the predominant mode of disassembly catalyzed by cofilin alone (Fig. 2 B), which is consistent with previous studies (Maciver et al., 1991; Andrianantoandro and Pollard, 2006). Our cofilin-alone data reinforce our conclusion that bursting disassembly catalyzed by cofilin, coronin, and Aip1 is a novel mechanism of filament disassembly.

Actin filaments are structurally polarized, so we next tested whether the bursts of disassembly occur preferentially from a particular end of the filament. To separately probe disassembly near barbed ends and pointed ends, we imaged disassembly of filaments elongated from fragments of Limulus polyphemus, acrosomal processes, which are highly bundled arrays of filaments all oriented in the same direction (Bonder and Mooseker, 1983). Filaments grown off the barbed end of acrosomal bundles have free barbed ends but pointed ends that are connected to the acrosome. The reverse applies at the pointed end. We polymerized bundles of filaments off the barbed ends and pointed ends of L. polyphemus acrosomes using 5 μM of fluorescently labeled actin monomer and 2 mM ATP and then perfused them with cofilin, coronin, and Aip1, as well as unlabeled actin monomer and ATP. To minimize filament aging effects, filaments were perfused within 2 min after initiation of polymerization. Interestingly, filament bundles grown from either barbed ends (Fig. 3 A, b) or pointed ends (Fig. 3 A, p) of acrosomal processes disassembled at comparable rates (Fig. 3 A). Most of the polymer mass in the filament bundles disappeared within the first 2 min; the remainder, which manifest as a short
similar rates. is consistent with both ends undergoing bursting disassembly at nin, and Aip1 frequently gave rise to two new filament ends that with previous kinetic studies (k on all standard barbed ends (unpublished data), which is consistent for 10 s. This CapZ incubation was sufficient to completely cap conditions, and then perfused them with 3 μM Alexa 488 CapZ somal fragments, treated them under different depolymerizing protein that caps standard filament barbed ends with fast kinetics (Caldwell et al., 1989; Schafer et al., 1996). When filament bundles were partially disassembled with coflin, coronin, and Aip1 before CapZ incubation, their fluorescence also tapered due to disassembly (Fig. 4 C, left). However, unlike coflin-treated filament bundles, these bundles recruited almost no CapZ (Fig. 4 C, middle). The CapZ/filament ratio for bundles treated with coflin, coronin, and Aip1 was much lower than that for untreated bundles (Fig. 4 A and B, right; 1-tailed t test, t = 1.6, degrees of freedom [df] = 14, P > 0.01). We conclude that filaments severed by coflin retain the ability to bind CapZ at their barbed ends, as expected.

When filament bundles were partially disassembled with coflin, coronin, and Aip1 before CapZ incubation, their not only at their tips but also along their lengths (Fig. 4 B, middle), which is consistent with binding of CapZ to barbed ends of severed filaments. To quantify the fraction of capped filaments before and after coflin treatment, we measured for individual filament bundles the CapZ/filament ratio, which reports on the proportion of capped filaments (see Materials and methods). This analysis showed that coflin-treated filament bundles showed no significant decrease in the fraction of capped filaments compared with untreated bundles (Fig. 4, A and B, right; 1-tailed t test, t = 11.1, df = 22, P < 0.01) and not significantly different from zero (Fig. 4 C, right; t test, t = 2.3, df = 15, P < 0.01). A similar decrease in CapZ recruitment was also seen when CapZ was incubated concurrently with coflin, coronin, and Aip1 (unpublished data). These results suggest that, unlike a pure severing reaction, bursting disassembly by the three-component system generates as reaction intermediates barbed ends that are not recognized by CapZ. To determine whether these barbed ends were stable in buffer, we partially disassembled filament bundles with coflin, coronin, and Aip1, then incubated them in buffer for 10 s before incubation with CapZ. These filament bundles recruited significantly more CapZ than bundles treated directly with CapZ (Fig. 4, C and D; 1-tailed t test, t = 4.8, df = 22, P < 10⁻⁴), which suggests that, in the absence of disassembly factors, nonstandard barbed ends rapidly revert to a standard CapZ-recognizable form. Consistent with this interpretation, others have observed that coflin and Aip1 do not inhibit filament elongation from barbed ends when washed out (Ono et al., 2004). These data suggest that bursting disassembly generates unstable filament intermediates with nonstandard barbed ends not recognized by CapZ.
Barbed end-capping agents inhibit actin disassembly in vitro

Our experiments with fluorescent CapZ argue against a disassembly mechanism where depolymerization factors form a stable cap with filament barbed ends to prevent monomer addition and filament annealing. They instead suggest that bursting disassembly may involve active disruption of filament internal structure, leading to ragged barbed ends that cannot recruit CapZ. To test these ideas further, we imaged single filaments after perfusion with coflin, coronin, and actin monomer with or without Aip1, and in the presence of CapZ or cytochalasin D (CytoD), a well-characterized barbed end–capping drug.

