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Accessibility
Glucocorticoid-induced TNF receptor family-related protein ligand is requisite for optimal functioning of regulatory CD4+ T cells

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INTRODUCTION

CD4+ CD25+ FoxP3+ regulatory T cells (Treg), which develop in the thymus or can be induced in peripheral organs, control many aspects of the immune response (1–4). Tregs constitutively express glucocorticoid-induced tumor necrosis factor receptor family-related protein ligand (GITR-L), which is expressed by antigen presenting cells, on the development and expansion of Tregs. We found that GITR-L is dispensable for the development of naturally occurring FoxP3+ Treg cells in the thymus. However, the expansion of Treg in GITR-L−/− mice is impaired after injection of the dendritic cells (DCs) inducing factor Flt3 ligand. Furthermore, DCs from the liver of GITR-L−/− mice were less efficient in inducing proliferation of antigen-specific Treg cells in vitro than the same cells from WT littermates. Upon gene transfer of ovalbumin into hepatocytes of GITR-L−/−FoxP3(GFP) reporter mice using adeno-associated virus (AAV8-OVA) the number of antigen-specific Treg in liver and spleen is reduced. The reduced number of Tregs resulted in an increase in the number of ovalbumin specific CD8+ T effector cells. This is highly significant because proliferation of antigen-specific CD8+ cells itself is dependent on the presence of GITR-L, as shown by in vivo experiments and by adoptive transfers into GITR-L−/− Rag-1−/− and Rag-1−/− mice that had received AAV8-OVA. Surprisingly, administering αCD3 significantly reduced the numbers of FoxP3+ Treg cells in the liver and spleen of GITR-L−/− but not WT mice. Because soluble Fc-GITR-L partially rescues αCD3 induced in vitro depletion of the CD103+ subset of FoxP3+ CD4+ Treg cells, we conclude that expression of GITR-L by antigen presenting cells is requisite for optimal Treg-mediated regulation of immune responses including those in response during gene transfer.

Keywords: GITR-L, TNFSF18, Flt3L, Treg, CX3CR1

Glucocorticoid-induced tumor necrosis factor receptor family-related protein (TNFRSF18, CD357) is constitutively expressed on regulatory T cells (Tregs) and is inducible on effector T cells. In this report, we examine the role of glucocorticoid-induced TNF receptor family-related protein ligand (GITR-L), which is expressed by antigen presenting cells, on the development and expansion of Tregs. We found that GITR-L is dispensable for the development of naturally occurring FoxP3+ Treg cells in the thymus. However, the expansion of Treg in GITR-L−/− mice is impaired after injection of the dendritic cells (DCs) inducing factor Flt3 ligand. Further, DCs from the liver of GITR-L−/− mice were less efficient in inducing proliferation of antigen-specific Treg cells in vitro than the same cells from WT littermates. Upon gene transfer of ovalbumin into hepatocytes of GITR-L−/−FoxP3(GFP) reporter mice using adeno-associated virus (AAV8-OVA) the number of antigen-specific Treg in liver and spleen is reduced. The reduced number of Tregs resulted in an increase in the number of ovalbumin specific CD8+ T effector cells. This is highly significant because proliferation of antigen-specific CD8+ cells itself is dependent on the presence of GITR-L, as shown by in vivo experiments and by adoptive transfers into GITR-L−/−Rag-1−/− and Rag-1−/− mice that had received AAV8-OVA. Surprisingly, administering αCD3 significantly reduced the numbers of FoxP3+ Treg cells in the liver and spleen of GITR-L−/− but not WT mice. Because soluble Fc-GITR-L partially rescues αCD3 induced in vitro depletion of the CD103+ subset of FoxP3+ CD4+ Treg cells, we conclude that expression of GITR-L by antigen presenting cells is requisite for optimal Treg-mediated regulation of immune responses including those in response during gene transfer.
(21). TGF-β, a cytokine highly expressed in mucosal tissues and sites of inflammation, plays a role in conversion of conventional peripheral CD4+ T cells into Treg, and TGF-β up-regulates expression of CD103 (Integrin αEβ7) (22), which is the primary ligand of E-cadherin, an epithelial adhesion molecule. Expression of CD103 marks a subset of peripheral inducible Tregs (about 20–30% of the CD4+FoxP3+ Tregs in the spleen), which inhibit graft-versus-host disease more potently than the CD4+CD25+ Tregs (23, 24).

