



# Inhibition of Patterned Cell Shape Change and Cell Invasion by Discs Large During *Drosophila* Oogenesis

## Citation

Goode, Scott, Norbert Perrimon. "Inhibition of Patterned Cell Shape Change and Cell Invasion by Discs Large During *Drosophila* Oogenesis." *Genes & Development* 11, no. 19 (1997): 2532-2544. DOI: 10.1101/gad.11.19.2532

## Published version

<https://doi.org/10.1101/gad.11.19.2532>

## Link

<https://nrs.harvard.edu/URN-3:HUL.INSTREPOS:37372474>

## Terms of use

This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material (LAA), as set forth at

<https://harvardwiki.atlassian.net/wiki/external/NGY5NDE4ZjgzNTc5NDQzMGIzZWZhMGFIOWI2M2EwYTg>

## Accessibility

<https://accessibility.huit.harvard.edu/digital-accessibility-policy>

## Share Your Story

The Harvard community has made this article openly available.

Please share how this access benefits you. [Submit a story](#)

# Inhibition of patterned cell shape change and cell invasion by Discs large during *Drosophila* oogenesis

Scott Goode<sup>1</sup> and Norbert Perrimon<sup>2</sup>

Department of Genetics,<sup>2</sup> Howard Hughes Medical Institute, Harvard Medical School, Boston, Massachusetts 02115 USA

***Drosophila Discs large (Dlg)* is a tumor suppressor gene whose loss in epithelial tissues causes disrupted cell polarity and increased cell proliferation. A human Dlg homolog, hDlg, has been implicated in tumorigenic processes via its association with the product of the Adenomatous Polyposis Coli (APC) gene. We show for the first time that *Drosophila* Dlg is required to block cell invasion. Loss of *dlg* activity during oogenesis causes follicle cells to change shape and invade in a pattern similar to border cells, a small population of cells that break from the post-mitotic follicular epithelium during wild-type oogenesis, yet *dlg* mutant cells have not adopted a border cell fate. Both functional and morphological evidence indicates that cooperation between germ cell and follicle cell Dlg, probably mediated by Dlg PDZ domains, is crucial for regulating cell mixing, suggesting a novel developmental mechanism and mode of action for the Dlg family of molecules. These findings suggest that Dlg does not simply inhibit individual cell behaviors during oogenesis, but rather acts in a developmental pathway essential for blocking cell proliferation and migration in a spatio-temporally defined manner. A model for Dlg action in blocking cell invasion is presented.**

[Key Words: Cell invasion; cell proliferation; cell shape; Discs large; membrane-associated guanylate kinases (MAGuKs); PDZ domain]

Received June 12, 1997; revised version accepted August 5, 1997.

Adjoining tissues must regulate their barriers during development and adult life. What factors determine whether cells remain in one place, retaining their associations with their neighbors, or dissociate and move elsewhere? If cells move, what determines where they go, where and when they stop, and whether or not they associate with like or unlike cells? These are important questions in understanding tissue morphogenesis, dynamic physiological processes, and pathologies such as metastasis (Hynes and Lander 1992; Gumbiner 1996).

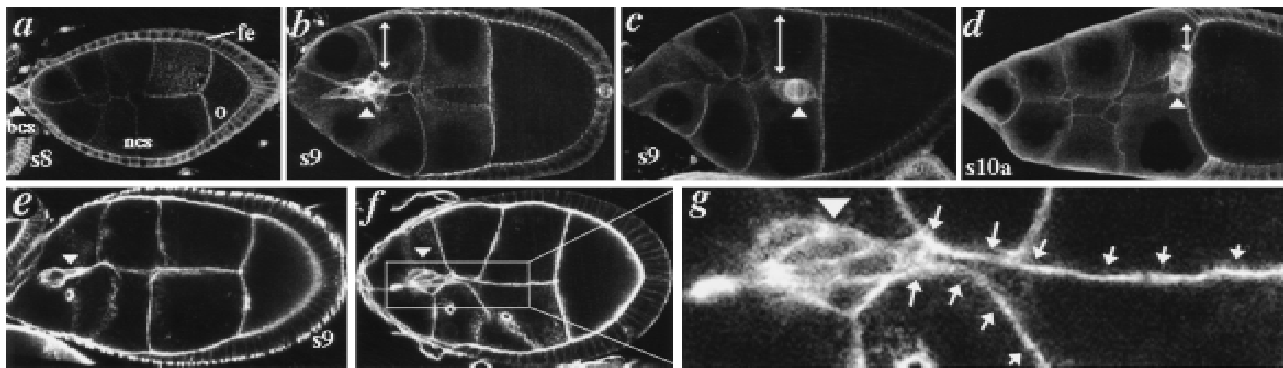
Maintenance of homotypic adhesion plays a crucial role in ensuring that cells do not mix. Blocking E-cadherin function turns cultured cells from noninvasive to invasive, whereas expressing E-cadherin in cancer cells reverts their tumorigenic phenotype (Takeichi 1993; Birchmeier and Behrens 1994). Physiological invasions and pathological metastasis also require adhesion of invasive cells to foreign matrices and heterologous cells (Mareel et al. 1991). What cellular mechanisms underpin the transfer of adhesion from sites of contact between homotypic cells to sites of contact between heterotypic cells?

An outstanding cell biological and genetic system for analyzing the problem of cell mixing and migration is

the movement of a small cluster of post-mitotic follicular cells, called border cells (BCs), between germ cells to the *Drosophila* oocyte (for review, see Montell 1994; described in Fig. 1). Several molecules have been identified as crucial players in regulating BC migration. The *slow border cells (slbo)* gene is specifically expressed in BCs and encodes the *Drosophila* homolog of a vertebrate basic region-leucine zipper transcription factor, CCAAT enhancer binding protein (C/EBP; Montell et al. 1992). In animals harboring strong *slbo* mutations, BCs fail to initiate migration, whereas weaker *slbo* mutations cause delayed initiation. Thus, *slbo* appears to play a role in determining the ability of BCs to disassociate from neighboring epithelial cells, their ability to attach to germ cells, or both of these processes. The *breathless (btl)* gene encodes a FGF receptor tyrosine kinase homolog that appears to be a direct transcriptional target of *slbo*, and mutations in *btl* dominantly enhance BC migration defects associated with weak *slbo* mutations (Murphy et al. 1995). Whereas *slbo* and *btl* are crucial for triggering BC differentiation and movement, little is known about the molecules that are involved in positively or negatively regulating the shape changes, migration, and interaction of BCs with germ cells in a specific pattern. We show that Discs large (Dlg), known for regulating cell shape in imaginal epithelia (Woods and Bryant 1991), is likely to be such a molecule.

<sup>1</sup>Corresponding author.

E-MAIL [goode@rascal.med.harvard.edu](mailto:goode@rascal.med.harvard.edu); FAX (617) 432-7688.



**Figure 1.** Border cell migration. Arrowheads point to BCs in each panel. (a–d) Expression of Armadillo (Arm)  $\beta$ -catenin from s8 to s10a. Arm is enriched in BCs before and during their migration through nurse cells (ncs) to the oocyte (o), in concert with the follicular epithelium (fe, double-headed arrows). Migration is complete at s10a (d). (e–g) Wild-type s9 egg chambers stained with phalloidin to reveal cellular cortices. (e) Nurse cells in this s9 egg chamber have the commonly observed quadrihedral shape. (f) When BCs have moved between anterior and central nurse cells, a posterior nurse cell, connected to the oocyte, extends a cytoplasmic process that contacts the BCs (boxed area). (g) An enlargement of the nurse cell process (arrows).

Dlg belongs to a conserved family of proteins termed membrane associated guanylate kinases (MAGuKs; Woods and Bryant 1991, 1993). These proteins share three amino-terminal PDZ domains (named for the family members PSD-95, Dlg, and ZO-1), an SH3 domain, and a carboxy-terminal guanylate kinase (GuK) homology domain (for review, see Anderson 1996; Sheng 1996). Dlg molecules may bind to cytoskeletal proteins through the SH3 domain, a known adaptor motif mediating direct association with cytoskeletal and signaling molecules (Mayer and Eck 1995). The GuK-homology domain has not been shown to have kinase activity, and is missing two amino acids thought to be crucial for catalytic activity (Woods and Bryant 1993). In *Drosophila*, the GuK domain has been shown to play a specific role in prohibiting cell proliferation (Woods et al. 1996).

The three PDZ motifs are perhaps the most intriguing aspect of MAGUK structure. Human Dlg binds via the third PDZ motif to APC, a commonly mutated tumor suppressor gene in human colon cancer cells (Matsumine et al. 1996). PDZ motifs have also been shown to bind to the extreme carboxy-terminal tail of several distinct families of signaling receptors and channels in a sequence-specific fashion (Songyang et al. 1997), and genetic evidence for PDZ involvement in receptor localization at specific subcellular sites has been obtained in *Caenorhabditis elegans* (Simske et al. 1996). PDZ domains have thus emerged as novel adaptor modules for specific protein-protein binding, important for clustering membrane proteins as well as linking signaling molecules in multiprotein complexes at specialized membrane sites (for review, see Anderson 1996; Sheng 1996).

*Drosophila* Dlg is essential for prohibiting cell growth and for maintaining cell adhesion and cell polarity in both embryonic and adult tissues (Gateff 1978; Perrimon 1988; Woods and Bryant 1989, 1990; Woods et al. 1996). In animals harboring *dlg* null mutations, aberrant cell polarity is revealed in the aberrant organization of both the actin and microtubule cytoskeletons, the transfor-

mation of columnar epithelial cells to a apolar morphology, and the delocalized distribution of cell adhesion molecules (Woods et al. 1996). The role of Dlg in maintaining cell polarity appears to be separable from its role in proliferation control, because several mutations that eliminate carboxy-terminal sequences, including the entire GuK domain, cause loss of proliferation control, without affecting cell polarity (Woods et al. 1996).