Omission of Aip1 from the three-component system stopped disassembly (Fig. 5 A, green) and greatly reduced the frequency of both bursts and scored severing events (Fig. 5 B, green). Omission of either coflin or coronin had the same effect (Fig. S2, available at http://www.jcb.org/cgi/content/full/jcb.200801027/DC1), which is consistent with a requirement for all three factors for rapid monomer-insensitive disassembly in L. monocytogenes comet tails (Brieher et al., 2006). CapZ could not substitute for Aip1 in the three-component reaction (Fig. 5, A and B, yellow), which suggests a role for Aip1 in disassembly that is distinct from barbed-end capping. When added to the full three-component system, both CytoD and CapZ strongly inhibited filament disassembly (Fig. 5 A, black and blue), blocking both endwise bursting and internal severing modes of filament disassembly (Fig. 5 B, black and blue). To separately test the effect of CytoD and CapZ on disassembly from filament barbed ends and pointed ends, we imaged acrosomal filament bundles in the presence of the full three-component system and either CytoD or CapZ (Fig. 5 C). Remarkably, disassembly of filaments grown off barbed ends and pointed ends was inhibited with equal efficacy, which suggests that filament segments near both ends disassemble through the same CytoD/CapZ–sensitive mechanism (Fig. 5 C). CytoD inhibited filament bundle disassembly with an IC_{50} of 90 nM (measured using total polymer mass from all filament bundles in a field of view), which is similar to its IC_{50} for inhibition of polymerization off acrosomal fragments (30 nM; Fig. 5 D). In contrast, although CapZ also inhibited polymerization off acrosomal fragments with a low IC_{50} (30 nM; Fig. 5 E), its inhibitory effects on disassembly did not saturate at 10 μM (Fig. 5 E). The difference in IC_{50}s between CytoD– and CapZ-mediated inhibition of disassembly suggests differences in the detailed mechanism between these two barbed end–capping agents. For example, barbed ends generated during disassembly may have a conformation that is recognized by CytoD but not by CapZ. Regardless of the detailed mechanism, their common inhibitory effects suggest an important role for barbed end–capping agents in controlling filament stability in the three-component system, even when disassembly occurs by bursting from the pointed end. In contrast, CytoD did not inhibit filament severing in high concentrations of coflin alone (Fig. 5 F), which is consistent with others’ results (Ono et al., 2004).

Barbed end-capping agents inhibit actin disassembly in cells

To determine whether physiological actin disassembly also occurs by a mechanism that is inhibited by barbed end–capping agents, we treated mammalian tissue culture cells with CytoD and measured disassembly by several different assays. As a control, we used latrunculin B (LatB), a drug that also inhibits polymerization but does so by sequestering a free monomer. We first used a simple bulk assay for total cellular F-actin. Human HeLa S3 cells grown in suspension were treated with either 10 μM CytoD or LatB, fixed at various time points after treatment, and then stained with TRITC-phalloidin to quantify bulk F-actin levels. Before drug treatment, F-actin was present throughout the entire cell and was enriched at the cell cortex (Fig. 6 A, top). Treatment of the cells with CytoD did not reduce the amount of F-actin per cell (Fig. 6 A, middle; and Fig. 6 B, blue). The TRITC-phalloidin signal remained high for >30 min after CytoD treatment. The drug was active in these cells as it induced extensive blebs after a few minutes (unpublished data). In contrast, treatment of cells with LatB led to a rapid drop in the amount of F-actin per cell (Fig. 6 A, bottom; and Fig. 6 B, red).

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kabiramide C (KabC), a barbed end–capping drug structurally unrelated to CytoD (Tanaka et al., 2003). We recorded time-lapse fluorescence videos of *L. monocytogenes* actin comet tails inside mammalian tissue culture cells expressing GFP-actin. Before drug addition, actin comet tail assembly propelled *L. monocytogenes* forward in the cytoplasm. Comet tail actin disassembled within a minute after assembly (Fig. 6C, control; and Fig. 6D, green; Theriot et al., 1992), causing a decrease in filament density away from the bacterium. Both CytoD and KabC stopped comet tail assembly and *L. monocytogenes* motility within a few seconds after addition (Fig. 6C), which is consistent with the drugs capping barbed ends to inhibit actin polymerization. Both CytoD and KabC also inhibited disassembly of comet tails with the purified protein system. The TRITC-phalloidin signal dropped by nearly threefold over the first 100 s, which implies a timescale of disassembly consistent with fluorescence recovery after photobleaching studies of cortical actin turnover (Murthy and Wadsworth, 2005) and our data with the purified protein system. The differential responses of cellular F-actin levels to CytoD and LatB treatment provides evidence that CytoD also actively inhibits actin filament disassembly in vivo, as it also does in our purified protein system in vitro.

