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Accessibility
Ly-6Ch	extsuperscript{hi} monocytes dominate hypercholesterolemia-associated monocytosis and give rise to macrophages in atheroma

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Macrophage accumulation participates decisively in the development and exacerbation of atherosclerosis. Circulating monocytes, the precursors of macrophages, display heterogeneity in mice and humans, but their relative contribution to atherogenesis remains unknown. We report here that the Ly-6Ch	extsuperscript{hi} monocyte subset increased dramatically in hypercholesterolemic apoE–/– deficient mice consuming a high-fat diet, with the number of Ly-6Ch	extsuperscript{hi} cells doubling in the blood every month. Ly-6Ch	extsuperscript{hi} monocytes adhered to activated endothelium, infiltrated lesions, and became lesional macrophages. Hypercholesterolemia-associated monocytosis (HAM) developed from increased survival, continued cell proliferation, and impaired Ly-6Ch	extsuperscript{lo} to Ly-6Ch	extsuperscript{hi} conversion and subsided upon statin-induced cholesterol reduction. Conversely, the number of Ly-6Ch	extsuperscript{lo} cells remained unaffected. Thus, we believe that Ly-6Ch	extsuperscript{hi} monocytes represent a newly recognized component of the inflammatory response in experimental atherosclerosis.

Introduction
During atherogenesis, an inflammatory process, leukocytes and lipids accumulate in the aortic intima (1, 2). Lipid-rich macrophages, known as foam cells in atheromas, secrete inflammatory mediators that stimulate smooth muscle cell migration and proliferation and participate in plaque development and rupture as well as thrombosis. Serum C-reactive protein and other molecular mediators of inflammation have broadened our understanding of the disease by illustrating that peripheral blood can afford valuable prognostic information (3, 4). Leukocyte counts rise in atherosclerotic patients. Yet, the extent to which circulating leukocyte subsets reflect the inflammatory response during atherogenesis remains less defined (5–9). This study tested the hypothesis that leukocyte heterogeneity in atherosclerosis could provide novel markers of and mechanistic insights into atherogenesis.

Prevailing concepts view monocytes as intermediary cells that continuously develop in the bone marrow, circulate in the bloodstream, and migrate unselected into tissue, where they become macrophages, dendritic cells, or other tissue descendants (10–12). Studies in atherosclerotic mice have shown that bone marrow–derived circulating monocytes populate atherosclerotic lesions (13–17), and many studies support an active role for monocytes/macrophages in atherosclerosis (reviewed in refs. 1, 2). The appreciation of monocyte heterogeneity (18–20) has led to the hypothesis that monocytes commit for specific functions while still in the circulation. Indeed, both human and mouse monocytes fall into at least 2 phenotypically distinct subsets: Ly-6Ch	extsuperscript{hi} (which are also phenotypically Gr-1 CCR2 CX3CR1hi) and Ly-6Ch	extsuperscript{lo} (which are...
Figure 1
Hypercholesterolemia induces peripheral blood Ly-6C\(^{hi}\) monocytopoiesis. (A) Mononuclear cells from blood of apoE\(^{+/+}\) and apoE\(^{-/-}\) mice consuming either chow or Western diet were stained with anti-CD11b, -CD90, -B220, -CD49b, -NK1.1, -Ly-6G, and -Ly-6C mAbs. Living cells were gated to determine presence and percentage of CD11b\(^{hi}\)CD90\(^{lo}\)B220\(^{lo}\)CD49b\(^{lo}\)NK1.1\(^{lo}\)Ly-6G\(^{lo}\) monocytes (top row) and further divided into Ly-6C\(^{hi}\) and Ly-6C\(^{lo}\) subsets (bottom row). Representative dot plots and histograms from individual mice are depicted. Percentages of cells are shown as mean ± SEM. (B) Total blood monocytes in apoE\(^{+/+}\) and apoE\(^{-/-}\) mice consuming either Western diet (+) or chow (−). (C) Total blood Ly-6C\(^{hi}\) monocytes. (D) Total blood Ly-6C\(^{lo}\) monocytes. (E) Total peripheral blood leukocytes. (F) Representative dot plots depicting expression of CD62L and CD11c among Ly-6C\(^{hi}\) and Ly-6C\(^{lo}\) monocytes. Percentages of cells in each quadrant are shown as mean ± SEM. (G) Representative cytospin preparations of purified blood Ly-6C\(^{hi}\) and Ly-6C\(^{lo}\) monocytes in apoE\(^{+/+}\) mice on chow and apoE\(^{-/-}\) mice on Western diet. Scale bar: 10 µm. Student's t test was used. Results are representative of 8 independent experiments with 5–14 mice per group.