In this paper we show for the first time that Dlg is required to block cell invasion and that Dlg activity appears to define a novel developmental pathway. Starting very early in oogenesis, loss of *dlg* activity causes follicle cells to overproliferate at the poles of the egg chamber and invade germ tissue. Invading follicle cells change shape and move in a pattern similar to BCs, suggesting that the BC migration pathway is established very early in oogenesis. Both functional and morphological evidence indicates that cooperation between germ cell and follicle cell Dlg, probably mediated by Dlg PDZ domains, is crucial for regulating cell mixing. On the basis of these findings, we suggest a model for Dlg action in prohibiting interactions between tissue layers during oogenesis, and for releasing this prohibition in a regulated manner when BCs migrate. We propose that Dlg does not simply inhibit individual cell behaviors during oogenesis, but rather acts in a developmental pathway essential for blocking cell proliferation, shape change, and migration in a spatio-temporally defined manner.

## Results

### BC migration

Because our analysis of Dlg function during oogenesis focuses on BC migration, we describe and extend previous observations of this process relevant to our studies. Two postmitotic follicle cell populations migrate at s9 of oogenesis (Fig. 1; see Fig. 5, below, for stages of oogenesis). Most follicle cells surrounding nurse cells move to

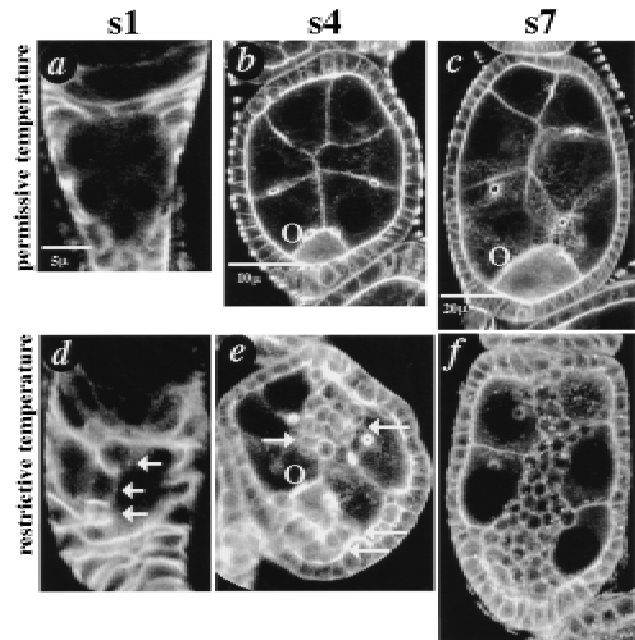
the oocyte as an epithelial sheet along the outside of the egg chamber, whereas the BCs move through the center of the egg chamber, in concert with the epithelium. Six to seven BCs break from the follicular epithelium and adopt a mesenchymal-like morphology, then migrate to the oocyte, as they contact anterior nurse cells, then posterior nurse cells, before reaching their destination (Fig. 1).

The interaction of BCs with posterior nurse cells is particularly dramatic. Nurse cells maintain an invariant quadrihedral-like architecture throughout most stages of oogenesis (Fig. 1e; Fig. 5b, below), but following initiation of BC movement, one nurse cell adjacent to the oocyte extends a cytoplasmic process that contacts the migrating cells (Fig. 1f,g). Nurse cell processes are never observed in s8 egg chambers, preceding BC migration, although BC fate has already been established (data not shown; see also Fig. 4, below). Further, nurse cell processes show a clear directionality. They are never observed to extend from anterior nurse cells to BCs that have moved close to the oocyte at the posterior of the egg chamber. These observations suggest that nurse cells adjacent to the oocyte play an active role in guiding the movement of BCs to the oocyte, a suggestion further supported by analysis described below.

#### Reduction of *dlg* activity leads to an invasive phenotype starting as early as s1 of oogenesis

To analyze the function of *dlg* during oogenesis, we took advantage of temperature-sensitive allele combinations (Perrimon 1988). *dlg<sup>hf321</sup>/dlg<sup>lv55</sup>* animals are viable and have completely normal egg chambers at 18°C. When these animals are shifted to 25°C for at least 6 hr, follicle cells exit the follicular epithelium and intermingle with germ cells at the anterior and posterior poles of the egg chamber (Fig. 2e; data not shown). Cells that remain in the epithelium retain their polarized, epithelial characteristics, whereas cells that have exited the epithelium have an apolar morphology. Follicle cells that have exited the epithelium in anterior regions of the egg chamber invade between nurse cells as they migrate towards the oocyte (Fig. 2, cf. e and f). The phenotype is evident as early as s1 of oogenesis, just following the birth of new egg chambers (Fig. 2d).

Several morphological characteristics suggest that *dlg* invasive cells behave like BCs during wild-type oogenesis. First, BCs always migrate as a cluster of interconnected cells (Fig. 1), and likewise, *dlg* invasive cells always migrate as streams of interconnected cells (Fig. 2d-f). Furthermore, like BCs, invasive cells always migrate through the center of the egg chamber. They never diverge on their path to move between lateral nurse cell membranes (Figs. 2-4, 6, 7, and 9). That invasive cells are attracted to the oocyte is further supported by simple temperature-shift experiments. Temperature shifts of increasing duration result in increasingly larger streams of follicle cells that move increasingly closer to the oocyte (data not shown). Furthermore, if *dlg<sup>hf321</sup>/dlg<sup>lv55</sup>* animals are shifted to the restrictive temperature for 6 hr,



**Figure 2.** *dlg* invasive phenotypes. Invasive phenotypes were analyzed as described in the text. Confocal microscopy was used to capture single optical sections through the center of egg chambers that had been stained with fluorescein-labeled probes. Egg chambers are not shown to scale (see scale bars and Fig. 5; oogenesis is reviewed in Spradling 1993). (a,d) s1 *dlg<sup>hf321</sup>/dlg<sup>lv55</sup>* egg chambers were stained with FasIII to reveal follicle cell outlines. (a) A s1 egg chamber that developed at 18°C for several days appears wild type. (d) An egg chamber that developed at 25°C for 2 days. Three follicle cells have migrated through the center of the egg chamber along the presumptive BC pathway (arrows). (b,c,e,f) s4 and s7 *dlg<sup>hf321</sup>/dlg<sup>lv55</sup>* egg chambers were stained with phalloidin to reveal cellular cortices. (b,c) s4 and s7 egg chambers that developed at 18°C for several days appear wild type. (e) s4 egg chamber that developed at 25°C for 18 hr. Apolar follicle cells have accumulated around the oocyte at the posterior pole of the egg chamber (arrows; O, oocyte), and are invading from the anterior pole (arrows). (f) s7 egg chamber that developed at 25°C for 36 hr. Follicle cells have invaded germ tissue in a pattern resembling BC migration.

then placed at the permissive temperature for several hours, a small clump of follicle cells separates from the anterior epithelium; 24 hr later, the small clump of cells is typically found adjacent to the oocyte, and never outside the center of the egg chamber (data not shown).

The same invasive phenotype, with varying degrees of severity, is found for many *dlg* mutant combinations (Table 2, below). The degree of severity directly correlates with that described for imaginal disc tissue (Table 1; Woods et al. 1996). For example, the stage at which the *dlg* invasive phenotype is first manifested in *dlg<sup>hf321</sup>/dlg<sup>1P20</sup>* (or *v59* or *X1-2*) animals directly correlates with the degree of GuK truncation associated with the *dlg<sup>1P20</sup>*, *dlg<sup>v59</sup>*, and *dlg<sup>X1-2</sup>* mutations (Table 1), with the weakest allele combinations only disrupting the latest stages of oogenesis, and increasingly stronger allele combinations disrupting increasing earlier stages of oogenesis. A simi-

**Table 1.** Characteristics of *dlg* mutations used in this study

Mutation	Invasive phenotype <sup>a</sup>	Genetic characteristic <sup>b</sup>	Phenotypic characteristic <sup>c</sup>	Molecular characteristic <sup>d</sup>
<i>m52</i>	yes	zygotic lethal, behaves like a null, fails to complement all <i>dlg</i> mutations	loss of PC, CP, and SJ structure <sup>e</sup>	truncated protein comprised of PDZ1 and PDZ2, expressed at very low levels compared to wild type
<i>hf321</i>	yes	zygotic lethal, plf <sup>f</sup> ts <sup>g</sup>	N.D.	5.5-kb insert at the 5' end of the gene; effect on Dlg protein not known
<i>sw</i>	no	viable, plf	loss of PC; CP and SJs normal	carboxyl-end protein truncation, just outside of the GKD <sup>h</sup>
<i>1P20</i>	N.D.	zygotic lethal, plf	loss of PC; CP and SJs normal	eliminates the carboxyl third of the GKD
<i>lv55</i>	no	zygotic lethal, plf	loss of PC; CP and SJs normal	not characterized
<i>v59</i>	no	zygotic lethal, plf	loss of PC; CP and SJs normal	eliminates the carboxyl two-thirds of the GKD, reduces protein levels by half
<i>X12</i>	no	zygotic lethal, plf	loss of PC; CP and SJs normal	truncated protein lacking the GKD
<i>m35</i>	no	zygotic lethal, plf	loss of PC; CP and SJs normal	L <sub>632</sub> > P missense mutation in the SH3 domain

<sup>a</sup>The mutations *m52*, *v55*, *v59*, *X12*, and *m35* were assayed by the germ cell-follicle cell clone technique described in the text; *hf321* was assayed by rearing homozygous animals at 18°C, the permissive temperature, then shifting them to 25°C, the restrictive temperature; *sw* is a female sterile allele that was directly assayed in homozygous females.

<sup>b</sup>The genetic characteristics of the mutations used in this study are described in detail in several reports (Perrimon 1988; Woods and Bryant 1989, 1991; Woods et al. 1996).

<sup>c</sup>Effect of *dlg* mutations on imaginal and salivary gland epithelial cells (Woods and Bryant 1989, 1991; Woods et al. 1996).

<sup>d</sup>Molecular characteristics were established in several publications (Woods and Bryant 1989, 1991; Woods et al. 1996).

<sup>e</sup>(PC) Proliferation control; (CP) cell polarity; (SJ) septate junction.

<sup>f</sup>(plf) Partial loss of function.