To probe disassembly on a well-characterized actin array in vivo, we imaged actin comet tails formed by the intracellular pathogen *L. monocytogenes* (Sanger et al., 1992; Theriot et al., 1992) while perturbing cells with CytoD or LatB. To control for secondary effects of CytoD, we also used in this assay cabiramide C (KabC), a barbed end–capping drug structurally unrelated to CytoD (Tanaka et al., 2003). We recorded time-lapse fluorescence videos of *L. monocytogenes* actin comet tails inside mammalian tissue culture cells expressing GFP-actin. Before drug addition, actin comet tail assembly propelled *L. monocytogenes* forward in the cytoplasm. Comet tail actin disassembled within a minute after assembly (Fig. 6C, control; and Fig. 6D, green; Theriot et al., 1992), causing a decrease in filament density away from the bacterium. Both CytoD and KabC stopped comet tail assembly and *L. monocytogenes* motility within a few seconds after addition (Fig. 6C), which is consistent with the drugs capping barbed ends to inhibit actin polymerization. Both CytoD and KabC also inhibited disassembly of comet tails with the purified protein system. The TRITC-phalloidin signal dropped by nearly threefold over the first 100 s, which implies a timescale of disassembly consistent with fluorescence recovery after photobleaching studies of cortical actin turnover (Murthy and Wadsworth, 2005) and our data with the purified protein system. The differential responses of cellular F-actin levels to CytoD and LatB treatment provides evidence that CytoD also actively inhibits actin filament disassembly in vivo, as it also does in our purified protein system in vitro.

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Lamellipodia and other dynamic cellular actin assemblies contain dense dendritic arrays resembling those seen in L. monocytogenes (Sechi et al., 1997; Cameron et al., 2001). To test if these arrays show similar sensitivities to CytoD versus LatB with respect to disassembly, we imaged the ruffling edge of BSC-1 cells stably expressing a fusion of actin to monomeric RFP and photoactivatable GFP (mRFP-PAGFP-actin; Fig. S4, available at http://www.jcb.org/cgi/content/full/jcb.200801027/DC1). PAGFP allows localized illumination of specific actin subpopulations, and the in-tandem mRFP permits simultaneous viewing of the bulk actin population. Like GFP-actin, this probe localized to actin-rich cellular structures like the ruffling cell edge and stress fibers (Fig. 7 A). During the experiments, we imaged both red and green fluorescence and tails, which showed no visible decrease in fluorescence for up to a minute after drug addition (Fig. 6, C and D, blue; and Videos 3 and 4, available at http://www.jcb.org/cgi/content/full/jcb.200801027/DC1). Cell rounding induced by both drugs hindered longer observation; however, tails were still visible in rounded-up cells up to 10 min after CytoD addition, which implies a reduction in the disassembly rate by at least 20-fold. Although LatB also rapidly stopped tail assembly and L. monocytogenes movement, it did not inhibit disassembly. Comet tails disappeared after drug addition at a rate similar to that in untreated cells (Fig. 6, C and D, red; and Video 5). Others have previously noted that comet tails are stable in CytoD-treated cells (Sanger et al., 1992) but did not attribute this observation to inhibition of disassembly.
Discussion

This study aimed to elucidate the mechanism of rapid monomer-insensitive actin disassembly by the three-component system consisting of coflin, coronin, and Aip1. By directly imaging filament dynamics before and after drug treatment both in bulk and in photoactivated subpopulations, we found that this three-component system disassembles filaments in abrupt bursts, where 100-subunit-long segments near both filament ends are lost very rapidly. Bursting disassembly generated filament barbed ends not recognized by CapZ, a protein that binds tightly to standard barbed ends. When added to the three-component system, CapZ and CytoD inhibited bursting disassembly, and did so from both ends with equal efficacy. CytoD and KabC, a structurally unrelated barbed-end capper, also inhibited actin disassembly in mammalian cells, which suggests that the three-component system reconstitutes a prevalent physiological disassembly pathway.