In this study, we provide evidence in support of the concept that the interactions between GITR and GITR-L are requisite for optimal functioning of Tregs. To this end, we analyze GITR-L−/− FoxP3(GFP) and GITR-L+/− CX3CR1(GFP) mice after gene transfer of ovalbumin into hepatocytes with adeno-associated virus (AAV8-OVA). Coordinate expansion of Treg and dendritic cells (DCs) was assessed after injection of ligand in GITR-L−/− mice. The interactions between antigen presenting cells and Tregs are also evaluated after administering αCD3 in GITR-L−/− mice or by co-activation with αCD3 and soluble Fc-GITR-L.

**MATERIALS AND METHODS**

**MICE**

B6, OT-II Tg, and CX3CR1(GFP) reporter mice were purchased from the Jackson Laboratory (Bar Harbor, ME, USA). OT-I × Rag−/− mice were purchased from Taconic Labs (Germantown, NY, USA). GITR-L−/− and FoxP3-IRE-EGF-SV40 knock-in (FoxP3(GFP)) B6 mice were described previously (8, 25). GITR-L−/− mice were crossed with FoxP3(GFP) and CX3CR1(GFP) mice to generate GITR-L−/−FoxP3(GFP) and GITR-L+/−CX3CR1(GFP) B6 mice. All animals were housed in the Center for Life Science animal facility of BIDMC. The Guide for the Care and Use of Laboratory Animals was followed in the conduct of the animal studies of the Institutional Animal Care and Use Committee at BIDMC. Veterinary care was given to any animals requiring medical attention.

**ANTIBODIES**

Anti-CD11b-PacBlu, αCD11b-FITC, αCD4-PE, αCD4-APC, αCD11c-APC, αCD11c-PE, αTCRvα2-PE, and αCD3s(145–2C11) were purchased from BioLegend (San Diego, CA, USA). Anti-Ly6C-PerCP and αFoxP3-APC were products of eBioscience (San Jose, CA, USA). Anti-Ly6G-PE, αNK1.1-PE, αCD8α-PacBlu, αCD25-PE, and αCD103-Alexa Fluor 647 were products from BD Biosciences (San Jose, CA, USA). Flt3L-Fc fusion protein was purchased from BioXCell (West Lebanon, NH, USA). Anti-IL-2 was purchased from R&D Systems (Minneapolis, MN, USA). Fc-GITR-L fusion protein was produced as described previously (9).

**AAV8-OVA MEDIATED expression of FOREIGN PROTEIN in HEPATOCYTES**

AAV8-OVA vector (containing an ovalbumin expression cassette driven by AAV-EF1α) was packaged into serotype 8 capsid as described previously (16). Vector was injected i.v. into FoxP3(GFP) and GITR-L−/−FoxP3(GFP) mice at a dose of 10^10 vector genome/mouse. Five weeks later, leukocytes from liver, spleen, and thymus were stained with TCRvα2.

Also, Ly6G NK1.1+ cells FACS sorted from the liver of CD3ε−/− FoxP3+ (GFP) mice 7 days after AAV8-OVA injection were incubated with anti-CD4+ or CSF1-labeled OT-I CD8+ T cells for 3 days. OT-I CD4+ T cell cultures were stained with TCRvα2 and FoxP3. OT-I CD8+ T cell culture was stained with TCRvα2 and proliferating CD8+ cells were evaluated by CFSE dilution.

**INDUCTION OF DENDRITIC CELLS AND TREG WITH Flt3L**

Flt3L-Fc fusion protein (10 ng/mouse/injection) was i.p. injected into FoxP3(GFP) and GITR-L−/−FoxP3(GFP) mice for nine consecutive days as described previously (26). Leukocytes from the spleen and liver were analyzed at day 10.