<sup>g</sup>(ts) Temperature sensitive.

<sup>h</sup>(GKD) Guanylate kinase domain.

lar temporally graded pattern of phenotypic expression has been observed for mutations disrupting components of other signaling and adhesion pathways during oogenesis (Goode et al. 1992, 1996a,b). Because a similar invasive phenotype is observed in many *dlg* mutant combinations, including clones of a null *dlg* mutation (see below), we conclude that the invasive phenotype is caused by a reduction or loss of *dlg* activity.

#### *Invasive follicle cells have not adopted a BC fate*

Because *dlg*<sup>hf321</sup>/*dlg*<sup>lv55</sup> follicle cells behave like BCs, we determined whether they have adopted a BC fate. The *slbo* gene (Montell et al. 1992) is specifically expressed in BCs at s8, before they break from the epithelium and initiate migration to the oocyte, and *slbo* continues to be expressed in BCs during their migration to the oocyte at s9 (Fig. 3a–c). We analyzed the expression of *slbo* in *dlg* mutant egg chambers. *slbo* is expressed in the wild-type pattern in s8 and s9 *dlg*<sup>hf321</sup>/*dlg*<sup>lv55</sup> mutant egg chambers and is not expressed in invasive follicle cells (Fig. 3d,e). *slbo* expression never initiates before s8 in *dlg* mutant animals (not shown). We also analyzed expression of FasIII (Patel et al. 1987), which is specifically expressed in two polar cells at the anterior of the egg chamber starting during mid oogenesis. The polar cells are included among the cells that will go on to migrate as border cells (not shown). We did not find an increase in the number of FasIII-positive cells in *dlg*<sup>hf321</sup>/*dlg*<sup>lv55</sup> mu-

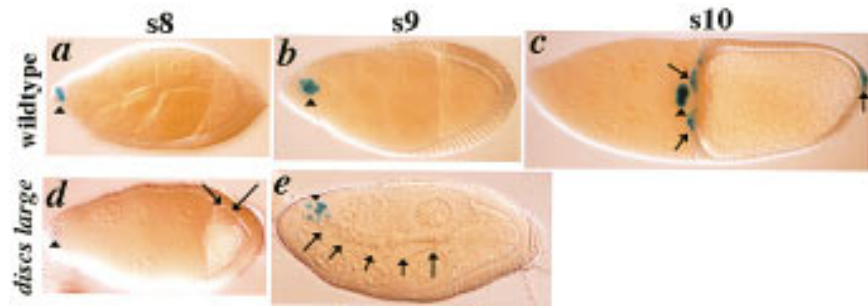
tant egg chambers, or in egg chambers harboring follicle cell clones of the null *dlg*<sup>m52</sup> mutation, further suggesting that overaccumulating follicle cells have not acquired a border cell fate (data not shown). Finally, we note that whereas *dlg*-invasive cells share an apolar morphology with border cells, other aspects of their morphology, such as absence of lamellipodia-like structures (Figs. 1, 2), are not shared with border cells. On the basis of these observations, we conclude that *dlg* invasive follicle cells have not adopted a BC fate, yet, as described below, these cells undergo and participate in several morphogenetic transitions characteristic of BC invasion.

#### *The oocyte and nurse cells attached to the oocyte extend processes that contact invasive follicle cells*

Although *dlg*<sup>hf321</sup>/*dlg*<sup>lv55</sup> follicle cells have not adopted a BC fate, they resemble BCs in their apolar morphology and migration pattern. We sought to determine if *dlg* follicle cells also behave like BCs in their pattern of interaction with nurse cells attached to the oocyte (Fig. 1e–g). To perform these experiments, *dlg*<sup>hf321</sup>/*dlg*<sup>lv55</sup> animals were shifted to the restrictive temperature for 6–12 hr, so that invasive follicle cells would initiate migration just past the first nurse cells (Fig. 4a), the point at which BCs come in contact with a process extended by an oocyte-associated nurse cell (Fig. 1e–g). In *dlg*<sup>hf321</sup>/*dlg*<sup>lv55</sup> mutant egg chambers, nurse cells extend processes to meet invasive follicle cells. This is clear by s6,



**Figure 3.** Invasive follicle cells are not border cells. Egg chambers harbor a single copy of a *lacZ* enhancer trap gene inserted in the *slbo* gene (Montell et al. 1992). *slbo* expression is revealed by X-gal staining (blue). (a–c) Expression of *slbo* in wild-type s8–s10 egg chambers. *slbo* expression initiates at s8 in a small cluster of presumptive BCs (arrowheads), and is maintained in BCs as they migrate to the oocyte. At s10, *slbo* is also expressed in follicle cells in the anterior follicular epithelium that move between the oocyte and nurse cells (arrows), and in a cluster of follicle cells at the posterior of the egg chamber (arrow). (d–e) Expression of *slbo* in *dlg<sup>hf321</sup>/dlg<sup>lv55</sup>* egg chambers. (d) s8 egg chamber that was shifted to the restrictive temperature for 8 hr in which *slbo* expression has initiated (arrowhead). A small cluster of follicle cells that do not express *slbo* have segregated from the epithelium surrounding the oocyte at the posterior of the egg chamber (arrows). (e) A s9 egg chamber that was shifted to the restrictive temperature for 36 hr in which *slbo* is expressed in border cells as they initiate migration but is not expressed in the stream of invasive cells that have moved to the oocyte (arrows).



when the egg chamber has acquired an elongated shape (Fig. 5a), but as for the invasive phenotype, these interactions probably start as early as s1 (see also Fig. 2e). We confirmed these interactions for other allele combinations. For example, the interactions are also observed in *dlg<sup>hf321</sup>/dlg<sup>1P20</sup>* animals, in which cells do not invade before s7 (Fig. 4b; Table 2).

We also found that invasive follicle cells show unusual interactions with the oocyte. Once invasive cells have migrated to the most posterior nurse cells, the oocyte elongates to contact the cells (Fig. 4c,d; see also Fig. 8d). Although the oocyte does not extend during BC migration in wild-type animals, this result documents the selective adhesive forces acting between the oocyte and BCs, and suggests that *dlg* plays a role in maintaining oocyte architecture during wild-type oogenesis.

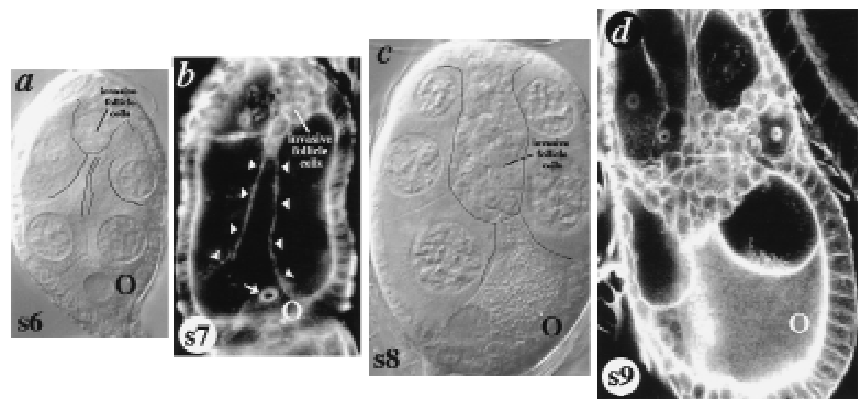
#### *Dlg* expression

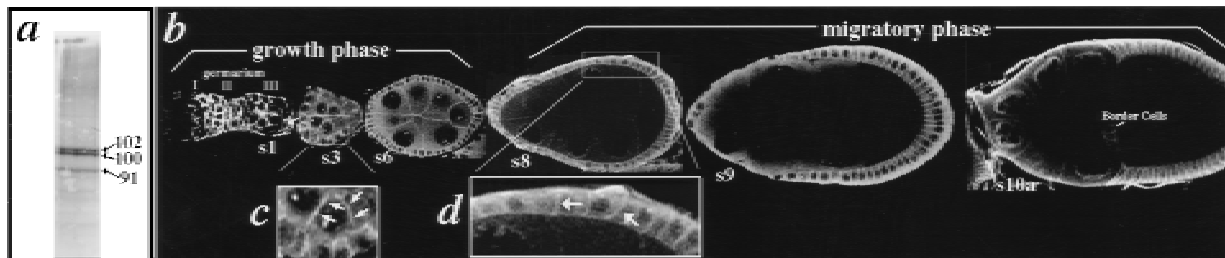
To help establish the basis for the invasive behavior of follicle cells in *dlg* mutant egg chambers, we analyzed

the spatio-temporal and subcellular distribution of Dlg proteins throughout oogenesis. Western analysis reveals three polypeptide species of about 91, 100, and 102 kD (Fig. 5a), approximately the same mass described for Dlg proteins in imaginal discs and larval muscles (Lahey et al. 1994; Woods et al. 1996). Dlg is expressed in both germ and follicle cell tissues from the time that the germ cell cyst becomes surrounded by follicle cells in the gerarium (Fig. 5b). Dlg appears to be expressed at equivalent levels in both tissues throughout the growth phases, as the germ cell cyst expands in size and the number of follicular epithelial cells increases >10-fold. Following cessation of follicle cell proliferation, levels of Dlg protein appear to dramatically decrease in germ cells, corresponding to the time just preceding and including BC migration to the oocyte (Fig. 5b).