Mechanism of actin filament bursting disassembly

What is the mechanism underlying bursting filament disassembly? We propose the following model (Fig. 8 A): during a burst of disassembly, cooperative weakening of lateral interactions between the two strands of the actin filament causes the strands to transiently separate near an end (Fig. 8 A, left). Coflin has been shown to disrupt lateral contacts between subunits in adjacent strands (Bobkov et al., 2004) as well as induce splaying or unwinding of strands observed by electron microscopy (Bremer et al., 1991; McGough et al., 1997). Aip1, which is...
believed to bind filaments at barbed ends and along their lengths (Okada et al., 2002), may also catalyze cofillin-induced strand separation. Consistent with this idea, Aip1 interacts with the F-actin subunit near surfaces that form lateral interactions with adjacent filament strands in the Holmes model (Holmes et al., 1990; Amberg et al., 1995; Rodal et al., 1999; Clark and Amberg, 2007). As separated strands lack stabilizing lateral interactions, they are highly unstable and rapidly disassemble into constituent subunits. Rapid disassembly of separated filament strands is plausible, as single-stranded polymers are not limited by dissociation of individual subunits from ends (Oosawa and Asakura, 1975).

This burst leaves behind a nonstandard filament end (Fig. 4), which can then initiate another burst at a significantly faster rate, giving rise to a successive series of bursts from the same filament end (Fig. 1 B, f1 and f2). Occasionally, strand separation also occurs in the middle of the filament, giving rise to internal bursts of disassembly scored as severing events in our assay (Fig. 1 B, f3; and Fig. 8 A, right). However, most disassembly occurs as bursts from either filament end (Fig. 2 B), as strands separate more easily near ends.

An alternative interpretation of our observations is that bursting disassembly simply involves filament severing, where a spatially localized disruption of the filament causes it to break at the site of disruption (Fig. 8 B). Both severed daughter filaments are completely intact but one daughter filament immediately diffuses away and cannot be detected. However, we favor cooperative strand separation over severing for several reasons: (1) strand separation accounts for preferential bursting from an end that has already undergone a burst (Fig. 1), whereas severing does not; (2) strand separation accounts for preferential initiation of disassembly near filament ends (Fig. 1), whereas severing does not (moreover, the classical severing we observed using cofillin alone was not biased toward ends [Fig. 2], which argues that nothing in our experimental setup, such as filament immobilization, is causing an artificial bias toward ends); (3) we were able to score severing by single filament imaging (Fig. 2), which argues that we would have seen severing if it contributed to bursting; (4) strand separation accounts for disassembly intermediates with lowered affinity for CapZ (Fig. 4); it is also possible that barbed ends are structurally intact but capped by Aip1 after a burst and thus unable to bind CapZ; however, this putative Aip1–barbed end interaction would be quite unstable (τ < 10 s; Fig. 4 D); and (5) strand separation plausibly accounts for inhibition of disassembly by capping factors (Figs. 5–7), whereas severing does not (Fig. 5 F). In our view, severing better describes the behavior of filaments treated with high concentrations of cofillin alone (Fig. 2). Future efforts to distinguish between these alternative models will require electron microscopy of intermediates or development of spectroscopic methods that report on filament internal structure.

The mechanism of inhibition of disassembly by barbed end–cing agents presents a paradox we have yet to fully resolve. Mutual antagonism between barbed end capping and bursting disassembly suggests a possible means of inhibition. Bursting disassembly may inhibit binding of CytoD/CapZ by separating filament strands; conversely, CytoD/CapZ may inhibit bursting disassembly by keeping strands together near barbed ends. However, filaments do not disassemble preferentially from barbed ends but instead disassemble near both ends with similar kinetics by a similar capping-sensitive mechanism. We considered several possible explanations. One possibility is that barbed end factors control the stability of the whole filament by binding only to the free barbed end. However, the effect of barbed-end binding would have to propagate through the L. polyphemus acrosomal bundle, which itself is highly stabilized. Alternatively, barbed end factors may act in solution as opposed to on the polymer to inhibit disassembly. For instance, CytoD may interact with an actin monomer to perturb the activity of the purified system (Goddette and Frieden, 1986). However, we failed to detect any possible indirect effects of CytoD in solution (Fig. S3, available at http://www.jcb.org/cgi/content/full/jcb.200801027/DC1). A third possibility is that barbed end factors may bind along the filament length to inhibit strand separation. Consistent with this idea, in vitro studies point to low-affinity cytochalasin B binding sites along the filament length (Hartwig and Stossel, 1979).