**CELLULARITY IN MICE AFTER αCD3-MEDIATED ACtivation of T CELLS BY IN VIVO**

Anti-CD3ε was i.p. injected into CX3CR1(GFP) and GITR-L−/−CX3CR1(GFP) mice (20 µg/mouse, one injection). After 72 h, leukocytes of the spleen and liver were stained with CD4 and FoxP3. CX3CR1+ cells were evaluated by expression of the reporter gene GFP.

**IN VITRO ACTivation of CD4+ T CELLS**

CD4+ T cells from the spleen of FoxP3(GFP) mice were negatively selected using a CD4+ T cells isolation kit (Miltenyi, Auburn, CA, USA) and were activated with αCD3-coupled microbeads in a round bottom 96-well plate in the presence or absence of Flt3L (1 µg/ml) for 2 days as described previously (9). Cells were stained with CD4 and CD103. Expression of FoxP3 was judged by the reporter gene EGFP. Cell numbers were counted with a Countess Automated Cell Counter (Invitrogen, Grand Island, NY, USA).

**ISOLATION OF LIVER leukocytes**

Liver leukocytes were isolated as described previously (27). Briefly, liver was washed and filtered through a 70 µM cell strainer. Hepatocytes and cell debris were removed by spinning at 300 rpm for 10 min. Supernatant was centrifuged at 1500 rpm for 10 min to collect cells. Leukocytes were isolated from the interface of a 40 and 70% Percoll gradient.

Statistical analysis used Prism 4.0c software (GraphPad, San Diego, CA, USA). Statistical comparisons were performed using the two-tailed Student’s t-test. Values of P < 0.05 were considered to be statistically significant.

**RESULTS**

Flt3L-Induced expansion of Treg was impaired in GITR-L deficient mice due to a partially reduced number of dendritic cell subpopulations

We previously found that after administering a Fc-GITR-L fusion protein to WT mice the number of Treg cells increased, which was confirmed by studies with GITR-L transgenic mice (9–11, 28). Surprisingly, we found that GITR-L was dispensable for the development of naturally occurring Treg, as the number of FoxP3+ Treg cells was normal in the thymus and spleen of GITR-L−/−FoxP3(GFP) mice under resting conditions (Figure 1A; Figure S1 in Supplementary Material).
After the injection of Fc-Flt3L fusion protein, both the numbers of pDCs in GITR-L WT their ial, the percentage of CD11c+ T cells from the thymus, spleen, and/or liver were analyzed by FACS for FoxP3 expression based on the expression of reporter protein EGFP. The number of CD11c+FoxP3+ T cells in the spleen and liver of FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice without stimulation. Percentages of CD11c+ and representative staining of Treg in the spleen and liver after administering Flt3L. Flt3L−/− FoxP3(GFP) mice after administering Flt3L. Filled circle represents FoxP3(GFP) mouse. Open circle represents GITR-L−/− FoxP3(GFP) mouse. Each circle represents one mouse.

To further investigate the role of GITR-L in controlling Treg development, we assessed the consequences of injecting Fms-related tyrosine kinase 3 ligand (Flt3L) into FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice for nine consecutive days. Not only is Flt3L a potent inducer of DC and macrophage proliferation (26, 29), several phagocyte subpopulations express GITR-L (12, 30). Indeed, the proliferation of CD8+ cells was affected by the absence of GITR-L from the surface of these DCs. Since activated CD8+ cells carry GITR on their surface, we also directly test whether the absence of GITR-L in DC subpopulations affects proliferation of antigen-specific GITR+ Treg and CD8+ cells, we immunized GITR-L−/−CX3CR1(GFP) and WT CX3CR1(GFP) mice by gene transfer with AA V8-OVA (Figure 3A). One week after injection of AA V8-OVA, liver CX3CR1(GFP)+ cells purified by FACS were incubated with OVA-specific OT-II CD4+ T cells or OT-I CD8+ cells for 3 days. GITR-L−/−CX3CR1+ cells were less efficient in inducing Treg as compared to the same cells isolated from WT mice (Figures 3B,C,D). Taken together, these data indicate that after Flt3L induction, GITR-L affects the expansion and differentiation of subpopulations of DCs, which in turn leads to expansion of Tregs.