At the cellular level, Dlg is localized to sites of contact between follicle cells and to sites of contact between germ cells, but appears to be excluded at sites of contact between germ cells and follicle cells (Fig. 5c,d). Likewise, before and during BC migration, Dlg is expressed at sites

**Figure 4.** Posterior germ cells extend processes that contact invasive follicle cells. (a) s6 *dlg<sup>hf321</sup>/dlg<sup>lv55</sup>* egg chamber shifted to the restrictive temperature for 6 hr. Nurse cells adjacent to the oocyte extend cytoplasmic processes that contact newly invading follicle cells (outlined). (b) s7 *dlg<sup>hf321</sup>/dlg<sup>1P20</sup>* egg chamber. A nurse cell attached to the oocyte (via the ring canal, arrow) extends a cytoplasmic process to the anterior pole of the egg chamber (arrowheads), contacting newly invading follicle cells. Note the resemblance of this process to that extended at s9 during BC migration in wild-type egg chambers (Fig. 1). (c) s8 *dlg<sup>hf321</sup>/dlg<sup>lv55</sup>* egg chamber in which invading follicle cells have migrated over half the distance to the oocyte. The oocyte extends to meet invading cells. (d) Phalloidin-stained s9 egg chamber harboring a presumptive simultaneous germ cell and follicle cell clone of the genetic null *dlg<sup>m52</sup>* mutation (see Fig. 5; Materials and Methods). This egg chamber documents a rare instance in which follicle cells invade from lateral portions of the egg chamber, as well as from the anterior pole. The dramatic extension of the oocyte towards the invading cells shows the invading cells attraction to the posterior egg cell.





**Figure 5.** Expression of Dlg during oogenesis. (a) Western blot of ovarian proteins probed with Dlg sera. Three polypeptides of 91, 100, and 102-kD are detected (arrowheads). (b) Ovarian tissue probed with Dlg sera and visualized by confocal microscopy. Dlg is expressed in germ cells and follicle cells starting in the germarium [where the germ cell cyst becomes ensheathed by follicle cells in region II; oogenesis is reviewed in Spradling (1993); cell types are labeled in Fig. 1]. Dlg protein expression continues in both cell types throughout the growth stages of oogenesis, as the germ cell cyst expands and the follicular epithelium increases from ~40 cells at s1 to 650 cells at s6. Dlg staining is detected in both the cytoplasm and membrane. Staining is enriched in membranes at sites of contact between germ cells (c, arrows), and sites of contact between follicle cells (d, arrows). Following cessation of follicle cell division at s6, Dlg protein levels dramatically decrease in germ cells during s8–s10 (b), a migratory phase when follicle cell populations move to the oocyte (see also Fig. 1).

of contact between border cells, but not at points of contact between border cells and germ cells (data not shown). Considerable focus has been placed on the localization and function of Dlg in epithelial septate junctions (Woods and Bryant 1991, 1993), but our analysis does not address the function of Dlg in these structures, because septate junctions are not found in the germ cell cyst or follicular epithelium throughout the stages preceding and including BC migration (Mahowald 1972).

#### *Dlg is required in both germ cells and follicle cells*

Because Dlg is expressed in both germ cells and follicle cells, we sought to determine the respective function of germ cell Dlg and follicle cell Dlg in the genesis of the invasive phenotype. To determine where Dlg is required, germ cell and follicle cell clones were generated with the genetic null mutation *dlg<sup>m52</sup>* (Fig. 6b). Eliminating *dlg* function in germ cells alone did not cause any defects, except for occasional misshapen germ cells (Fig. 6c, not shown). Eliminating *dlg* function in follicle cells alone led to overaccumulation of follicle cells at the anterior and posterior poles of the egg chamber (as in *dlg<sup>hf321</sup>*/*dlg<sup>lv55</sup>* egg chambers, see Fig. 2e), but these cells invade germ tissue only rarely (Fig. 6d; invasion occurs in ~1/50 egg chambers,  $n = 425$ , Fig. 6f). When *dlg* function is eliminated in both germ cells and follicle cells, follicle cells always invade along the BC pathway (Fig. 6e). We conclude that Dlg acts in both germ cells and follicle cells to block cellular invasion.

#### *Dlg appears to be required for prohibiting cell proliferation at the poles of egg chambers*

We considered two possibilities for the overaccumulation of follicle cells at the poles of *dlg* mutant egg chambers (Fig. 2e; see below). One possibility is that follicle cells have redistributed from lateral regions of the egg chamber to the egg chamber poles. This does not seem

likely because even after 36 hr at the restrictive temperature, and the massive invasion of follicle cells, lateral regions of the follicular epithelium do not appear to differ in cell density from wild-type epithelia (Fig. 2, c and f). A second possibility is that follicle cell overaccumulation results from follicle cell overproliferation.

To obtain evidence for overproliferation of follicle cells, we compared the degree to which follicle cells at the poles of wild-type and mutant egg chambers incor-

**Table 2.** Phenotypic characteristics of heteroallelic *dlg* combinations

Strength	Follicle cells invade <sup>a</sup>	Follicle cells do not invade
weak <sup>b</sup>	sw/m35, sw/hf321	
intermediate <sup>c</sup>	1P20/hf321, v59/hf321	1P20/m35, lv55/m35, v59/m35
strong <sup>d</sup>	X12/hf321, lv55/hf321	sw/sw, 1P20/sw, v59/sw, lv55/sw, X12/sw, m52/sw
lethal <sup>e</sup>	hf321/m52, 1P20/m52, lv55/m52, v59/m52, X12/m52, m35/m52, lv55/v59, 1P20/v59, X12/v59, 1P20/lv55, X12/lv55, 1P20/X12, m35/X12, hf321/m35	

The phenotypes associated with the homozygous mutants is given in Table 1.

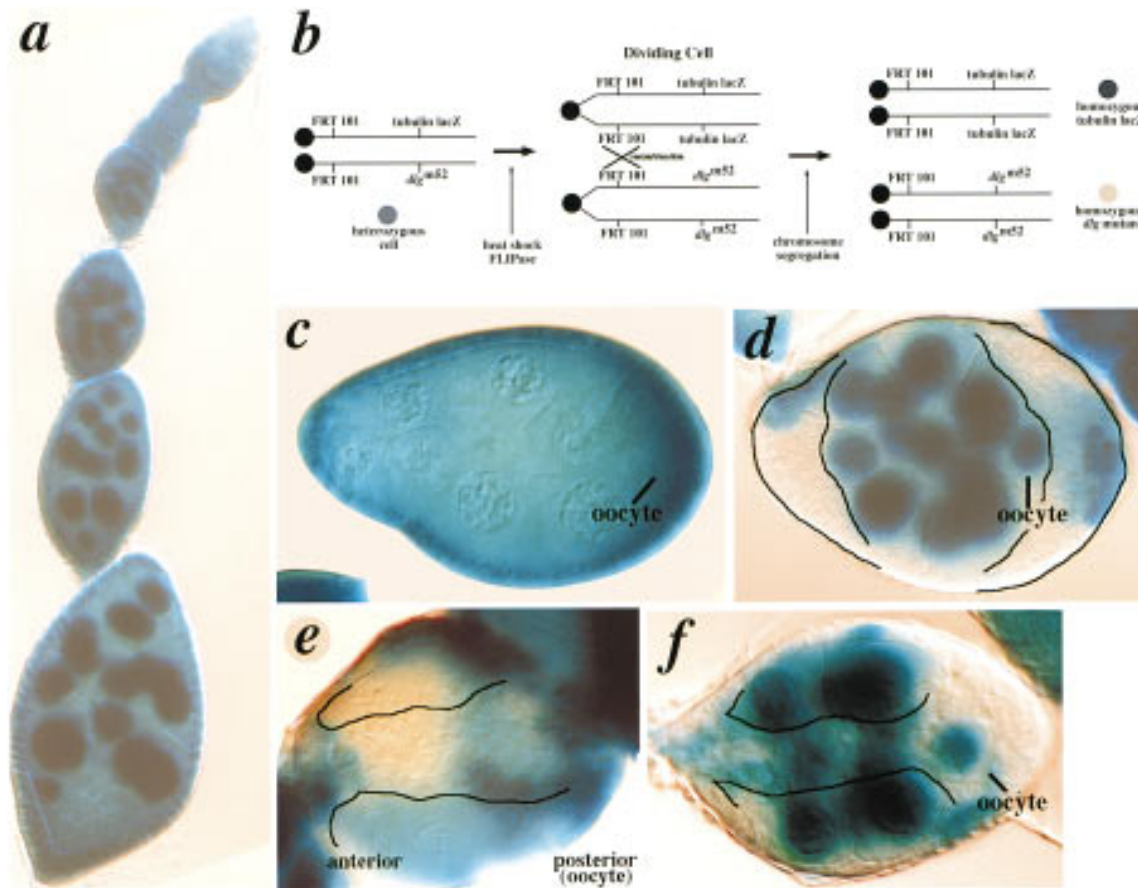
<sup>a</sup>Invasive vs. noninvasive phenotypes are described and pictured in the text.

<sup>b</sup>BCs leave prematurely at s8, or move prematurely at s9. Since BCs are determined at s8 and migrate between germ cells at s9, the designation of “Follicle cells invade” or “Follicle cells do not invade” is not meaningful.

<sup>c</sup>Follicle cells accumulate at the poles and/or invade typically starting no earlier than s5.

<sup>d</sup>Follicle cells accumulate at the poles and/or invade starting at least as early as s3, and for most allele combinations, the earliest time that a mutant phenotype was observed is s1.

<sup>e</sup>Phenotypic analysis not possible because the mutant combinations are lethal at both 18°C and 25°C.



**Figure 6.** Clonal analysis. Except in rare instances, cell invasion depends on loss of *dlg* in both germ and follicle cells. (a) X-gal stained ovariole from an FRT<sup>101</sup>, TUBlacZ/ FRT<sup>101</sup>, *dlg*<sup>m52</sup>; hsFLPase animal. The tubulin promoter drives *lacZ* expression (TUBlacZ) in both germ and follicle cell nuclei (see Materials and Methods). (b) Scheme for generating germ and follicle cell clones (see Materials and Methods). Following sister chromatid exchange, homozygous mutant cells do not stain because they no longer harbor the TUBlacZ gene. (c) *dlg*<sup>m52</sup> germ-line clone. Egg chambers with homozygous *dlg*<sup>m52</sup> germ cells and heterozygous follicle cells usually appear wild type. (d) *dlg*<sup>m52</sup> follicle cell clone. Egg chambers with homozygous *dlg*<sup>m52</sup> follicle cells and heterozygous germ cells have multiple layers of follicle cells at the anterior and posterior poles (outlined). (e) Egg chamber with homozygous mutant *dlg*<sup>m52</sup> germ cells and follicle cells. Follicle cells invade germ tissue (outlined). (f) *dlg*<sup>m52</sup> follicle cell clone. Although germ cells harbor a wild-type copy of *dlg*, follicle cells invade (outlined). These egg chambers are observed at ~2% frequency ( $n > 200$ ).

porate BrdU, a marker for newly dividing cells, as a function of distance along the anterior–posterior axis. Wild-type follicle cells incorporate BrdU at the same frequency independent of anterior–posterior position (Fig. 7a,c). In contrast, the relative frequency with which *dlg*<sup>hf321</sup>/*dlg*<sup>lv55</sup> follicle cells incorporate BrdU is higher at the anterior and posterior poles of the egg chamber (Fig. 7b,c). The difference between mutant and wild type was even more pronounced if the degree of incorporation of BrdU in follicle cells thought to be most susceptible to loss of *dlg* activity, those residing strictly within the plane of invasive cells, was analyzed (Fig. 7c).