**Implications for actin dynamics in cells**

What are the implications of this new disassembly reaction for dynamic organization and morphogenesis of actin assemblies in cells, assuming that it is physiologically relevant? Actin cytoskeletal networks in cells must disassemble through a mechanism that is both fast and insensitive to monomers. Monomer insensitivity is a basic thermodynamic requirement for non-equilibrium polymer turnover in the cell, and speed is important to allow fast remodeling of the actin cytoskeleton. The three-component reaction may achieve speed by disassembling entire filaments in cells in a single burst, as many actin filaments do not exceed hundreds of subunits in length in cells (Sechi et al., 1997; Cameron et al., 2001). Moreover, the three-component reaction may achieve monomer insensitivity by generating non-standard filament ends (Fig. 4 C) that cannot readily add subunits, which is akin to the low affinity of shrinking microtubule (+) ends for tubulin. We have not visualized these non-standard ends and speculate that the two filament strands may be unequal in length or partially separated (Fig. 8 A).

In contrast to bursting disassembly by the three-component system, pure severing by cofillin alone nucleates assembly by generating filament ends that can grow (Ichetovkin et al., 2002; Andrrianantoandro and Pollard, 2006). The opposite effects of these two reactions may help resolve the paradoxical role of cofillin in cells, where it promotes disassembly in some cases.
(Kiuchi et al., 2007; Okreglak and Drubin, 2007) and assembly in others (Ichetovkin et al., 2002; Ghosh et al., 2004). Local activity of Aip1 and/or coronin may be important in determining whether coflin catalyzes assembly or disassembly in cells.

The inhibitory effects of barbed-end cappers in our experiments suggest a role for barbed end–associated factors such as CapZ, formin, or gelsolin in modulating not only filament assembly but also disassembly. Conversely, bursting disassembly may antagonize barbed end–capping agents in cells, possibly accounting for the coflin-dependent fast dissociation of CapZ from dynamic actin assemblies in cells (Miyoshi et al., 2006). Similarly, the side-binding factors (Siripala and Welch, 2007) may also modulate filament stability in cells. We note that the rate of bursting disassembly in our in vitro assays decreased when filaments were immobilized with N-ethylmaleimide–inactivated myosin instead of filamin (unpublished data). It will be a challenge to examine how filaments disassemble in the full repertoire of factors that interact with actin filaments in cells.

Differential drug effects

We finally note that CytoD and LatB are widely used research tools and are often used interchangeably. Previous studies demonstrated, but did not explain, differences between these drugs with respect to actin polymerization (Higashida et al., 2004), gross morphology (Spector et al., 1989), cell mechanical properties (Wakatsuki et al., 2001), and other cellular processes (Forer and Pickett-Heaps, 1998; Morales et al., 2000; Omata et al., 2000). The dramatic difference in their effects on disassembly (Figs. 6 and 7) has not been recognized before and may explain these different effects on cell physiology. In the future, differential effects of these two drugs can be used to test the importance of disassembly in different actin-dependent cellular processes, a question that has largely been overlooked.

Materials and methods

Protein purification

Actin was prepared from rabbit skeletal muscle according to the method of Pardee and Spudich (1982). The F-actin was then depolymerized by dialysis into G buffer (2 mM Tris-Cl, 0.2 mM CaCl2, 0.2 mM ATP, 0.1% β-mercaptoethanol, and 0.005% NaNO3, pH 8.0) and further purified by an additional cycle of polymerization/dem polymerization as described by Pardee and Spudich (1982). Actin was then repolymerized and labeled on lysines using approximately stoichiometric amounts of Alexa 488 and Alexa 647 N-hydroxysuccinimide–ester (Invitrogen) according to the procedure of Kellogg et al. (1988). The labeled actin was subject to two additional cycles of polymerization and depolymerization and then frozen in aliquots at −80°C. Cofflin, coronin, and Aip1 were purified as described by Brieher et al. (2006).

Construction of perfusion chambers

Perfusion chambers were assembled using glass cover slips (48366–227; VWR international), glass slides (3050; Gold Seal), and thin parafilm strips. Glass cover slips were cleaned for imaging by sonication for 30 min in 1% Micro-90 detergent (99999–01; Cole-Parmer), 60 min in 2 M KOH, 30 min in acetone, and 30 min in ethanol. During each solution change, cover slips were rinsed extensively using 10 or more changes of distilled water. Cleaned cover slips were then stored in ethanol until the day of the experiment.

To assemble the perfusion chamber, two parafilm strips were placed on the surface of the glass slide. The cover slips were then removed from ethanol, dried using filtered air, and pressed onto the parafilm strips to create a fluid channel. The chamber was then baked at 85°C for 20 s to create a seal between the parafilm and the glass. Assembled chambers were stored in a box and used within 2 d.