0.27 ± 0.03 × 10\(^6\) cells/ml; Western diet, 0.96 ± 0.11 × 10\(^6\) cells/ml) and lymphocytes decreased slightly (chow, 2.4 ± 0.6 × 10\(^6\) cells/ml; Western diet, 2.0 ± 0.2 × 10\(^6\) cells/ml). As expected (25, 26), apoE\(^{-/-}\) mice on Western diet had increased serum cholesterol levels (479 ± 20 mg/dl) when compared with apoE\(^{+/+}\) mice on chow (286 ± 25 mg/dl). Macroscopic and histologic examination of aortas revealed fatty streaks and fibrous plaque lesions in the root and descending aorta of apoE\(^{-/-}\) mice regardless of diet. Lesions were identified along the entire aorta in older mice (i.e., mice that consumed Western diet for 50 weeks; data not shown). apoE\(^{-/-}\) mice consuming Western diet had 3–5 times more extended and widespread atherosclerotic lesions than did apoE\(^{+/+}\) mice on chow.
apoE<sup>−/−</sup> mice fed a Western diet had serum cholesterol levels of 227 ± 39 mg/dl, lower than those observed in apoE<sup>−/−</sup> mice on Western diet but higher than those in apoE<sup>+/+</sup> mice on chow (101 ± 10 mg/dl).

apoE<sup>−/−</sup> mice fed a Western diet did not show a significant increase in the number of circulating monocytes or Ly-6C<sub>hi</sub> monocytes (Figure 1, B–D) and did not develop atherosclerotic lesions during the 25 weeks of diet consumption (data not shown).

apoE<sup>−/−</sup> mice also had elevated numbers of CD11b<sup>+</sup>CD90<sup>−/−</sup>B220<sup>−</sup>CD49b<sup>−</sup>NK1.1<sup>−</sup>Ly-6G<sup>+</sup> cells (chow, 3.0% ± 0.1% cells; Western diet, 3.8% ± 0.4% cells) than apoE<sup>+/+</sup> mice (chow, 1.8% ± 0.3% cells; Western diet, 1.9% ± 0.2% cells). These cells were phenotypically distinct from monocytes and were not examined further.

Although the number of circulating monocytes increased dramatically in apoE<sup>−/−</sup> mice fed Western diet, Ly-6C<sub>hi</sub> cells consistently expressed CD62 ligand (CD62L; also known as L-selectin) but not CD11c, while Ly-6C<sub>lo</sub> cells consistently expressed low levels of CD11c but not CD62L (Figure 1F), as previously reported for these monocyte subsets (21). Morphologic analysis of flow-sorted cells also showed that cells of both subsets retained their size as well as their characteristic kidney- or horse-shoe-shaped nuclei (Figure 1G).

Having determined that numbers of circulating monocytes increased in apoE<sup>−/−</sup> mice on Western diet, we assessed the spatial and temporal course of monocytosis development by quantification of monocytes and their subsets in the bone marrow, peripheral blood, and spleen over 250 days of Western diet consumption (Figure 2A). Analysis included additional compartments because the bone marrow produces monocytes and the spleen may serve as a reservoir for monocytes in the periphery. Monocytosis developed progressively in all 3 compartments, and the blood and spleen showed predominant expansion of the Ly-6C<sub>hi</sub> subset. Statistical analysis matched the data to an exponential growth curve, permitting determination of doubling time for each tissue. The Ly-6C<sub>hi</sub> subset showed the lowest doubling times (95% confidence interval, 33 to 38 days in the blood), while, as expected, the Ly-6C<sub>lo</sub> monocyte subset had the highest doubling times (95% confidence interval, 145 to 256 days in the blood).

![Figure 2](image-url)

**Figure 2**

Peripheral blood monocytosis develops over the course of 250 days on an atherogenic diet. (A) Number of total monocytes and Ly-6C<sub>hi</sub> and Ly-6C<sub>lo</sub> subtypes in bone marrow, blood, and spleens of apoE<sup>−/−</sup> mice at various days of Western diet. Statistical analysis was based on an exponential growth curve and known cell numbers on day 0. Curve fit (solid line) and 95% confidence intervals (dashed lines) are shown. Doubling time (DT) of cell number is shown. (B) The same analysis was conducted with peripheral blood from apoE<sup>+/+</sup> and apoE<sup>−/−</sup> mice that remained on chow diet. Doubling time of cell number in days is shown. (C) Splenic CD11b<sup>+</sup>CD90<sup>−/−</sup>B220<sup>−</sup>CD49b<sup>−</sup>NK1.1<sup>−</sup>Ly-6G<sup>+</sup> cells were divided into F4/80<sup>−</sup>CD11c<sup>−/−</sup>I-A<sup>−</sup>–high macrophages/dendritic cells (gate i) and F4/80<sup>−</sup>CD11c<sup>−/−</sup>I-A<sup>−</sup>–low monocytes, which were further divided into Ly-6C<sub>hi</sub> (gate ii) and Ly-6C<sub>lo</sub> (gate iii) subsets. These 3 subsets were isolated and stained with HEMA 3 for microscopic analysis. Scale bar: 10 μm. Results are pooled from 8 independent experiments.
Control experiments used blood from apoE\(^{-/-}\) and apoE\(^{+/-}\) mice during 250 days of chow consumption (Figure 2B). apoE\(^{-/-}\) mice on chow did not develop monocytosis, excluding the possibility that age drives the increase. apoE\(^{-/-}\) mice on chow showed moderately increased survival (Figure 3A) while retaining their monocytic Ly-6C\(^{hi}\) phenotype as identified in Figure 1. Statistical analysis matched the data to an exponential growth curve, though doubling times (e.g., 95% confidence interval, 63 to 86 days for Ly-6C\(^{hi}\) monocytosis in the blood) were 1.9–2.3 times longer than in apoE\(^{-/-}\) mice on Western diet. Thus, atherosclerosis and Ly-6C\(^{hi}\) monocytosis arise concomitantly in apoE\(^{-/-}\) mice and show aggravation by Western diet.