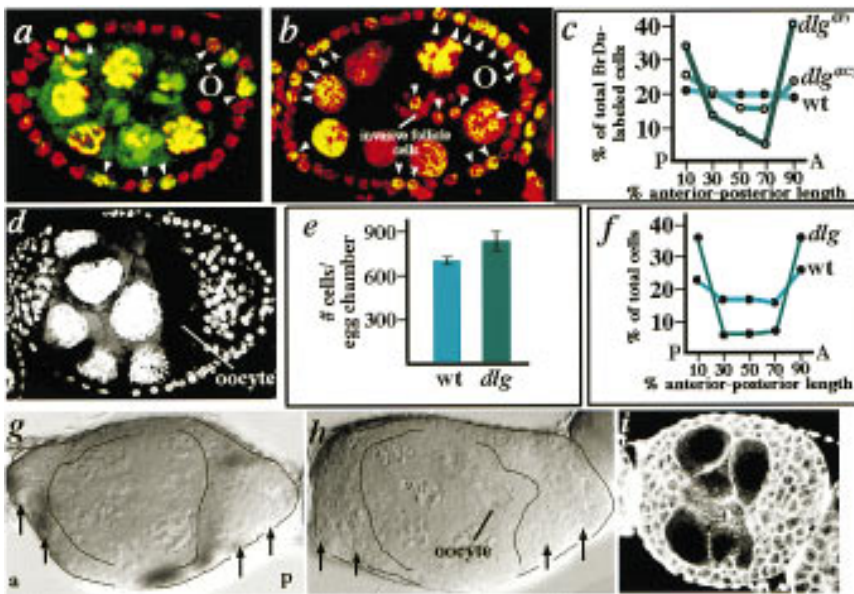
To further substantiate that follicle cells overproliferate in *dlg* egg chambers, the total number of follicle cells in *dlg*<sup>sw</sup> versus wild-type s5–s6 egg chambers was compared. In *dlg*<sup>sw</sup> egg chambers, in which follicle cells accumulate at the poles of the egg chamber without invading (Fig. 7d), there are on average ~150 more follicle cells per egg chamber compared with wild type (Fig. 7e). Fur-

ther, these follicle cells appear to overaccumulate at the anterior and posterior poles of the egg chambers, where we observed increased BrdU incorporation in *dlg*<sup>hf321</sup>/*dlg*<sup>lv55</sup> egg chambers (Fig. 7f). Combined, these findings strongly indicate that BrdU incorporation at the poles of *dlg*<sup>hf321</sup>/*dlg*<sup>lv55</sup> egg chambers results from loss of proliferation control, and not from a redistribution of follicle cells.

#### *dlg* mutations that map to the SH3 and GuK domains do not confer premature cell mixing

*dlg* mutations that behave as genetic nulls, such as *dlg*<sup>m52</sup> (described above), cause loss of proliferation control, cell polarity, and adhesion in imaginal epithelia (Woods et al. 1996; Table 1), and an invasive phenotype when removed from germ cells and follicle cells during oogenesis (Fig. 6). In contrast, *dlg* mutations that specifically disrupt the SH3 and GuK domains cause loss of





**Figure 7.** Separation of Dlg invasion and proliferation phenotypes. (a–c) Follicle cells at the anterior and posterior poles of  $dlg^{hf321}/dlg^{lv55}$  mutant egg chambers preferentially incorporate BrdU. BrdU is a nucleotide analog that is incorporated into nuclei that are in S-phase, or have entered S-phase during the experimental period. Note the intense accumulation of BrdU in nurse cell nuclei (yellow), which are polyploid. (a) s6 wild-type egg chamber. All follicle cells have been labeled with propidium iodide (red nuclei), whereas follicle cells that have incorporated BrdU during the 12 hr experimental period have yellow nuclei (arrow heads; see Materials and Methods). (b) s6  $dlg^{hf321}/dlg^{lv55}$  egg chamber with invading follicle cells migrating to the oocyte. Follicle cells at the poles of the mutant egg chambers appear to have incorporated BrdU to a greater extent than wild type. (c) Graph comparing the average distribution of BrdU-labeled follicle cells

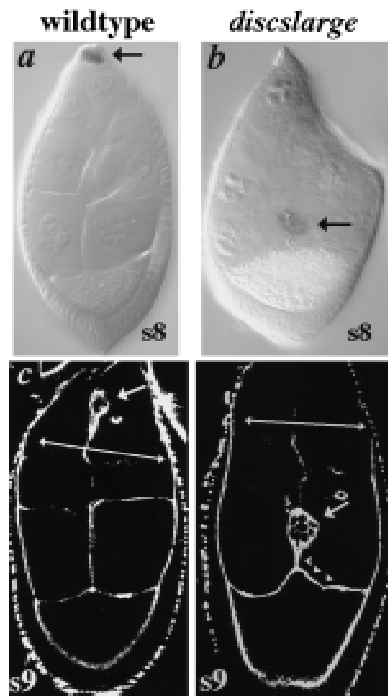
at the anterior and posterior poles of wild-type and mutant egg chambers. BrdU-labeled follicle cells are found at about equal frequency along the anterior-posterior axis of wild-type egg chambers, but are found at greater frequency at the anterior and posterior poles of  $dlg^{hf321}/dlg^{lv55}$  mutant egg chambers [ $dlg^{ec}$ ]. The difference between mutant and wild type is more pronounced if the degree of BrdU incorporation in follicle cells that reside strictly within the plane that includes invasive cells is analyzed [ $dlg^{lp}$ ]. (d–i) Characterization of  $dlg$  genotypes and phenotypes in which follicle cells overaccumulate, but do not invade. (d) A  $dlg^{sw}$  egg chamber. Follicle cells accumulate at the poles of the egg chamber, but do not invade. (e) Graph comparing the total number of follicle cells in  $dlg^{sw/+}$  (wild type) vs.  $dlg^{sw}$  egg chambers ( $n = 4$  egg chambers for each genotype). There are ~150 more follicle cells in mutant egg chambers on average. (f) Graph comparing the distribution of follicle cells in the center of s7  $dlg^{sw/+}$  vs. the center of s7  $dlg^{sw}/dlg^{sw}$  egg chambers (numbers obtained from confocal sections like those shown in a and d;  $n = 6$  egg chambers for each genotype). Many more follicle cells are found at the poles of s7  $dlg^{sw}$  egg chambers compared with  $dlg^{sw/+}$  egg chambers. Note that supernumerary follicle cells are found in the same position wherever increased BrdU incorporation is found in  $dlg^{hf321}/dlg^{lv55}$  egg chambers (c). (g,h) Egg chambers harboring simultaneous germ cell and follicle cells clones of  $dlg^{X1-2}$  and  $dlg^{m35}$ , respectively (produced as in Fig. 6). Follicle cells accumulate at the poles of egg chambers (outlined, arrows), but do not invade germ tissue. (i)  $dlg^{v59}/dlg^{m35}$  egg chamber stained with phalloidin to reveal actin cortices. Follicle cells overaccumulate at the poles of egg chambers, but do not invade.

proliferation control, but have no effect on cell polarity (Table 1; Woods et al. 1996). To assay the requirement of the SH3 and GuK domains in blocking cell invasion, we analyzed the phenotype of SH3 and GuK mutations by employing the germ cell–follicle cell clone technique described (Fig. 6). In egg chambers in which both germ cells and follicle cells are mutant for  $dlg^{lv55}$  or  $dlg^{X1-2}$  mutations, which partially and almost completely eliminate the GuK domain, respectively, or for the  $dlg^{m30}$  mutation, which disrupts the SH3 domain, follicle cells accumulate at the poles of the egg chamber, but do not invade between germ cells (Fig. 7g,h). Further, in animals heterozygous for  $dlg^{1P20}$  or  $lv55$  (that disrupt GuK sequences), and  $dlg^{m52}$  (that disrupts the SH3 domain), follicle cells accumulate at the poles of egg chambers without invading (Table 2; Fig. 7i). These results indicate that Dlg molecules lacking the activity of the GuK domain, and probably the activity of the SH3 domain, retain the Dlg activity that prohibits follicle cell invasion.

