



Genome Sequences of Three Pseudoalteromonas Strains (P1-8, P1-11, and P1-30), Isolated from the Marine Hydroid Hydractinia echinata

Citation

Klassen, Jonathan L., Maja Rischer, Thomas Wolf, Huijuan Guo, Ekaterina Shelest, Jon Clardy, and Christine Beemelmans. 2015. "Genome Sequences of Three Pseudoalteromonas Strains (P1-8, P1-11, and P1-30), Isolated from the Marine Hydroid Hydractinia echinata." *Genome Announcements* 3 (6): e01380-15.
doi:10.1128/genomeA.01380-15. <http://dx.doi.org/10.1128/genomeA.01380-15>.

Published version

<https://doi.org/10.1128/genomeA.01380-15>

Link

<http://nrs.harvard.edu/urn-3:HUL.InstRepos:23993623>

Terms of use

This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material (LAA), as set forth at

<https://harvardwiki.atlassian.net/wiki/external/NGY5NDE4ZjgzNTc5NDQzMGIzZWZhMGFIOWI2M2EwYTg>

Accessibility

<https://accessibility.huit.harvard.edu/digital-accessibility-policy>

Share Your Story

The Harvard community has made this article openly available.
Please share how this access benefits you. [Submit a story](#)

Genome Sequences of Three *Pseudoalteromonas* Strains (P1-8, P1-11, and P1-30), Isolated from the Marine Hydroid *Hydractinia echinata*

Jonathan L. Klassen,^a Maja Rischer,^b Thomas Wolf,^b Huijuan Guo,^b Ekaterina Shelest,^b Jon Clardy,^c Christine Beemelmans^b

Department of Molecular and Cell Biology, University of Connecticut, Storrs, Connecticut, USA^a; Leibniz Institute for Natural Product Research and Infection Biology e.V., Jena, Germany^b; Department of Biological Chemistry and Molecular Pharmacology, Harvard Medical School, Boston, Massachusetts, USA^c

The genomes of three *Pseudoalteromonas* strains (P1-8, P1-11, and P1-30) were sequenced and assembled. These genomes will inform future study of the genes responsible for the production of biologically active compounds responsible for these strains' antimicrobial, biofouling, and algicidal activities.

Received 5 October 2015 Accepted 23 October 2015 Published 10 December 2015

Citation Klassen JL, Rischer M, Wolf T, Guo H, Shelest E, Clardy J, Beemelmans C. 2015. Genome sequences of three *Pseudoalteromonas* strains (P1-8, P1-11, and P1-30), isolated from the marine hydroid *Hydractinia echinata*. *Genome Announc* 3(6):e01380-15. doi:10.1128/genomeA.01380-15.

Copyright © 2015 Klassen et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Christine Beemelmans, christine.beemelmans@hki-jena.de.

Marine pseudoalteromonads are commonly associated with diverse marine eukaryotic hosts (1, 2) and exhibit a remarkable ability to produce small molecules with a broad range of bioactivities, including antibacterial (3), (anti)biofouling (4, 5), and algicidal (6) activities. We isolated three *Pseudoalteromonas* strains from the tissue of *Hydractinia echinata*, a colonial marine hydroid growing on gastropod shells inhabited by hermit crabs (*Pagurus pollicaris*). Sequencing these strains' genomes will assist the manipulation of *Pseudoalteromonas* genomes, facilitate the discovery and production of new and biologically active molecules (7), and might provide insights into the molecular cues and mechanisms involved in the recruitment and settlement of *H. echinata* larvae (8).

Freshly collected *H. echinata* were purchased from the Marine Biological Laboratory (Woods Hole, MA, USA), and the tissue surface of feeding polyps were investigated for the presence of bacteria from the *Pseudoalteromonas* genus. Clean isolates were cultured in marine broth (Difco 2216) for 3 days at 30°C (150 rpm), and metabolites were extracted using standard solid-phase extraction methods. The resulting organic extracts were tested for antimicrobial activity against a broad range of human pathogenic bacteria and fungi, and showed weak to moderate antimicrobial activity against Gram-positive bacteria (e.g., *Staphylococcus aureus*). Genomic DNA was extracted using the GenElute blood genomic DNA kit (Sigma-Aldrich) according to the manufacturer's protocol. Sequencing was performed at the Harvard Medical School Biopolymers Facility using Illumina TruSeq 50-bp paired-end libraries and a HiSeq2000 instrument (Illumina CASAVA version 1.8.2). A fraction of these reads representing ~50× coverage were assembled using the A5 pipeline version 201401013 (9) and screened for potential contaminations using blobology (10). Genomes were annotated using Prokka version 1.10 (11), and statistics were calculated using scripts from the Assemblathon 2 project (12).