GITR-L−/−CX3CR1+ DCs ISOLATED FROM THE LIVER ARE LESS EFFICIENT THAN WT CX3CR1+ DCs IN THE IN VITRO INDUCTION OF OVA-SPECIFIC TREG AND CD8+ T CELLS

To directly test whether the absence of GITR-L in DC subpopulations affects proliferation of antigen-specific GITR+ Treg and CD8+ cells, we immunized GITR-L−/−CX3CR1(GFP) and WT CX3CR1(GFP) mice by gene transfer with AA V8-OVA (Figure 3A). One week after injection of AA V8-OVA, liver CX3CR1(GFP)+ cells purified by FACS were incubated with OVA-specific OT-II CD4+ T cells or OT-I CD8+ cells for 3 days. GITR-L−/−CX3CR1+ cells were less efficient in inducing Treg as compared to the same cells isolated from WT mice (Figures 3B,C,D). Taken together, these data indicate that after Flt3L induction, GITR-L affects the expansion and differentiation of subpopulations of DCs, which in turn leads to expansion of Tregs.
Impair the induction of antigen-specific Tregs (16–18, 21, 33), which may at least partially compromise their immunosuppressive capability.

As the in vitro data suggest that GITR-L expression on DCs causes the expansion of CD8+ cells, this in vivo result might underestimate the consequences of the reduced number of the Tregs in the GITR-L−/− mice. To test whether GITR-L is implicated in the in vivo expansion of antigen-specific CD8+ cells, we used a system in which the Treg-mediated suppression is absent. To this end, we injected AA V8-OVA into Rag−/− and GITR-L−/− Rag−/− mice followed by the adoptive transfer of OT-I CD8+ T cells. Eight weeks after transfer of OT-I CD8+ T cells, the number of CD8+ T cells in the blood of the GITR-L−/− Rag−/− recipients was significantly lower than that of the Rag−/− recipients (Figure 5B). This was not due to an inadequate amount of OVA antigen production in the GITR-L−/− Rag−/− recipients (Figure 5C). Taken together, the data indicate that GITR-L is required for optimal induction and/or expansion of antigen-specific Treg in the context of hepatic AA V8 gene transfer.

**DEPLETION OF CX3CR1+ (GFP) CELLS BY αCD3 IN GITR-L−/− MICE CORRELATES WITH A REDUCED NUMBER OF FOXP3+ TREG CELLS**

*In vitro* expansion of FoxP3+ Treg cells can be achieved by stimulation with a combination of αCD3 and soluble GITR-L (Fc-GITR-L) (9). We then assessed whether injection of αCD3 into WT and GITR-L−/− mice would affect the Treg population. As shown in Figures 6A,B, αCD3 induced a significant reduction of the percentage of FoxP3+ Treg in the spleen and liver of GITR-L−/− CX3CR1(GFP) mice, but not in WT CX3CR1(GFP) mice. In support of our observations in this paper, the reduced number of Tregs coincided with a reduction of CX3CR1+ DCs in the spleen.