Imaging of single actin filaments

Alexa 488– or Alexa 647–labeled actin was treated with 1 mM EGTA and 0.2 mM MgCl2 on ice for 5 min to convert calcium-ATP-actin to magnesium-ATP-actin. To remove residual filaments, the actin was then diluted in G buffer (0.2 mM CaCl2, 0.2 mM ATP, and 2 mM Tris-Cl, pH 8), left on ice for 20 min, and centrifuged at 436,000 g with a k-factor of 7 for 20 min. The supernatant, which contained actin monomer, was retained and kept on ice. Alexa 488, an actin cross-linking protein, was diluted into assay buffer (50 mM KCl, 2 mM MgCl2, 2 mM ATP, and 100 mM K-Hepes, pH 7.8) to a final concentration of 10–50 μg/ml and adsorbed onto coverslips of perfusion chambers using a 10-min incubation. Chambers were then washed with blocking solution (5 mg/ml casein, 0.2% Tween 20, and 0.1% Pluronic F-127 in assay buffer) for 5 min. Immediately before polymerization of Alexa 488 and Alexa 647, actin monomer was mixed to a final concentration of 10 μM, 30% Alexa 647–labeled and 5% Alexa 488–labeled. 10x polymerization buffer (500 mM KCl, 20 mM MgCl2, 20 mM ATP, and 1 M K-Hepes, pH 7.8) was then added, and the mixture was allowed to polymerize in the perfusion chamber for 30–60 s. The chamber was then perfused with 6–7 chamber volumes of assay buffer supplemented with an oxygen scavenging system (4.5 mg/ml glucose, 0.2 mg/ml glucose oxidase, and 35 μg/ml catalase) and 10% blocking solution for imaging. Actin filaments in the perfusion chamber were imaged immediately at room temperature using an upright wide-field epifluorescence microscope (E90i) with a 60x 1.4 NA oil objective (both from Nikon). To distinguish newly polymerized filaments from filaments already present before salt addition, an image of filaments in the field of view was first taken in both the far-red channel (Alexa 647) and the green channel (Alexa 488). The disassembly process was diluted into imaging buffer and then perfused into the chamber, and a streaming video of filament disassembly was taken in the far-red channel using either an intensifying charge-coupled device (CCD) camera (ORCA; Hamamatsu) or an electron microscopy CCD camera (iKon; Andor Technology PLC) with image acquisition software (Metamorph 6.0; Invitrogen).

Segmentation of single filaments by automated image analysis

Single actin filaments in the time-lapse images were segmented using a custom image analysis algorithm written in MATLAB (Mathworks). For each image in a time-lapse sequence, background correction was performed by subtracting a grayscale morphological opening of the image from its parent. The contrast of the image was then adjusted. The low contrast value was set to: \[ I_{\text{low}} = I_{\text{mode}} - n \times I_{\text{width}}, \] where \( I_{\text{mode}} \) is the grayscale value at the peak of the intensity histogram, \( I_{\text{width}} \) is the width of the peak of the intensity histogram, and \( n \) is an adjustable parameter. The high contrast value was set to: \[ I_{\text{high}} = I_{\text{mode}} + n \times I_{\text{max}} - I_{\text{mode}}, \] where \( I_{\text{max}} \) is the highest grayscale value in the image and \( 0 < n < 1 \) is the degree of saturation. The image was then segmented and the segmentation was iteratively refined using the following procedure: the contrast-adjusted image was passed through a Canny edge detector, which generated a binary image with closed contours around the actin filaments. Single-pixel gaps in the contours in the binary images were bridged, and the regions enclosed in the contours were filled. Unfilled open contours were removed by morphological opening, and the filled objects were thinned to lines. The shape of each object was automatically inspected, and only objects without branch points (corresponding to linear segments) were retained. The grayscale values from pixels corresponding to objects in the segmentation were then averaged to obtain mean filament intensity \( I_0 \) from this estimate, and the new high contrast value of the image was then set to: \[ I_{\text{new}} = I_{\text{high}} + n \times I_0 - I_{\text{high}}. \] The procedure was then repeated until \( I_0 \) converged to a constant value. A typical image would require approximately three or four iterations before convergence. This iterative procedure achieves automated segmentation robust to fluctuations in fluorescence intensity, variations in filament density, and the presence of bright outliers. It also does not require input parameters that explicitly depend on microscope acquisition settings.

Once segmentation was achieved, lengths of individual linear contours were estimated by the number of pixels occupied by the contours in the binary image. The lengths from all linear objects were then averaged to give the mean filament length.