The draft genome of strain P1-8 was sequenced to 50× coverage and comprises 37 contigs in 29 scaffolds, totaling 4,488,653 bases in length and having a G+C content of 41.2%. Its annota-

tion includes 3,992 coding sequences (CDSs), 36 tRNAs, and 3 rRNAs.

The draft genome of strain P1-11 was sequenced to 51× coverage and comprises 44 contigs in 31 scaffolds, totaling 4,377,754 bases in length and having a G+C content of 41.0%. Its annotation includes 3,885 CDSs, 39 tRNAs, and 3 rRNAs.

The draft genome of strain P1-30 was sequenced to 51× coverage and comprises 51 contigs in 35 scaffolds, totaling 4,337,278 bases in length and having a G+C content of 40.9%. Its annotation includes 3,824 CDSs, 36 tRNAs, and 3 rRNAs.

Genes associated with biofilm formation and surface attachments, including genes encoding for *curli*, type II secretion system, type IV pili, and capsular polysaccharide (O-antigen) were identified, reflecting the adaptation to successful persistence and competition on marine surfaces (13). Genes encoding for secondary metabolite production (e.g., alterochromides), bacteriocins, and siderophore function (e.g., desferrioxamines) were detected using antiSMASH (14) and SMIPS (15). These genomes will promote the genetic analysis of the *Pseudoalteromonas* genus and will provide insights into secondary metabolite production and the molecular cues and mechanisms involved in the recruitment and settlement of *H. echinata* larvae (8).

Nucleotide sequence accession numbers. The whole-genome shotgun projects for strains P1-8, P1-11, and P1-30 have been deposited in DDBJ/EMBL/GenBank under the accession numbers [LJSO0000000](https://www.ncbi.nlm.nih.gov/nuccore/LJSO0000000), [LJSP0000000](https://www.ncbi.nlm.nih.gov/nuccore/LJSP0000000), and [LKBC0000000](https://www.ncbi.nlm.nih.gov/nuccore/LKBC0000000), respectively. The versions described in this paper are the first versions, [LJSO0100000](https://www.ncbi.nlm.nih.gov/nuccore/LJSO0100000), [LJSP0100000](https://www.ncbi.nlm.nih.gov/nuccore/LJSP0100000), and [LKBC0100000](https://www.ncbi.nlm.nih.gov/nuccore/LKBC0100000).

ACKNOWLEDGMENTS

We are grateful for financial support from the NIH to J.C. (GM086258) and the German National Academy of Sciences Leopoldina for a postdoctoral fellowship to C.B. (LPDS 2011-2). J.L.K. was supported by funds from the University of Connecticut. M.R. was supported by the graduate school Jena School for Microbial Communication (JSMC), financed by the Deutsche Forschungsgemeinschaft, and T.W. was supported by the

International Leibniz Research School for Microbial and Molecular Interactions (ILRS), as part of the JSMC.