#### Dlg prohibits premature BC migration

Our analysis indicated that Dlg is required for inhibiting several interactions and activities in growing egg cham-

bers that appear to be fundamental to BC migration. Does Dlg play a similar role during the process of BC migration? To analyze the role of  $dlg$  during post-mitotic stages, we used extremely weak mutant combinations, such as  $dlg^{hf321}/dlg^{m35}$  and  $dlg^{hf321}/dlg^{sw}$ , in which no detectable defect on egg chamber development is found until s8–s9, when BCs are determined and migrate (Table 1). At s8, the only observable defect is the occasional premature breaking and movement of BCs from the anterior follicular epithelium (Fig. 8a,b). We do not think it is likely that this phenotype results from delayed migration of the follicular epithelium towards the oocyte, because we do not observe defective epithelium migration in stronger  $dlg$  mutant combinations. At s9, in addition to the premature arrival of BCs to the oocyte, we sometimes observe cytoplasmic extensions between the oocyte and BCs, suggesting excessive interaction between these cells (Fig. 8c,d). These extensions are not found during BC migration in wild-type egg chambers. These results indicate at least two similarities for the role of Dlg during BC migration and the earlier stages of oogenesis. First, the premature breaking of BCs suggests a functional connection to the early invasive phenotype, and second, the pulling of the oocyte by the BCs is remi-



**Figure 8.** Weak *dlg* mutations cause BCs to migrate in advance of the follicular epithelium. (a–b) Egg chambers harbor a single copy of a *lacZ* enhancer trap gene inserted in the *slbo* gene (Montell et al. 1992). *slbo* expression is revealed by X-gal staining of egg chambers that have developed at 25°C. (a) Typical s9 wild-type egg chamber in which BCs migrate in concert with the follicular epithelium (arrows). (b) Typical s9 *dlg<sup>sw</sup>/dlg<sup>hf321</sup>* egg chamber in which BCs have migrated in advance of the follicular epithelium. This is the only defect observed in the ovaries of *dlg<sup>sw</sup>/dlg<sup>hf321</sup>* animals. (c–d) Egg chambers stained with fluorescein-phalloidin and imaged by scanning laser confocal microscopy to reveal cortices of germ and follicle cells. (c) Typical s9 wild-type egg chamber in which BCs migrate in concert with the follicular epithelium (arrows). (d) Typical s9 *dlg<sup>sw</sup>/dlg<sup>hf321</sup>* egg chamber in which BCs have migrated in advance of the follicular epithelium. The oocyte extends to meet the BCs while the BCs extend to meet the oocyte (arrowheads). These excessive interactions between the oocyte and follicle cells are never observed in wild-type egg chambers.

niscient of the cytoplasmic extensions observed between posterior germ cells and invasive follicle cells in egg chambers mutant for stronger *dlg* mutations (Fig. 4c,d).

#### *kek-1* is expressed in BCs and in overproliferating and invasive follicle cell populations

Our analysis indicates that follicle cells at the poles of egg chambers throughout the growth phases, and BCs during the migratory phases, respond differentially to loss of *dlg* during oogenesis. This finding was unexpected because Dlg protein is expressed at uniform levels in all follicle cells throughout oogenesis (Fig. 5). To determine if follicle cells sensitive to loss of Dlg function might be genetically distinct, we screened enhancer trap lines. We found that the *Drosophila kekkon-1* (*kek-1*)

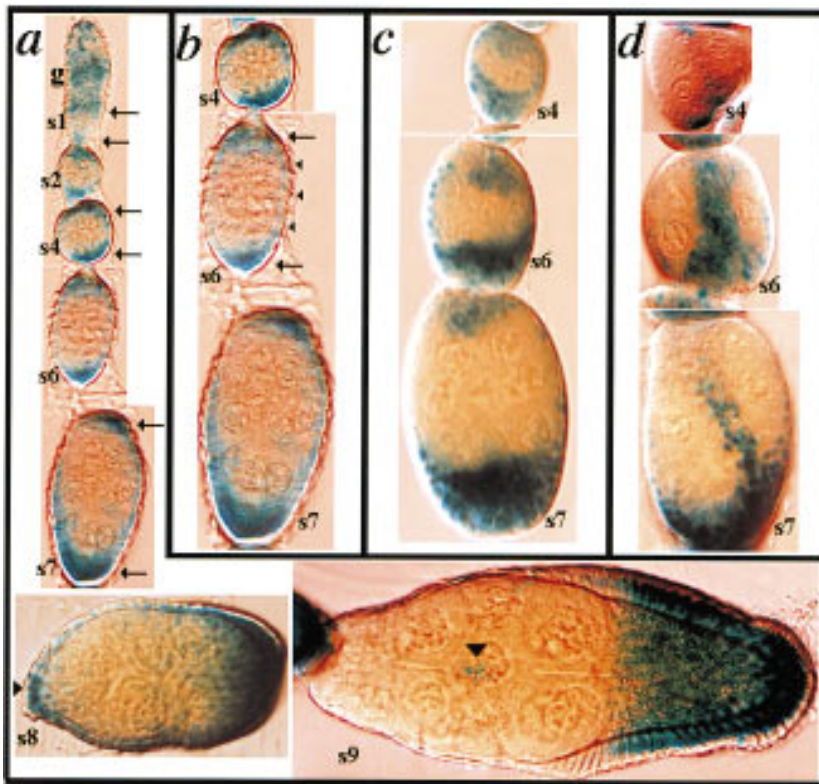
gene (Mussachio and Perrimon 1996) is specifically expressed in fields of follicle cells at the poles of the egg chamber, and in BCs before and during their migration to the oocyte (Fig. 9). This finding suggests that BCs and follicle cells at the poles of the egg chamber are functionally distinct, on the basis of sensitivity of these cells to loss of Dlg function, and genetically distinct, on the basis of their expression of the *kek-1* gene.

#### Discussion

*Dlg acts in a developmental pathway that prohibits the cell shape changes and morphogenetic transitions characteristic of BC migration*

We show for the first time that Dlg is required to prohibit cell invasion, in a pattern resembling border cell migration. BCs are distinguished from other follicle cells by expression of a C/EBP-like factor, encoded by the *slbo* gene (Montell et al. 1992). Following C/EBP expression, BCs become further distinguished by their adoption of an apolar morphology, their migration, and their interaction with germ cells in the posterior of the egg chamber (Fig. 1). We found that the Dlg plays no role in BC specification, but rather is likely to function following C/EBP expression to regulate morphogenetic activities of both BCs and germ cells. In support of this hypothesis, we find that BCs migrate prematurely in animals mutant for weak Dlg mutations (Fig. 8). Premature migration may result from both decreased Dlg activity between BCs and adjacent follicle cells, as indicated by the expression of Dlg at sites of contact between these cells (Fig. 5), and from the excessive interaction of the BCs with the germ cells, as indicated by the hypertrophic cell extensions between BCs and the oocyte (Fig. 8). We speculate that simultaneous reduction and loss of Dlg activity in BCs and germ cells, respectively, allows BC migration in wild-type animals. BC differentiation, induced by *slbo*, may lead to reduction of Dlg activity specifically in BCs (see below), whereas unknown factors apparently lead to a loss of Dlg protein in germ cells (Fig. 5).

That reduction of Dlg in germ and follicle cells is essential for BC migration is indicated by the cellular invasion phenotype observed when Dlg activity is lost or reduced in both germ cells and follicle cells, starting as early as s1 of oogenesis. The cellular invasion phenotype is associated with several cellular events indistinguishable from those guiding BC movement. Both BCs and *dlg* invasive cells undergo an epithelial–mesenchyme-like transition, in which they move from adjacent epithelial cells as they shift their morphology from a polarized epithelial phenotype to an apolar morphology. *dlg* invasive follicle cells then move to the oocyte, along a path that is indistinguishable from the BC pathway. Invasive cells never turn from the central path, between lateral nurse cell membranes. The fidelity of this migration pattern is likely to result from specific interaction of invasive cells with posterior nurse cells, as indicated by the hyperplastic extension of a cytoplasmic process by a nurse cell adjacent to the oocyte (Fig. 4), resembling the contacts



**Figure 9.** *kek-1* is expressed in follicle cells sensitive to loss of *dlg* activity. Egg chambers harbor a single copy of a *lacZ* enhancer trap gene inserted in the 5' noncoding region of *kek-1* (Musacchio and Perrimon 1996). *kek-1* expression is revealed by X-gal staining. (a) *kek-1* is intensely expressed at the anterior and posterior poles of egg chambers throughout the time that follicle cells are proliferating, from s1 to s6, and continuing to s7 (arrows). At s8, *kek-1* is expressed in the presumptive BCs at the anterior of the egg chamber (arrowhead). At s9, *kek-1* is expressed in migrating BCs, and in follicular epithelial cells covering the oocyte, but not in nurse cell-associated oothelial cells. (b) Higher magnification of s4-s7 egg chambers shown in (a). In addition to polar staining, *kek-1* is expressed at lower levels in lateral follicle cells (arrowheads). (c) Multiple layers of follicle cells at the anterior and posterior poles of *dlg<sup>sw</sup>* egg chambers express *kek-1*. (d) Invasive follicle cells in *dlg<sup>h321</sup>/dlg<sup>lv55</sup>* egg chambers express *kek-1*.

established between BCs and an oocyte-adjacent nurse cell during normal oogenesis (Fig. 1). Loss or reduction of Dlg appears to be required in both cell types to permit these interactions, because the invasive phenotype is always manifested when *dlg* activity is eliminated or reduced in both germ cells and follicle cells, but is typically not manifested when one tissue is mutant. This functional requirement is consistent with morphological evidence indicating that both germ and follicle cells play a role in guiding BC movements.

Significantly, our findings imply that Dlg does not simply inhibit individual cell behaviors during oogenesis, but rather may be a component in a developmental pathway essential for blocking cell proliferation and migration in a spatio-temporally defined pattern. *dlg* mutant egg chambers indicate that this pathway acts in germ and BCs and in fields of follicle cells at the anterior and posterior poles of egg chambers. Functionally, this is indicated by anterior and posterior follicle cell shape change, overaccumulation, and movement towards the oocyte (Figs. 2, 6-8; the movement of apolar posterior cells towards the oocyte is best documented in Fig. 2e), and genetically, this is indicated by *kek-1* expression in BCs and follicle cells at the poles of the egg chamber. Strikingly, *slbo* is also expressed at both the anterior and posterior poles of the egg chamber (Fig. 3), further suggesting a relationship between polar follicle cells.

BCs and *dlg* invasive cells are distinguished in that the latter are supplied from a field of overproliferating follicle cells that migrate as a continuous stream, whereas

BCs, derived from a postmitotic epithelium, completely dissociate from the follicular epithelium. These distinctions result from the requirement of Dlg in proliferation control (see below), and from the inability of *dlg* mutant cells to completely break from the follicular epithelium. These two processes appear to be intimately connected. In animals harboring *dlg* temperature-sensitive mutations, follicle cell overproliferation and cell invasion can be turned off simply by placing the animals at the permissive temperature (not shown). Under these circumstances, the epithelium heals, while invading cells continue to move towards the oocyte. Thus, the breaking of BCs from the follicular epithelium during wild-type oogenesis may involve a cycle of reduced Dlg activity for the BCs to exit the epithelium, followed by a return to higher activity to reseal the follicular epithelium.