Imaging of actin filament bundles grown off L. polyphemus acrosomal processes

L. polyphemus acrosomal actin bundles (a gift from G. Weller and P. Matsudaira, Massachusetts Institute of Technology, Cambridge, MA) were washed with blocking solution (5 mg/ml casein, 0.2% Tween 20,
and 0.1% Pluronic F-127 in assay buffer) for 5 min. Filaments were elongated off acrosomal bundles by incubation with 3 μM Alexa 647-labeled actin monomer in assay buffer for 2 min and then washed. For time-lapse imaging experiments, filament bundles were perfused with disassembly factors and 100 μg/ml filamin, which held bundles together for imaging. For experiments with Alexa 488 CapZ, filament bundles were treated under different depolymerizing conditions, incubated with 3 μM Alexa 488 CapZ for 10 s, washed again, bundled with 200 μg/ml filamin, and then imaged. Unless otherwise stated, all perfusion steps were performed using assay buffer supplemented with oxygen scavenging system and 10% blocking solution. Imaging was performed using the fluorescence microscope described for single filaments.

Measurement of polymer mass decay in acrosome filament bundles
To quantify polymer mass in fluorescent filament bundles, we wrote an automated image analysis routine in MATLAB (Mathworks). The routine measures integrated intensity of a bright object over a dark background. The grayscale value for the background in each image was estimated by finding the mode of the image histogram, assuming that the objects of interest were small in the field of view. The pixel intensities were background-subtracted using the obtained value and then summed to obtain the integrated fluorescence intensity. This procedure was repeated for all images in the time-lapse sequence. The polymer mass decay curves of all filament bundles were obtained by running this routine for the entire field of view (Fig. S1 A, left). Polymer mass decays for long bundles and short bundles were obtained by running the routine on the appropriate regions of interest (Fig. S1 A, middle and right). The polymer mass exhibited a rapidly decaying component followed by a decay with significantly slower kinetics. We then subtracted from the polymer mass decay curve the slow-varying component, which was obtained by fitting the last segment of the curve to a straight line and extrapolating to time 0 (Fig. S1 B, red and orange lines). The resultant decay curve was then normalized to unity at time 0, and the characteristic rate of polymer mass decay was then obtained by fitting the first few points of the normalized decay curve to a straight line (Fig. S1 C).

For experiments involving Alexa 488 CapZ (capping protein was a generous gift of A. Lebehn, Harvard Medical School, Boston, MA), the integrated CapZ fluorescence in a single filament bundle was measured in a region of interest surrounding the filament bundle. Background was measured in a neighboring region of interest where no filament bundles were present. The number of actin filaments in the bundle was estimated by taking the maximal fluorescence intensity along the filament bundle length. The integrated CapZ fluorescence was divided by estimated actin filament number to obtain the CapZ/actin filament ratio (Fig. 3 B), which measures the proportion of CapZ-bound filaments in the filament bundle.

Drug treatment of suspension cells
HeLa S3 cells were cultured in suspension in shaking Fernbach flasks in CO2-independent medium (Invitrogen) plus 10% fetal calf serum. 10 ml of log phase culture (~10^6 cells/ml) were removed, disaggregated by passing twice through a 27-gauge needle, and kept in a water bath at 37°C with occasional agitation. At time 0, CytoD or LatB was added from a 2,000x stock in DMSO to a final concentration of 10 μM. 0.5 ml aliquots of cells were removed at each time point and fixed by dilution into 1 ml of PBS (1.55 mM NaCl, 1 mM potassium phosphate, 3 mM sodium phosphate, 1 mM MgCl2, and 1 mM CaCl2, pH 7.4) + 2% formaldehyde. After 10–20 min at 25°C, the fixed cells were collected by centrifugation (1,000 g for 5 min) and resuspended in 100 μl of PBS + 0.1% Triton X-100, 1 μM TRITC-phalloidin + 1 μM Hoechst dye. This was cooled for 30 min at 25°C with occasional agitation, the cells were diluted with 1 ml of PBS + 0.1% Triton X-100, collected by centrifugation, resuspended in 50 μl of PBS + 60% glycerol, and mounted between a slide and coverslip. Random fields were imaged with a 10x objective. Integrated intensity of the TRITC-phalloidin signal in each field of view was measured and used in the image analysis routine described for acrosome filament bundles. The total intensity in the field of view was then divided by the number of cells present to obtain the integrated intensity per cell.

Infection of tissue culture cells with L. monocytogenes
Before infection, BSC-1 cells were cultured in DME media on polylysine-coated glass coverslips. Cells were then infected with an adenovirus expressing GFP-actin and incubated overnight. These GFP-actin-expressing cells were then infected with L. monocytogenes for 3 h, washed with DME containing 50 μg/ml gentamicin, and incubated at 37°C for up to 24 h before live imaging.