REFERENCES

- Gauthier G, Gauthier M, Christen R. 1995. Phylogenetic analysis of the genera *Alteromonas*, *Shewanella*, and *Moritella* using genes coding for small-subunit rRNA sequences and division of the genus *Alteromonas* into two genera, *Alteromonas* (emended) and *Pseudoalteromonas* gen. nov., and proposal of twelve new species combinations. *Int J Syst Bacteriol* 45:755–761. <http://dx.doi.org/10.1099/00207713-45-4-755>.
- Holmström C, Kjelleberg S. 1999. Marine *Pseudoalteromonas* species are associated with higher organisms and produce biologically active extracellular agents. *FEMS Microbiol Ecol* 30:285–293. [http://dx.doi.org/10.1016/S0168-6496\(99\)00063-X](http://dx.doi.org/10.1016/S0168-6496(99)00063-X).
- Bowman JP. 2007. Bioactive compound synthetic capacity and ecological significance of marine bacterial genus *Pseudoalteromonas*. *Mar Drugs* 5:220–241. <http://dx.doi.org/10.3390/md504220>.
- Qian P-Y, Lau SCK, Dahms H-U, Dobretsov S, Harder T. 2007. Marine biofilms as mediators of colonization by marine macroorganisms: implications for antifouling and aquaculture. *Mar Biotechnol* 9:399–410. <http://dx.doi.org/10.1007/s10126-007-9001-9>.
- Holmström C, Egan S, Franks A, McCloy S, Kjelleberg S. 2002. Antifouling activities expressed by marine surface associated *Pseudoalteromonas* species. *FEMS Microbiol Ecol* 41:47–58. [http://dx.doi.org/10.1016/S0168-6496\(02\)00239-8](http://dx.doi.org/10.1016/S0168-6496(02)00239-8).
- Lovejoy C, Bowman JP, Hallegraef GM. 1998. Algicidal effects of a novel marine *Pseudoalteromonas* isolate (class *Proteobacteria*, gamma subdivision) on harmful algal bloom species of the genera *Chattonella*, *Gymnodinium*, and *Heterosigma*. *Appl Environ Microbiol* 64:2806–2813.
- Machado H, Sonnenschein EC, Melchiorson J, Gram L. 2015. Genome mining reveals unlocked bioactive potential of marine gram-negative bacteria. *BMC Genomics* 16:158–170. <http://dx.doi.org/10.1186/s12864-015-1365-z>.
- Frank U, Leitz T, Müller WA. 2001. The hydroid *Hydractinia*: a versatile, informative cnidarian representative. *Bioessays* 23:963–971. <http://dx.doi.org/10.1002/bies.1137>.
- Coil D, Jospin G, Darling AE. 2015. A5-miseq: an updated pipeline to assemble microbial genomes from Illumina MiSeq data. *Bioinformatics* 31:587–589. <http://dx.doi.org/10.1093/bioinformatics/btu661>.
- Kumar S, Jones M, Koutsovoulos G, Clarke M, Blaxter M. 2013. Blobology: exploring raw genome data for contaminants, symbionts and parasites using taxon-annotated GC-coverage plots. *Front Genet* 4:237. <http://dx.doi.org/10.3389/fgene.2013.00237>.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <http://dx.doi.org/10.1093/bioinformatics/btu153>.
- Bradnam KR, Fass JN, Alexandrov A, Baranay P, Bechner M, Birol I, Boisvert S, Chapman JA, Chapuis G, Chikhi R, Chitsaz H, Chou W, Corbeil J, Del Fabbro C, Docking T, Durbin R, Earl D, Emrich S, Fedotov P, Fonseca NA, Ganapathy G, Gibbs RA, Gnerre S, Godzaridis E, Goldstein S, Haimel M, Hall G, Haussler D, Hiatt JB, Ho IY, Howard J, Hunt M, Jackman SD, Jaffe DB, Jarvis ED, Jiang H, Kazakov S, Kersey PJ, Kitzman JO, Knight JR, Koren S, Lam T-W, Lavenier D, Laviolette F, Li Y, Li Z, Lio B, Liu Y, Luo R, MacCallum I, MacManes MD, Maillat N, Melnikov S, Naquin D, Ning Z, Otto TD, Paten B, Paulo OS, Phillippy AM, Pina-Martins F, Place M, Przybylski D, Qin X, Qu C, Ribeiro FJ, Richards S, Rokhsar DS, Ruby JG, Scalabrin S, Schatz MC, Schwartz DC, Sergushichev A, Sharpe T, Shaw TI, Shendure J, Shi Y, Simpson JT, Song H, Tsarev F, Vezzi F, Vicedomini R, Vieira BM, Wang J, Worley KC, Yin S, Yiu S-M, Yuan J, Zhang G, Zhang H, Zhou S, Korff IF. 2013. Assemblathon 2: Evaluating *de novo* methods of genome assembly in three vertebrate species. *GigaScience* 2:10. <http://dx.doi.org/10.1186/2047-217X-2-10>.
- Thomas T, Evans FF, Schleheck D, Mai-Prochnow A, Burke C, Penevyan A, Dalisay DS, Stelzer-Braid S, Saunders N, Johnson J, Ferreira S, Kjelleberg S, Egan S. 2008. Analysis of the *Pseudoalteromonas tunicata* genome reveals properties of a surface-associated life style in the marine environment. *PLoS One* 3:e3252. <http://dx.doi.org/10.1371/journal.pone.0003252>.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Brucoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. *Nucleic Acids Res* 43:W237–W243. <http://dx.doi.org/10.1093/nar/gkv437>.
- Wolf T, Shelest V, Nath N, Shelest E. 2015. CASSIS and SMIPS—promoter-based prediction of secondary metabolite gene clusters in eukaryotic genomes. Manuscript under revision. Program available at <https://sbi.hki-jena.de/smips>.