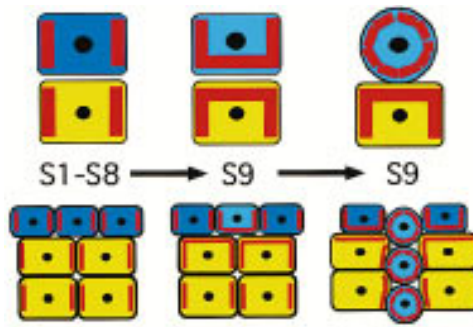
Whereas we have presented several lines of positive evidence implicating Dlg in a pathway that inhibits the invasion characteristic of BCs, specificity implies that Dlg should not be required in many other cellular interactions during oogenesis. Both anterior-posterior and dorsal-ventral specification of follicle cells, which establish the oocyte axes, are known to depend on intimate interactions between germ and follicle cells (Gonzalez-Reyes et al. 1995; Roth et al. 1995), yet we do not observe any defects in these processes in *dlg* mutant egg chambers. Furthermore, we have not observed a role for Dlg in the movement of the follicular epithelium along the outside of the egg chamber during s9. Many aberrant interactions between germ cells and follicle cells could be



imagined, such as simultaneous invasion of follicle cells between the interface of each nurse cell, but such patterns are not observed. Finally, we suggest that the requirement for Dlg in prohibiting follicle cell proliferation may not be distinct from *dlg* functions maintaining epithelial integrity, because the loss of epithelial characteristics of follicle cells during the early stages of oogenesis occurs in the environment of a mitotically active epithelium. Thus, the requirement of Dlg for inhibiting follicle cell division may reflect an indirect consequence of the protein's requirement in maintaining appropriate contact between the epithelial cells, which may be indirectly tied to the regulation of follicle cell proliferation.

#### *Dlg PDZ domains appear to be required for prohibiting follicle cell invasion*

We found that the Dlg SH3 and GuK domains do not appear to play a role in blocking cell invasion (Fig. 7), indicating that the PDZ domains inhibit germ cell–follicle cell mixing. We have not tested this hypothesis directly, because *dlg* mutations that specifically disrupt the PDZ domains do not exist. What role might the PDZ domains play in blocking cell mixing? PDZ domains bind to the cytoplasmic tail of transmembrane molecules and localize them to specific regions on the cell surface (for review, see Anderson 1996; Sheng 1996). During oogenesis, Dlg is localized to sites of contact between germ cells and to sites of contact between follicle cells, but Dlg is absent from sites of contact between germ cells and follicle cells (Fig. 5). This localization almost certainly ensures that presumptive Dlg-bound cell surface molecules retain tissue-specific segregation, and further suggests that Dlg PDZ domains serve as components of a germ cell–follicle cell tissue barrier (Fig. 10). Because the germ cell–follicle cell interactions that we observe in *dlg* mutant egg chambers specifically resemble BC interactions (see previous section), we suggest that Dlg binds to a class of cell surface molecules that have the potential to mediate the cellular interactions characteristic of BC development, starting at s1 (Fig. 10). We propose that at the time of BC migration, via a regulated step that allows the redistribution of at least some of the cell surface molecules bound by Dlg PDZ domains to the interface between germ cells and follicle cells, germ cells and BCs intermingle. Obvious candidates for this regulated step are loss or reduction of Dlg in germ cells, and expression of *slbo* in BCs. Loss or reduction of Dlg in germ cells might lead to the diffusion of previously bound molecules to cellular regions other than the interface with other germ cells. We can only speculate how *slbo* might elicit a freeing of Dlg bound molecules, but precedent for regulated interactions involving MAGuKs is the interaction between the MAGuK PSD-95 and the inward rectifier K<sup>+</sup> channel Kir 2.3, which appears to be blocked by phosphorylation of a serine within the consensus PDZ-binding motif of Kir 2.3 by protein kinase A (Cohen et al. 1996). We suggest that in *dlg* mutant egg chambers, molecules bound by



**Figure 10.** Model for Dlg in prohibiting cell invasion during oogenesis. Germ cells are yellow and follicle cells are blue. The red stripes within in each cell represent the distribution of putative membrane spanning proteins held in place by Dlg PDZ domains (see text). During s1 to s8 of oogenesis, Dlg restricts these putative membrane spanning proteins to sites of contact between germ cells and to sites of contact between follicle cells, as indicated by the localization of Dlg protein during oogenesis. We propose that at s9, differentiation of BCs (light blue), leads to a redistribution of at least some Dlg-bound molecules to sites of contact between germ and follicle cells, whereas loss or reduction of Dlg in germ cells has a similar consequence. The interaction of membrane spanning molecules on the surface of germ and follicle cells then contributes to the driving force for BC invasion between germ cells. In *dlg* mutant egg chambers, the redistribution of PDZ-bound molecules happens in the absence of specific signals, leading to invasion as early as s1 of oogenesis.

PDZ domains are freed prematurely, in the absence of specific signals, resulting in patterns of cell transformation and intercellular interaction resembling BC development. This model suggests that the BC migration pathway has been established very early in oogenesis and provides an explanation for the variegated adhesion phenomena mediated by Dlg, apparently promoting adhesion between follicle cells, while inhibiting contact and excessive interaction between germ cells and follicle cells.

#### *Dlg may help to pattern follicle cell divisions*

Loss or reduction of Dlg activity in the follicular epithelium causes follicle cells to overaccumulate at the anterior and posterior poles of the egg chamber (Figs. 2, 6, 7, and 9). We do not believe that this overaccumulation results from redistribution of follicle cells, because we did not find discontinuities in lateral regions of the epithelium (Figs. 2, 6, 7, and 9). Further, egg chambers appear to have a greater total number of follicle cells, and follicle cells incorporate BrdU higher levels in the region of the epithelium that contains overaccumulating cells (Fig. 7). The spatial specificity of inhibited follicle cell proliferation does not reflect an underlying pattern in follicle cell divisions, because follicle cells in the wild-



type follicular epithelium appear to enter both S phase and M phase at a uniform frequency relative to the anterior-posterior axis of the egg chamber (this study; Goode et al. 1996b). Rather, the differential requirement of Dlg would appear to reflect an underlying distinction between follicle cells at the poles of the egg chamber. The fields of cells at the poles are distinguished by their expression of *kek-1*, and perhaps by greater intercellular tension caused by their position in a curved region of the epithelium. Dlg may be a component of a signaling/adhesion pathway that helps to establish a spatially uniform rate of cell proliferation by differentially inhibiting cell proliferation at the egg chamber poles.

#### A *Drosophila* model for cellular invasion

How do *dlg* invasive phenotypes compare with normal and pathological cell invasions in other organisms? The cell invasions that we have analyzed have no relevance to those in which cells traverse extracellular matrix, because basement membrane does not exist between either nurse cells or between the follicular epithelium and nurse cells. Rather, our data may be useful for understanding how cells move between heterologous cells during developmental processes such as migration of gonocytes, adult processes such as movement of leukocytes to specific destinations throughout the organism, and during pathological states such as cancer. Whereas most malignant cancer cells traverse an extracellular matrix before reaching their destination, some cancerous cells, particularly in the central nervous system, apparently metastasize strictly through cellular environments, such as axon fascicles (Mareel et al. 1991).

Regardless of interactions with matrices, at a cellular level, normal and pathological cell invasions involve common transitions, including loss of homotypic cell adhesion, epithelial-mesenchymal conversion, gain of heterotypic cell adhesion, and cell migration. In our studies, we have shown that Dlg plays a role in mediating all of these cell behaviors. These cell activities provide a common link between normal and pathological cell invasions, and have led to the suggestion that developmental genes that block invasion are switched off in malignant cells, whereas genes that promote invasion become activated on developmental maturation (Mareel et al. 1991). Although our developmental studies do not make a direct connection to adult malignancy, human Dlg has been implicated in the development of the tumorous state via its association with APC, the most commonly mutated gene in human colon cancer (Matsumine et al. 1996). Further, we have shown that *dlg* invasive cells resemble malignant cells in several regards, including loss of homotypic adhesion, loss of proliferation control, and cell polarity, and enhanced heterotypic interactions and migratory ability. The Dlg invasive phenotype may serve as an excellent genetic and cell biological model for analyzing intercellular interactions underlying cell shape and cell mixing phenomena in a variety of normal and pathological cell invasions.

## Materials and methods

### *Dlg* mutations

*dlg* mutations used in this study have been described previously (Table 1; Perrimon 1988; Woods and Bryant 1989, 1991). All flies were reared on standard media at 25°C or were reared at 18°C, and then shifted to 25°C, as described in the text.

### Histochemistry

Fluorescein-phalloidin staining was performed as described (Goode et al. 1996a). Immunohistochemistry, by use of FITC-conjugated secondary antibodies, was performed as described previously (Goode et al. 1996b). The following sera were used in this study: Dlg (Woods and Bryant 1991), Arm (Peifer et al. 1993), and FasIII (Patel et al. 1987). All fluorescein-labeled tissues were analyzed with a Bio-Rad laser scanning confocal microscope, attached to a Zeiss Axiophot microscope.

### BrdU labeling and analysis of follicle cell division patterns

BrdU was incorporated into ovarian tissue *in vivo*. Females that had been reared at 18°C were placed with an equal number of males for a few days and starved for 2 hr at 18°C (to induce hunger). The flies were then shifted to 25°C onto a yeast paste mixture consisting of 0.5 grams of dry yeast:1.5 ml of BrdU-labeling solution (Amersham). After feeding for 12 hr, females were sacrificed and their ovarian tissue was fixed in Carnoy's fixative, washed in PBT, hydrolyzed in 2 N HCl for 1 hr, and rinsed twice in PBT. Ovarian tissue was then treated with 50 µg/ml of RNase in PBS for 2 hr. Following RNase, the tissue was stained with antibodies to BrdU (Amersham), by use of a FITC-conjugated secondary antibody, as described above. The tissue was then stained for 20 min with propidium iodide and rinsed with PBS. The relative position of follicle cells that had incorporated BrdU was established with a laser scanning confocal microscopy, as described previously (Goode et al. 1996b).