Although we used the same criteria to define monocytes in the bone marrow and spleen as in blood (e.g., the CD11b\(^{+}\)CD90\(^{+}\)B220\(^{lo}\)CD49b\(^{-}\)NK1.1\(^{hi}\)Ly-6C\(^{hi}\) phenotype), the preponderance of macrophages and dendritic cells in the spleen necessitated a secondary step to ensure that the cells were indeed monocytes. In the spleen, monocytes were further defined as F4/80\(^{+}\)CD11c\(^{-}\)I-A\(^b\)^{lo} phenotype among splenic CD11b\(^{+}\)CD90\(^{+}\)B220\(^{lo}\)CD49b\(^{-}\)NK1.1\(^{hi}\)Ly-6C\(^{hi}\) monocytosis on days 1 and 5 or in age-matched untreated mice. (C) Percent of splenic Ly-6C\(^{hi}\) monocytes recovered after clodronate liposome injection in apoE\(^{-/-}\) mice on chow and Western diet compared with absolute number of cells in age-matched untreated mice. Shown are 1 of 2–3 independent experiments.

Supplementation of medium with M-CSF (50 μg/ml) for 24 hours partially fostered Ly-6C\(^{hi}\) monocyte survival (Figure 3A). These data suggest that LDL and/or its derivatives directly and indirectly promote the survival of Ly-6C\(^{hi}\) monocytes. Nevertheless, in vitro conditions used may not reproduce the in vivo environment in apoE\(^{-/-}\) mice on Western diet, since the number of Ly-6C\(^{hi}\) monocytes did not increase in vitro. Therefore, we sought to compare directly the proliferation and survival of monocytes in vivo. apoE\(^{-/-}\) animals consuming either chow or Western diet received daily i.p. injections of BrdU for 3 days and were sacrificed 1 day later. Mononuclear cells were purified from the bone marrow, blood, and spleen, and monocytes were analyzed for apoptosis (annexin V) and proliferation (anti-BrdU mAb). The Western diet decreased the number of annexin V\(^{+}\) cells within Ly-6C\(^{hi}\) monocytes in the bone marrow and peripheral blood but not in the spleens of apoE\(^{-/-}\) mice (Figure 3, B and C). The Western diet concurrently increased the number of BrdU\(^{+}\) cells within Ly-6C\(^{hi}\) monocytes in peripheral blood and the spleen (Figure 3, B and C).
The vast majority of bone marrow monocytes incorporated BrdU regardless of diet, likely reflecting the continuous medullary production of these cells. These observations reflect the increased survival of dividing Ly-6C<sup>hi</sup> monocytes in apoE<sup>+/+</sup> mice consuming Western diet, but may result from accelerated production in the bone marrow and/or from increased extramedullary proliferation. Interestingly, the higher levels of BrdU incorporation by Ly-6C<sup>hi</sup> monocytes in the spleen (mean fluorescence intensity, 528 ± 111) compared with the bone marrow (mean fluorescence intensity, 293 ± 63) indicate either continued proliferation in the periphery or selective emigration from the bone marrow of cells with longer proliferative histories.

Analysis of apoptosis and proliferation in apoE<sup>+/+</sup> mice showed that Western diet increased survival and proliferation of monocytes in the spleen but not in the bone marrow or blood (Figure 3, B and C). Because apoE regulates apoptosis and cell cycle (28, 29), it may participate in the differences observed between apoE<sup>+/+</sup> and apoE<sup>−/−</sup> mice.

To determine whether the Western diet also affects conversion of Ly-6C<sup>hi</sup> to Ly-6C<sup>lo</sup> monocytes, apoE<sup>−/−</sup> mice on chow and Western diet received clodronate liposomes to deplete endogenous circulating monocytes (24). Mice administered clodronate liposomes had a dramatically reduced number of Ly-6C<sup>lo</sup> monocytes in blood (chow, 0.6 ± 0.2 × 10<sup>4</sup> versus 8.7 ± 4.6 × 10<sup>4</sup> cells/ml; Western diet, 1.1 ± 0.6 × 10<sup>4</sup> versus 9.6 ± 1.3 × 10<sup>4</sup> cells/ml) and spleen (chow, 2.4 ± 1.1 × 10<sup>4</sup> versus 32.5 ± 7.3 × 10<sup>4</sup> cells/ml; Western diet, 3.8 ± 0.9 × 10<sup>4</sup> versus 381.0 ± 84.2 × 10<sup>4</sup> cells/ml) 1 day after injection. Clodronate also strongly reduced the number of Ly-6C<sup>lo</sup> monocytes in athero-digest aortas and determined the in vivo relevance of Ly-6C<sup>hi</sup> monocytes to atherosclerosis. Aortas from apoE<sup>−/−</sup> mice consum-
and/or dendritic cells in apoE⁺/− mice on Western diet (gate ii; 4.9 ± 1.7 × 10⁴ cells) than on chow (1.1 ± 0.3 × 10⁴ cells).