**FIGURE 3 | In vitro induction of OVA-specific Treg and CD8+ T cells with hepatic CX3CR1+ DCs**

(A) Schematic for in vitro priming of CD8+ OT-I and CD4+ OT-II T cells. Briefly, AA V8-OVA(10^6 vector genome/mouse) was i.v. injected into CX3CR1(GFP) and GITR-L−/− CX3CR1(GFP) mice. After 7 days, Ly6G-NK1.1-CX3CR1(GFP) cells were purified from the liver and incubated with CD8+ OT-I and CD4+ OT-II T cells at different ratios for 3 days. Divisions of CD8+ OT-I cells were evaluated by CFSE dilution. (B) Percentages of OT-I T cells. Consistent with the results support of our observations in this paper, the reduced number of the Tregs in CX3CR1(GFP) mice (Figure 4A). Conversely, AA V8-OVA vector into in FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice and determined the number of OVA-specific Treg and CD8+ T cells. Consistent with the results when administering Flt3L, there was a reduced percentage of OVA-specific FoxP3+ TCRva2+ T cells in the spleen and liver of GITR-L−/− FoxP3(GFP) mice as compared to that of WT mice 5 weeks after vector administration (Figure 4A). Conversely, AA V8-OVA vector into in FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice and determined the number of OVA-specific Treg and CD8+ T cells. Consistent with the results when administering Flt3L, there was a reduced percentage of OVA-specific FoxP3+ TCRva2+ T cells in the spleen and liver of GITR-L−/− FoxP3(GFP) mice as compared to that of WT mice 5 weeks after vector administration (Figure 4A). Conversely, AA V8-OVA vector into in FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice and determined the number of OVA-specific Treg and CD8+ T cells. Consistent with the results when administering Flt3L, there was a reduced percentage of OVA-specific FoxP3+ TCRva2+ T cells in the spleen and liver of GITR-L−/− FoxP3(GFP) mice as compared to that of WT mice 5 weeks after vector administration (Figure 4A). Conversely, AA V8-OVA vector into in FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice and determined the number of OVA-specific Treg and CD8+ T cells. Consistent with the results when administering Flt3L, there was a reduced percentage of OVA-specific FoxP3+ TCRva2+ T cells in the spleen and liver of GITR-L−/− FoxP3(GFP) mice as compared to that of WT mice 5 weeks after vector administration (Figure 4A). Conversely, AA V8-OVA vector into in FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice and determined the number of OVA-specific Treg and CD8+ T cells. Consistent with the results when administering Flt3L, there was a reduced percentage of OVA-specific FoxP3+ TCRva2+ T cells in the spleen and liver of GITR-L−/− FoxP3(GFP) mice as compared to that of WT mice 5 weeks after vector administration (Figure 4A). Conversely, AA V8-OVA vector into in FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice and determined the number of OVA-specific Treg and CD8+ T cells. Consistent with the results when administering Flt3L, there was a reduced percentage of OVA-specific FoxP3+ TCRva2+ T cells in the spleen and liver of GITR-L−/− FoxP3(GFP) mice as compared to that of WT mice 5 weeks after vector administration (Figure 4A). Conversely, AA V8-OVA vector into in FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice and determined the number of OVA-specific Treg and CD8+ T cells. Consistent with the results when administering Flt3L, there was a reduced percentage of OVA-specific FoxP3+ TCRva2+ T cells in the spleen and liver of GITR-L−/− FoxP3(GFP) mice as compared to that of WT mice 5 weeks after vector administration (Figure 4A). Conversely, AA V8-OVA vector into in FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice and determined the number of OVA-specific Treg and CD8+ T cells. Consistent with the results when administering Flt3L, there was a reduced percentage of OVA-specific FoxP3+ TCRva2+ T cells in the spleen and liver of GITR-L−/− FoxP3(GFP) mice as compared to that of WT mice 5 weeks after vector administration (Figure 4A). Conversely, AA V8-OVA vector into in FoxP3(GFP) And cell number (x10^6) of FoxP3(GFP) and GITR-L−/− FoxP3(GFP) mice. Each circle represents one mouse.
and liver of GITR−/−CX3CR1(GFP) mice (Figures 6C,D). In contrast, the numbers of CX3CR1+ cells in the spleen and liver were comparable in the two mouse strains under homeostasis (Figure S3 in Supplementary Material).

To further investigate the role of GITR-L in the expansion of FoxP3+ Treg, CD4+ T cells were purified from the spleen of FoxP3(GFP) mice and stimulated in vitro with αCD3 with either Fc-GITR-L or IgG. Forty-eight hours after exposure to αCD3, the number of total CD4+ and FoxP3+CD4+ Treg was significantly higher in the presence of Fc-GITR-L than that of IgG (Figures 7A,B). Interestingly, a subset of CD103+ Treg cells, which is induced in epithelium and in sites of inflammation (23, 34) and comprises approximately 20% of all FoxP3+ Treg cells in the spleen, was also expanded by Fc-GITR-L (Figures 7C,D).