Live imaging of L. monocytogenes comet tails in tissue culture cells
Imaging was performed using a 100x 1.4 NA oil objective on an epifluorescence microscope (TE300, Nikon). BSC-1 cells expressing GFP-actin were imaged in DME containing 50 μg/ml gentamicin and 20 mM Hepes at pH 7.4 and were maintained with a cooled-CCD camera (ORCA-ER; Hamamatsu) using image acquisition software (MetaMorph; MDS Analytical Technologies). For experiments involving drug treatment, the imaging media was exchanged for media containing the drug of interest during the course of image acquisition. Drug concentrations used: 6 μM LatB, 5 μM CytoD, and 0.5 μM KacB (provided by G. Marriot, University of Wisconsin, Madison, WI). In all cases, the concentration of DMSO was <0.5%.

Measurement of polymer mass decay in L. monocytogenes comet tails in cells
Polymer mass in L. monocytogenes actin comet tails were obtained using the same image processing routine detailed for analysis of L. polyphemus acrosomal filament bundles. Polymer mass decay curves were obtained in regions of interest around the part of the comet tail that had polymerized immediately before drug addition. Decay curves from multiple cells were then normalized and averaged together.

Preparation of mRFP-PAGFP-actin in a retroviral vector
The mRFP-PAGFP-actin plasmid was based on an EGFP-tagged human β-actin from Clontech Laboratories, Inc. First, EGFP was excised using an Nhel–BglII digest and replaced by mRFP (a gift of R. Tsien, University of California, San Diego, La Jolla, CA). Second, PAGFP (a gift from R. Wedlich-Soldner, Max Planck Institute, Martinsried, Germany) was PCR amplified and ligated into the mRFP-actin between XhoI and BglII. In addition, to enhance flexibility of the linking regions, glycine residues were inserted between mRFP and PAGFP (one added) and between PAGFP and actin (three added; Fig. S4). The plasmid sequence was verified by sequencing. For reliable expression inside cells, a retrovirus encoding mRFP-PAGFP-actin was generated. mRFP-PAGFP-actin was PCRed and ligated into pLNCX2 (Clontech Laboratories, Inc.) between HindIII and NotI, and this construct was transfected into a GIPS-293 packaging cell line for production of retrovirus (Ory et al., 1996).

Preparation of a cell line stably expressing mRFP-PAGFP actin
BSC-1 cells were infected with DME media containing retrovirus with the mRFP-PAGFP construct and a G418 selectable marker. 3 d later, cells were split into media containing 500 μg/ml G418 (Invitrogen). Upon confluence, cells were then reseeded in selective media at very low density (1:1,000, 1:5,000, and 1:10,000). 3 wk later, individual colonies were picked and regrown to confluence. Individual clones were then selected based on cell vitality and morphology, as well as actin morphology and mRFP-PAGFP-actin fluorescence intensity, and cultured on glass coverslips for imaging.

Imaging of photoactivated fluorescent actin in tissue culture cells
Imaging was performed using a 100x 1.4 NA oil objective on an inverted microscope (TE2000U, Nikon) with an Ultraview spinning disk confocal unit (PerkinElmer). BSC-1 cells expressing PAGFP-mRFP-actin were imaged in CO2-independent medium (18045-088; Invitrogen) supplemented with 5-glutamine and were maintained at 37°C by a heated metal stage. Cells were excited with 488- or 568-nm light from an argon-krypton laser (Melles Griot), and images were taken with a cooled-CCD camera (ORCA-ER) using image acquisition software (MetaMorph). During the time lapse, media containing the drug was applied to the cells, and the PAGFP-mRFP-actin was photoactivated using a nitrogen pulse laser tuned to 405 nm (Photonic Instruments, Inc.). Both LatB and CytoD were diluted in CO2-independent media to a final concentration of 5 μM in 0.5% DMSO.

Online supplemental materials
Fig. S1 shows decay rates of L. polyphemus actin bundles with exposed barbed versus pointed ends. Fig. S2 shows that monomer-insensitive disassembly of single filaments requires cofillin, coronin, and Aip1. Fig. S3 tests the effects of CytoD acting in solution on actin disassembly. Fig. S4 is a diagram of the mRFP-PAGFP-actin construct used for the photoactivation experiments in cells. Video 1 shows disassembly of single actin filaments in the presence of collyrin, Aip1, and G-actin. Video 2...
shows disassembly of actin grown off L. polyphemus bundles under the same conditions. Videos 3, 4, and 5 show decay of L. monocytogenes actin comets in the presence of CytoD, KabC, or LatB, respectively. Video 6 shows actin turnover in ruffling BSC-1 cells as a function of CytoD. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.200801027.DC1.

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References


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