We counted relatively low numbers of Ly-6C⁻/⁻ and Ly-6C⁺⁺ monocytes and macrophages/dendritic cells in the aortas of apoE⁻/− mice regardless of diet (Western diet, 260 ± 10 Ly-6C⁺⁺ monocytes, 310 ± 160 Ly-6C⁻/⁻ monocytes, 1.3 ± 0.5 × 10⁴ macrophages/dendritic cells; chow, 220 ± 170 Ly-6C⁻/⁻ monocytes, 230 ± 120 Ly-6C⁺⁺ monocytes, 1.2 ± 0.3 × 10⁴ macrophages/dendritic cells; Figure 4B). The above cell numbers likely underestimate actual values, since the enzymatic digestion of aortic tissue required to obtain single-cell suspensions caused the death of many cells (90% ± 1% of events and Ly-6C⁺⁺ monocyte subsets in apoE⁻/− mice on Western diet suggested that greater than 95% of cells capable of binding to activated endothelium would belong to the Ly-6C⁺⁺ subset (Figure 5C).

Further exploration used adoptive transfer of Ly-6C⁺⁺ monocytes from CD45.2 apoE⁻/− mice into congenic CD45.1 apoE⁻/− recipients (both donor and recipient mice on Western diet for 25 weeks). Because peripheral blood contains few monocytes, splenic monocytes served as surrogates for circulating monocytes. After 24 hours, aortas from recipient mice were digested enzymatically, and single-cell suspensions were analyzed by flow cytometry. We counted 135 ± 4 donor cells in recipient aortas, of which approxi-

**Figure 5**

Ly-6C⁺⁺ monocytes adhere preferentially to TNF-α–activated endothelium, accumulate in atherosclerotic plaques, and rapidly become lesional macrophages. Further examination followed the activity and fate of Ly-6C⁺⁺ and Ly-6C⁺⁺ monocytes isolated from either apoE⁺/− or apoE⁻/− mice on Western diet or chow. More than 70% of the cells remained alive in culture 24 hours after the isolation procedure (data not shown) and preserved monocyctic markers (i.e., Ly-6C⁺⁺ or Ly-6C⁺⁺, CD11b⁺CD90⁺B220⁺CD49b⁺NK1.1⁺Ly-6C⁺⁺, and F4/80⁺CD11c⁺I-A⁺–low; Figure 5A). We initially determined the capacity of monocyte subsets to adhere to TNF-α–activated murine cardiac endothelium under laminar flow conditions. Freshly isolated blood Ly-6C⁺⁺ monocytes from apoE⁺/− mice on either Western diet or chow adhered efficiently to the endothelium within minutes, while Ly-6C⁺⁺ cells adhered significantly less well (Figure 5B, black bars). As did naive lymphocytes (data not shown). Similar results were observed with monocytes isolated from apoE⁻/− mice (Figure 5B, black bars). These results indicate that Ly-6C⁺⁺ monocytes adhere preferentially to activated endothelium independent of diet or of the presence or absence of apoE. The relative proportion of circulating Ly-6C⁺⁺ monocytes contributed to debris and dead cells, as defined by low forward scatter) that were excluded from the analysis. The actual difference in Ly-6C⁺⁺ and Ly-6C⁺⁺ monocyte numbers is overestimated if Ly-6C⁺⁺ monocytes are more likely to die during isolation.

We performed immunohistochemical examination of aortic roots isolated from the apoE⁻/− mice described above to evaluate the spatial distribution of Ly-6C⁺⁺ cells in severe (fibrous plaque) and early (fatty streak) lesions (Figure 4C and data not shown). Ly-6C colocalized with mononuclear-like cells on the intimal face of CD31⁺ endothelial cells but not with the bulk of Mac-3⁺ macrophage-rich areas in both types of lesions. These observations suggest that Ly-6C⁺⁺ monocytes migrate to early and severe lesions and that differentiation into macrophages accompanies transmigration into the artery, although some Ly-6C⁺⁺ monocytes may reside in the innermost layer of the intima. The relative number of Ly-6C⁺⁺ cells compared with the number of Mac-3⁺ cells in these regions was 4.8 times higher in severe than in early lesions, suggesting that Ly-6C⁺⁺ monocytes migrate more efficiently to severe lesions.