We conclude that while the induction or expansion of Treg is impaired in the absence of GITR-L, Fc-GITR-L provides a positive signal to GITR+ Treg.

**DISCUSSION**

The receptor-ligand pair GITR/GITR-L (TNFRSF18/TNFSF18) appears to be involved in the development of a variety of inflammation-related diseases in murine models (6, 8, 12, 35, 36). It was originally thought that the suppressor function of Treg cells, which constitutively express GITR, would be abrogated by anti-GITR thus breaking immune self-tolerance (2). More recent additional evidence shows that GITR engagement by its natural ligand GITR-L causes an extensive expansion of functionally competent Tregs (9–11), although the relative role of GITR on Treg and Teff cells remains only partly understood. In this study we find that in the absence of GITR-L the expansion of FoxP3+ Treg cells is impaired in an antigen-specific manner, which can be mimicked by in vivo and in vitro activation of CD4+ Treg cells with αCD3. Our results are consistent with the findings of the Chatila group that expansion and contraction of Teff and Treg dynamically control primary immune responses to foreign antigen (25).

Glucocorticoid-induced TNF receptor family-related protein ligand impacts immune regulation in gene replacement therapy at least at three levels. First, the induction/expansion of antigen-specific Treg cells in the liver after AAV-mediated gene therapy is impaired directly by the absence of GITR-L. Second, the expansion of antigen-specific CD8+ T cells is reduced by GITR-L deficiency. However, impaired expansion of Treg cells can on the other hand up-regulate CD8+ T cell expansion indirectly. Third, GITR-L deficiency affects the infiltration of monocyte-derived MØ to the sites where exogenous protein is expressed and/or the sites of inflammation (30), which changes the local function of different immune cells. These GITR-L-expressing, monocyte-derived MØ may provide a microenvironment for the expression of CD103 in Treg cells, an integrin that facilitates the retention of Treg cells in the sites of inflammation or infection.

Surprisingly, we found that administering αCD3 causes the depletion of CX3CR1+ DCs in the spleen and liver of GITR−/−.
mice, which correlates with a reduced number of FoxP3^+ Tregs. It is reported that IL10-secreting GITR^+ T1r1 cells may suppress immune responses by granzyme B-mediated killing of myeloid APCs (37, 38). Granzyme B is also important for the ability of +
cytotoxicity. Depletion of CX3CR1 hi (41). The mechanism how this Ly6C subpopulation is educated to be either protagonist or antagonist is still not well understood. Anti-CD3-mediated depletion of CX3CR1 DCs in the liver may provide an important tool for the study of migration, colonization, and education of this special DC subset (30).

In conclusion, our data show that GITR and GITR-L have important implications for gene therapy. Optimal induction of an immune regulatory response, which is crucial for tolerance to the transgene product and for immune modulatory gene therapy, requires co-stimulation by GITR-L, which enhances Treg induction and function. Expression of GITR-L on hepatic APCs may in part explain the tolerogenic/Treg inducing capacity of hepatic gene transfer.

AUTHOR CONTRIBUTIONS
Gongxian Liao performed all the experiments; Michael S. O’Keeffe helped in processing the samples and editing the manuscript; Guoxing Wang and Boaz van Driel helped in processing the samples and discussing the results. Rene de Waal Malefyt generated GITR-L deficient mice; Hans-Christian Reinecker brought deeper insight into αCD3-inducing murine model. Roland W. Herzog helped in discussing and writing the manuscript; Cox Terhorst is the major organiser of this work and designed the experiments with Gongxian Liao.

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SUPPLEMENTARY MATERIAL
The Supplementary Material for this article can be found online at http://www.frontiersin.org/Journal/10.3389/fimmu.2014.00035/abstract

Figure S1 | Representative staining of Figure 1A.
Figure S2 | (A,B) Representative staining of Figures 2A,B
Figure S3 | CX3CR1(GFP)^+ phagocytes in spleen and liver leukocytes of CX3CR1(GFP)^+ and GITR-L^−/− CX3CR1(GFP)^+ mice under resting condition.

REFERENCES