### Clonal analysis

To analyze the phenotypic consequence of introducing *dlg* mutations in germ cells, follicle cells, or both tissues simultaneously, FRT-mediated recombination was induced with a heat shock FLPase (hsFLPase) chromosome (Golic and Lindquist 1989). Clones were marked by loss of a constitutively expressed *lacZ* gene driven by a tubulin promoter, TUBlacZ [(Fig. 6; originally X37 *lacZ* (D. Harrison, pers. comm.)]. A nuclear localization signal is fused to the β-gal protein produced by the X37 *lacZ* gene, thus targeting expression to both germ and follicle cell nuclei. FRT<sup>101</sup> was used in all experiments. The progeny of *dlg* FRT<sup>101</sup>/FM7, B animals crossed to TUBlacZ FRT<sup>101</sup>/Y; hsFLPase/+ animals were heat shocked for 2 hr on consecutive days during early pupal development. Adult B<sup>+</sup> females were allowed to lay eggs for several days before dissecting ovaries. Ovaries were stained for *lacZ* expression by standard methods.

### Western analysis

Western analysis was performed as described (Woods et al. 1996).

## Acknowledgments

Dan Woods provided Dlg sera and stimulating and helpful discussion throughout this study. We thank Kevin Fitzgerald, Denise Montell, and Dan Woods for helpful comments on the

## Goode and Perrimon

manuscript. We thank Beth Noll and Betsy Wilder for technical advice concerning the BrdU labeling protocol and Allan Spradling for helpful discussion concerning follicle cell counts. Doug Harrison kindly provided the X37 tubulin *lacZ* marker. Arm sera were a gift of Mark Peiffer, and FasIII sera were a gift of Nipam Patel. S.G. was supported by a National Institutes of Health postdoctoral fellowship (GM17511-02) and is currently a Leukemia Society of America Special Fellow (3232-98). N.P. is an investigator of the Howard Hughes Medical Institute.

The publication costs of this article were defrayed in part by payment of page charges. This article must therefore be hereby marked "advertisement" in accordance with 18 USC section 1734 solely to indicate this fact.

## References

- Anderson, J.A. 1996. Cell signaling: MAGUK magic. *Curr. Biol.* **4**: 382–384.
- Birchmeier, W. and J. Behrens. 1994. Cadherin expression in carcinomas: Role in the formation of cell junctions and the prevention of invasiveness. *Biochim. Biophys. Acta* **1198**: 11–26.
- Cohen, N.A., J.E. Brenman, S.H. Snyder, and D.S. Bredt. 1996. Binding of the inward rectifier K<sup>+</sup> channel Kir 2.3 to PSD-95 is regulated by protein kinase A phosphorylation. *Neuron* **17**: 759–767.
- Gateff, E. 1978. Malignant neoplasms of genetic origin in *Drosophila melanogaster*. *Science* **200**: 1448–1459.
- Golic, K.G. and S. Lindquist. 1989. The FLP recombinase of yeast catalyzes site-specific recombination in the *Drosophila* genome. *Cell* **59**: 499–509.
- Gonzalez-Reyes, A., H. Elliott, and R.D. St. Johnston. 1995. Polarization of both major body axes in *Drosophila* by *gurken-torpedo* signalling. *Nature* **375**: 654–658.
- Goode, S., D. Wright, and A.P. Mahowald. 1992. The neurogenic locus *brainiac* cooperates with the *Drosophila* EGF receptor to establish the ovarian follicle and to determine its dorsal-ventral polarity. *Development* **116**: 177–192.
- Goode, S., M. Morgan, Y.-P. Liang, and A.P. Mahowald. 1996a. *brainiac* encodes a novel, putative secreted protein that cooperates with *grk* TGF $\alpha$  to produce the follicular epithelium. *Dev. Biol.* **178**: 35–50.
- Goode, S., M. Melnick, T.-B. Chou, and N. Perrimon. 1996b. The neurogenic genes *egghead* and *brainiac* define a novel signaling pathway essential for epithelial morphogenesis during *Drosophila* oogenesis. *Development* **122**: 3863–3879.
- Gumbiner, B.M. 1996. Cell adhesion: The molecular basis of tissue architecture and morphogenesis. *Cell* **84**: 345–357.
- Hynes, R.O. and A.D. Lander. 1992. Contact and adhesive specificities in the associations, migrations, and targeting of cells and axons. *Cell* **68**: 303–322.
- Lahey, T., M. Gorczyca, X.X. Jia, and V. Budnik. 1994. The *Drosophila* tumor suppressor gene *dlg* is required for normal synaptic bouton structure. *Neuron* **13**: 823–835.
- Mahowald, A.P. 1972. Ultrastructural observations on oogenesis in *Drosophila*. *J. Morphol.* **137**: 28–48.
- Mareel, M.M., P. De Baetselier, and F.M. Van Roy. 1991. *Mechanisms of invasion and metastasis*. CRC Press, Boca Raton, FL.
- Matsumine, A., O. Akiko, S. Tadao, N. Okumura, K. Satoh, B. Gyeong-Hun, T. Kawahara, S. Kobayashi, M. Okada, K. Toyoshima, and T. Akiyama. 1996. Binding of APC to the human homolog of the *Drosophila* discs large tumor suppressor protein. *Science* **272**: 1020–1023.
- Mayer, B.J. and M.J. Eck. 1995. SH3 domains. Minding your p's and q's. *Curr. Biol.* **5**: 364–367.
- Montell, D.J. 1994. Moving right along: Regulation of cell migration during *Drosophila* development. *Trends Genet.* **10**: 59–62.
- Montell, D.J., P. Rorth, and A.C. Spradling. 1992. *slow border cells*, a locus required for a developmentally regulated cell migration during oogenesis, encodes *Drosophila* C/EBP. *Cell* **71**: 51–62.
- Murphy, A.M., T. Lee, C.M. Andrews, B.Z. Shilo, and D.J. Montell. 1995. The breathless FGF receptor homolog, a downstream target of *Drosophila* C/EBP in the developmental control of cell migration. *Development* **121**: 2255–2263.
- Musacchio, M. and N. Perrimon. 1996. The *Drosophila* *kekkon* genes: Novel members of both the leucine-rich repeat and immunoglobulin superfamilies expressed in the CNS. *Dev. Biol.* **178**: 63–76.
- Patel, N.H., P.M. Snow, and C.S. Goodman. 1987. Characterization and cloning of fasciclin III: A glycoprotein expressed on a subset of neurons and axon pathways in *Drosophila*. *Cell* **48**: 975–988.
- Peifer, M., S. Orsulic, D. Sweeton, and E. Wieschaus. 1993. A role for the *Drosophila* segment polarity gene *armadillo* in cell adhesion and cytoskeletal integrity during oogenesis. *Development* **118**: 1191–1207.
- Perrimon, N. 1988. The maternal effect of *l(1)discs-large*: A recessive oncogene of *Drosophila melanogaster*. *Dev. Biol.* **127**: 392–407.
- Roth, S., F.S. Neuman-Silberberg, G. Barcelo, and T. Schupbach. 1995. *cornichon* and the EGF receptor signaling process are necessary for both anterior-posterior and dorsal-ventral pattern formation in *Drosophila*. *Cell* **81**: 967–978.
- Sheng, M. 1996. PDZs and receptor/channel clustering: Rounding up the latest suspects. *Neuron* **17**: 575–578.
- Simske, J.S., S.M. Kaech, S.A. Harp, and S.K. Kim. 1996. LET-23 receptor localization by the cell junction protein *lin-7* during *C. elegans* vulval induction. *Cell* **85**: 195–204.
- Songyang, Z., A.S. Fanning, C. Fu, J. Xu, S.M. Marfatea, A.H. Chishti, A. Crompton, A.C. Chan, J.M. Anderson, and L.C. Cantley. 1997. Recognition of unique carboxyl-terminal motifs by distinct PDZ domains. *Science* **275**: 73–77.
- Spradling, A.C. 1993. Developmental genetics of oogenesis. In *The development of Drosophila melanogaster* (ed. M. Bate and M. Arias), pp. 1–70. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- Takeichi, M. 1988. The cadherins: Cell-cell adhesion molecules controlling animal morphogenesis. *Development* **102**: 639–655.
- Woods, D.F. and P.J. Bryant. 1989. Molecular cloning of the *lethal (l) discs large-1* oncogene of *Drosophila*. *Dev. Biol.* **134**: 222–235.
- . 1991. The *discs-large* tumor suppressor gene of *Drosophila* encodes a guanylate kinase homolog localized at septate junctions. *Cell* **66**: 451–464.
- . 1993. ZO-1, DlgA and PSD-95/SAP90: Homologous proteins in tight, septate, and synaptic cell junctions. *Mech. Dev.* **44**: 85–89.
- Woods, D.F., C. Hough, D. Peel, G. Callaini, and P.J. Bryant. 1996. Dlg protein is required for junction structure, cell polarity, and proliferation control in *Drosophila* epithelia. *J. Cell Biol.* **134**: 1469–1482.



## Inhibition of patterned cell shape change and cell invasion by Discs large during *Drosophila* oogenesis

Scott Goode and Norbert Perrimon

*Genes Dev.* 1997, 11:

Access the most recent version at doi:[10.1101/gad.11.19.2532](https://doi.org/10.1101/gad.11.19.2532)

---

### References

This article cites 30 articles, 9 of which can be accessed free at:  
<http://genesdev.cshlp.org/content/11/19/2532.full.html#ref-list-1>

### License

### Email Alerting Service

Receive free email alerts when new articles cite this article - sign up in the box at the top right corner of the article or [click here](#).

---

**CRISPR KO, CRISPRa,  
CRISPRi libraries.**  
Custom or genome-wide.

[VIEW PRODUCTS >](#)