**Ly-6C⁺⁺ monocytes adhere preferentially to activated endothelium, accumulate in atherosclerotic plaques, and rapidly become lesional macrophages.** Further examination followed the activity and fate of Ly-6C⁺⁺ and Ly-6C⁺⁺ monocytes isolated from either apoE⁺/− or apoE⁻/− mice on Western diet or chow. More than 70% of the cells remained alive in culture 24 hours after the isolation procedure (data not shown) and preserved monocyctic markers (i.e., Ly-6C⁺⁺ or Ly-6C⁺⁺, CD11b⁺CD90⁺B220⁺CD49b⁺NK1.1⁺Ly-6C⁺⁺, and F4/80⁺CD11c⁺I-A⁺–low; Figure 5A). We initially determined the capacity of monocyte subsets to adhere to TNF-α–activated murine cardiac endothelium under laminar flow conditions. Freshly isolated blood Ly-6C⁺⁺ monocytes from apoE⁺/− mice on either Western diet or chow adhered efficiently to the endothelium within minutes, while Ly-6C⁺⁺ cells adhered significantly less well (Figure 5B, black bars). As did naive lymphocytes (data not shown). Similar results were observed with monocytes isolated from apoE⁻/− mice (Figure 5B, black bars). These results indicate that Ly-6C⁺⁺ monocytes adhere preferentially to activated endothelium independent of diet or of the presence or absence of apoE. The relative proportion of circulating Ly-6C⁺⁺ monocytes contributed to debris and dead cells, as defined by low forward scatter) that were excluded from the analysis. The actual difference in Ly-6C⁺⁺ and Ly-6C⁺⁺ monocyte numbers is overestimated if Ly-6C⁺⁺ monocytes are more likely to die during isolation.
Approximately 25% were Ly-6C<sup>hi</sup> and approximately 75% were Ly-6C<sup>lo</sup> (Figure 5D). Many donor cells showed enhanced expression of F4/80 and I-A<sub>b</sub> (Figure 5D). Remarkably, the combined analysis of Ly-6C and F4/80 expression by donor cells revealed the existence of at least 3 distinct populations, Ly-6C<sup>hi</sup>F4/80<sup>-</sup>, Ly-6C<sup>hi</sup>F4/80<sup>+</sup>, and Ly-6C<sup>lo</sup>F4/80<sup>+</sup>, resembling monocytes, monocytes in the process of differentiating into macrophages, and mature macrophages, respectively (Figure 5E). These phenotypic relationships are comparable to those observed among endogenous populations (Figure 4A). In contrast, donor cells retrieved from the spleens of the same mice remained phenotypically unchanged (Figure 5D).

These results demonstrate recruitment of Ly-6C<sup>hi</sup> monocytes to atherosclerotic aortas, followed by local and rapid (<24 hours) differentiation into macrophages. Combined with the restricted localization of Ly-6C<sup>hi</sup> monocytes to the luminal face of the endothelium, these data suggest that differentiation into macrophages accompanies transmigration and also support the notion that aortic monocyte detection did not result from contamination of circulating cells.

To determine the relative capacity of Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> monocytes to migrate to atherosclerotic aortas and to determine whether accumulation of Ly-6C<sup>hi</sup> cells mapped to lesions, we labeled equal numbers of splenic Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> monocytes from apoE<sup>−/−</sup> mice on Western diet with [111<sup>In</sup>]oxine and injected them separately into apoE<sup>−/−</sup> mice on Western diet. After 24 hours, we excised the aortas and calculated the percent injected dose per gram of tissue, which revealed that Ly-6C<sup>hi</sup> cells preferentially accumulated in aortas (Figure 5F). Autoradiography showed discrete regions of activity only in recipients of Ly-6C<sup>hi</sup> cells (Figure 5G). The lack of such regions in animals receiving Ly-6C<sup>lo</sup> cells suggests that the signals detected in these mice corresponded to background activity. The radioactive signal observed for Ly-6C<sup>hi</sup> cells always mapped directly to areas containing lesions as determined microscopically, but not all lesions showed focal areas of radioactivity (data not shown). The relative proportion of circulating Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> monocyte subsets in apoE<sup>−/−</sup> mice on Western diet suggests that greater than 90% of cells accumulating in atherosclerotic lesions originate from the Ly-6C<sup>hi</sup> subset (Figure 5H). Taken together, these results suggest that circulating Ly-6C<sup>hi</sup> monocytes are direct precursors of lesional macrophages.

Expression of MCP-1 by a subset of cells in atherosclerotic lesions suggests active recruitment of monocytes to developing lesions in vivo (31, 32), and mice lacking MCP-1 or C-C motif chemokine receptor 2 (CCR2) show reduced atherosclerosis (33–35). Thus monocyte recruitment into lesions may require CCR2 expression, a feature of the Ly-6C<sup>hi</sup> subtype (19). Testing this hypothesis involved the adoptive transfer of Ly-6C<sup>hi</sup> monocytes from bone marrow of CCR2<sup>−/−</sup> mice into peripheral blood of atherosclerotic (CCR2<sup>+/+</sup>) apoE<sup>−/−</sup> mice. We counted only 14 ± 2 donor cells in recipient aortas, suggesting that Ly-6C<sup>hi</sup> monocyte accumulation in lesions does indeed depend on CCR2.

Statin administration attenuates Ly-6C<sup>hi</sup> monocytopsis. apoE<sup>−/−</sup> mice consumed Western diet supplemented or not with atorvastatin for 25 weeks. A control group of apoE<sup>−/−</sup> mice received regular chow. (A) Serum cholesterol after 25 weeks of diet. (B) Number of leukocytes, monocytes, and Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> subtypes in bone marrow, blood, and spleen. (C) Association between serum cholesterol and number of circulating Ly-6C<sup>hi</sup> or Ly-6C<sup>lo</sup> monocytes after 25 weeks of diet. Mean ± SEM are shown for apoE<sup>−/−</sup> mice on chow (filled circles), Western diet (open circles), and Western diet supplemented with atorvastatin (gray circles). Results are pooled from 9 independent experiments (n = 3–14 per group). Student’s t test was used.
reduction of Ly-6C<hi> monocytes attenuates disease. Repeated administration of anti-Ly-6C mAb could theoretically control Ly-6C<hi> monocytosis. This approach is impractical, given the chronic nature of atherogenesis, and may not keep the size of the Ly-6C<hi> monocyte population at homeostatic levels, but rather trigger transient cell depletion upon each mAb injection. In contrast, since inhibitors of hydroxymethylglutaryl coenzyme A reductase (statins) decrease cholesterol levels, exert antiinflammatory effects, and attenuate atherosclerosis (36, 37), we sought to determine whether concurrent treatment of apoE<–/>– mice on Western diet with statin also modulates the extent of monocytosis. Mice analyzed after 25 weeks of atorvastatin treatment had significantly attenuated serum cholesterol levels when compared with age-matched littersates on Western diet (Figure 6A). The statin treatment also reduced monocytosis (Figure 6B). Specifically, the numbers of Ly-6C<hi> monocytes declined significantly in the spleen and peripheral blood (although they remained higher than in apoE<–/>– mice on chow), whereas the numbers of Ly-6C<lo> monocytes in the bone marrow fell to levels found in apoE<–/>– mice on chow (Figure 6B). The statin treatment also reduced the numbers of the Ly-6C<lo> monocytes in the spleen. Antinflammatory effects of statins beyond LDL lowering could also participate in the attenuation of Ly-6C<lo> cell numbers.

Blood Ly-6C<hi> monocyte counts positively associated with serum cholesterol levels in apoE<–/>– (data not shown) and apoE<–/>– mice (Figure 6C). Furthermore, correlation between Ly-6C<hi> numbers from statin-treated apoE<–/>– mice conformed to a linear axis between chow- and Western diet–fed animals. Such correlative analysis did not apply for blood Ly-6C<lo> monocytes, since Ly-6C<lo> cell counts were similar for all serum cholesterol concentrations.

Discussion

This study shows that hypercholesterolemia in apoE<–/>– mice induced the progressive and selective expansion of Ly-6C<hi> monocytes, a phenomenon we have termed HAM. Ly-6C<hi> monocytes participate in atherosclerosis because they preferably adhered to activated endothelium, accumulated in lesions, and locally differentiated into macrophages, the predominant leukocytes during plaque inflammation and development. Although we did not describe a role for Ly-6C<lo> cells in atherosclerosis, future studies may determine whether this subset, as a possible precursor of dendritic cells (38), influences the adaptive arm of the immune system, particularly with respect to antigen presentation and lymphocyte activation, processes known to occur in atherosclerosis (1).

HAM results from continued bone marrow production of Ly-6C<hi> monocytes, increased survival of these cells in the periphery, and impaired conversion to the Ly-6C<lo> subset. Monocytosis was characterized by a doubling of the Ly-6C<hi> population every month in peripheral blood, generating up to 1.4 × 10^8 cells/mL after 250 days of diet; although the rate of Ly-6C<hi> monocytosis was slightly slower in the spleen (doubling time of around 2 months), this organ served as a much larger reservoir of Ly-6C<hi> monocytes, reaching approximately 2 × 10^7 cells after 250 days of diet. Moreover, since the number of Ly-6C<hi> monocytes continuously increased during the 250 days of the study, monocytosis expansion would likely have persisted beyond this point.

We suggest that Ly-6C<hi> monocytosis requires elevated concentrations of cholesterol or lipid derivatives, since (a) monocytosis occurred in mice fed Western diet; (b) the presence of a cholesterol-lowering drug of the statin class at least partially controlled monocytosis in vivo, and (c) the addition of LDL for 24 hours to purified Ly-6C<hi> monocytes in vitro allowed for their increased survival without promoting differentiation into macrophages. These findings demonstrate that augmented serum cholesterol levels, in addition to promoting lipid deposition in lesions (39) and inducing M-CSF production for local monocyte maturation into macrophages (40, 41), also trigger the expansion of circulating monocytes. These data suggest that a cholesterol-rich diet promotes murine atherogenesis, at least in part through the development of monocytosis. Molecules such as M-CSF, GM-CSF, PI3K, and NF-kB are induced by constituents of modified LDL and mediate the expansion and maturation of macrophages (42, 43). It will be of interest to determine whether similar pathways also elicit Ly-6C<hi> monocytosis and why circulating Ly-6C<lo> monocytes appear less sensitive to the same stimuli.

Following the fate of adoptively transferred, purified monocyte subsets revealed that lesional macrophages derived predominantly from circulating Ly-6C<hi> monocytes, pointing to an active role in this process for the membrane-bound, glyco-sphingosylphosphatidylinositol-linked protein Ly-6C (44–46). Future studies will show whether Ly-6C serves only as a marker for or also contributes functionally to inflammatory monocytes, whether Ly-6C characterizes inflammatory cells capable of tissue transmigration, and whether therapeutic targeting of Ly-6C, or a human equivalent, contains monocytosis and consequently reduces atherosclerosis.

The observation that Ly-6C<lo> cells trafficked differentially to lesions supports the concept of heterogeneity of atheromata (47, 48) and suggests that these cells migrate preferentially to lesions of high inflammatory activity. Interestingly, fatty streaks harbored few Ly-6C<lo> monocytes, indicating low monocyte accumulation at this stage. Expression of CD62L also suggests a role for Ly-6C<hi> monocytes in antigen presentation in secondary lymphoid organs and participation in adaptive immunity.

Activated macrophages, operationally categorized as M1 cells (classically activated macrophages), are induced by IFN-γ and generate inflammation, while M2 cells (alternatively activated macrophages) generated in response to IL-4 or IL-13 can exhibit immunoregulatory function (49, 50). Ly-6C<hi> monocytes selectively populate sites of experimentally induced inflammation and can locally differentiate into M1 cells. However, whether the cytokine milieu recruits and/or activates specific monocyte subsets, or indiscriminately polarizes already accumulated subsets for particular macrophage function will require further study.

Clinical data have shown a positive correlation between white blood cell count and acute myocardial infarction (5–7). More recently, a study showed that monocytosis is an independent risk factor for coronary artery disease (51). Here we propose that monocytosis is an etiologic constituent of experimental atherogenesis. Future research will determine whether humans with elevated serum cholesterol and/or C-reactive protein exhibit CD14<hi>CD16<–/>– monocytosis, and whether this variable relates to atheroma burden or complications. Overall, our present results indicate a striking and dynamic alteration of the monocyte repertoire during experimental atherogenesis, recognize circulating Ly-6C<hi> monocytes as key mediators of chronic inflammation, and suggest what we believe to be novel mechanistic hypotheses regarding the interface of innate immunity and atherosclerosis that will require future testing experimentally and in humans.
Methods

Animals. apoE<sup>+/−</sup> (C57BL/6) and apoE<sup>−/−</sup> (B6.129P2-Apoem<sup>tm1Unc</sup>) mice were purchased from The Jackson Laboratory. The apoE<sup>−/−</sup> animals had been backcrossed to the C57BL/6 background for at least 10 generations. apoE<sup>−/−</sup> CD45.1<sup>−/−</sup> mice were generated after backcrossing apoE<sup>−/−</sup> mice to C57BL/6 CD45.1<sup>−/−</sup> mice (The Jackson Laboratory). C57BL/6 CCR2<sup>−/−</sup> mice were a gift from B. Rollins (Dana-Farber Cancer Institute, Boston, Massachusetts, USA), A. Luster (Massachusetts General Hospital and Harvard Medical School), and I. Charo (UCSF, San Francisco, California, USA). At 10 weeks of age, groups of animals were placed on a Western diet (21.2% fat/weight, 0.2% cholesterol; Harlan Teklad), which was defined as day 0 in this study. The remaining animals consumed a regular chow diet. A group of animals received supplemental atorvastatin in their diet (52) (0.01% w/w) starting on day 0. All studies were conducted with age-matched animals after 20–25 weeks of diet unless otherwise stated. All protocols were approved by the Subcommittee on Animal Research Care (SARC) at Massachusetts General Hospital.

Cells. Cells were harvested from bone marrow, peripheral blood, spleens, and aortas at the time points indicated in Results. Bone marrow from both tibias was harvested, and the cells were collected by inserting a needle into the bone and washing with HBSS supplemented with 0.2% (w/v) BSA and 1% (w/v) FCS. Peripheral blood was drawn via cardiac puncture with citrate solution (100 mM Na-citrate, 130 mM glucose, pH 6.5) as anticoagulant, and mononuclear cells were purified by density centrifugation (16). Total leukocyte numbers were determined using acetic acid lysis solution (3% HEM 3 solution II, 94% dH2O, 3% glacial acetic acid). Blood smears were prepared to corroborate monocyte numbers and subtract percent granulocytes from total leukocyte counts. Spleens were removed, triturated in HBBS (cellgro; Mediatech Inc.) at 4°C with the end of a 3-ml syringe, and filtered through nylon mesh (BD Biosciences). Aortas were excised and placed into a cocktail of collagenase I, collagenase XI, DNase I, and hyaluronidase (Sigma-Aldrich) at 37°C for 1 hour, as described previously (30). Cells were then trituated through nylon mesh. The cell suspensions were centrifuged (1500 g, 4°C), red blood cells were lysed with ACK lysis buffer, and the resulting single-cell suspensions were washed with HBBS supplemented with 0.2% (w/v) BSA and 1% (w/v) FCS.

mAbs and flow cytometry. For visualization of monocytes, cells were incubated with a cocktail of mAbs against T cells (CD90-PE, 53-2-1), B cells (B220-PE, RA3-6B2), NK cells (CD49b-PE, DX5 and NK1.1-PE, PK136), granulocytes (Ly-6G–PE, 1A8), myeloid cells (CD11b-APC, M1/70) and monocyte subsets (Ly-6C–FITC, AL-21). In Figure 1F, labeling with Ly-6C–biotin–strept-PerCP was performed so as to allow combination with CD11c–FITC (HL3) and CD62L–FITC (MEL-14). CD45.2–FITC and CD45.2–APC (A20) mAbs were also used (all from BD Biosciences). Monocytes were identified as CD11b<sup>hi</sup>CD90<sup>−/−</sup>B220<sup>−/−</sup>CD49b<sup>−/−</sup>NK1.1<sup>−/−</sup>Ly-6G<sup>−/−</sup> cells. F4/80 (BM8)-biotin–strept-PerCP, I-A<sup>−/−</sup> (AF6-120.1)-biotin–strept-PerCP and CD11c (HL3)-biotin–strept-PerCP mAbs also served to determine macrophage and dendritic cell differentiation. Monocyte numbers were calculated as total leukocytes (corrected for granulocyte content, as determined on blood smears) multiplied by percent cells within the monocyte gate of the mononuclear cell fraction. Within this population, subsets were identified as either Ly-6C<sup>hi</sup> or Ly-6C<sup>lo</sup>. The number of cells in each subset was calculated by multiplying monocyte number by percent of cells in the subset. For proliferation studies, mice received 1 mg BrdU i.p. on 3 consecutive days prior to sacrifice on day 4. Cells were stained with anti-BrdU or annexin V, according to the manufacturer’s protocol (BD Biosciences), and analyzed on a FACSCalibur or flow sorted on a FACSArray (all from BD Biosciences). For depletion of monocytes (24), mice were injected i.v. with 0.2 ml of dichloromethylene-bisphosphonate (clodronate) (Sigma-Aldrich) liposomes. Clodronate was incorporated into liposomes, as previously described (53). For in vitro studies, cells were placed in complete medium (RPMI 1640 with 1 mM sodium pyruvate, 10 mM HEPES, 2 mM glutamine, 1% [w/v] penicillin/streptomycin and 10% [v/v] FCS previously heat inactivated for 1 hour at 56°C) alone or supplemented with either 100 μg/ml AcLDL or 50 μg/ml M-CSF (R&D Systems). For morphologic characterizations, sorted cells were spun, resuspended in PBS, prepared on slides by cytocentrifugation (Shandon; Thermo Electron Corp.) at 10 g for 2 minutes, and stained with HEMA-3 (Biochemical Sciences Inc.). Monocytes were identified as cells 10–30 μm in diameter with a horseshoe- or kidney-shaped nucleus, a variable nucleus-to-cytoplasm ratio, and fine granules and vacuoli.

Mouse heart EC isolation. Mouse heart ECs (MHECs) were isolated and cultured from 7-day-old mice (54). Briefly, harvested tissues were digested with collagenase (Worthington Biochemical Corp.), and ECs were isolated with 2 rounds of magnetic bead purification (Invitrogen), the first with anti-PECAM-1 and the second with anti-ICAM-2 mAbs (BD Biosciences).

Monocyte adhesion under defined laminar flow conditions. Monocytes from blood of apoE<sup>−/−</sup> and apoE<sup>−/−</sup> mice were isolated. Interactions with MHECs were examined under conditions of fluid shear stress in a parallel plate flow chamber (55). Briefly, confluent MHECs were grown on glass coverslips (25-mm diameter; Carolina Biological Supply Co.) coated with 5 μg/ml fibronectin (Sigma-Aldrich), stimulated with 120 ng/ml TNF-α for 4 hours, and inserted into the flow chamber. Monocytes (10<sup>6</sup> cells) were suspended in flow buffer (Dulbecco PBS, 0.75 mM Ca<sup>2+</sup> and Mg<sup>2+</sup>, containing 0.1% human serum albumin) and drawn through the flow chamber as a bolus at an estimated shear stress of 0.76 dynes/cm<sup>2</sup>. Monocyte accumulation was measured after 3 minutes of flow by counting stably adherent monocytes in 12–16 different fields. Monocyte-MHEC interactions were viewed using videomicroscopy (x20 phase-contrast objective) and digitally recorded to a PC using VideoLab software (version 2.2.1; Ed Marcus Laboratories).

Immunohistochemistry. Aortas were excised, frozen in O.C.T. compound (Sakura), sectioned in 5-μm slices, and stained by the avidin-biotin-peroxidase method as previously described (56). Cell types were identified with primary antibodies: monoclonal anti-CD31 (MEC 13.3) for endothelial cells, rat anti-mouse Mac-3 (M3/84) for macrophages, and anti-Ly-6C (AL-21) antibody to mouse macrophage precursor cells (BD Biosciences). Images were captured with a digital camera (Nikon DXM1200F) using imaging software ACT-1 (version 2.63; Nikon).

In vivo accumulation. Purified Ly-6C<sup>hi</sup> or Ly-6C<sup>lo</sup> monocytes from the spleen of apoE<sup>−/−</sup> or apoE<sup>−/−</sup> CCR2<sup>−/−</sup> mice were injected into apoE<sup>−/−</sup> CD45.1<sup>−/−</sup> mice or labeled with <sup>111</sup>Inoxine according to the manufacturer’s protocol (Amersham Biosciences). For imaging and autoradiography experiments, cells were washed with HBSS, spun, and resuspended in <sup>111</sup>Inoxine for 15 minutes at 37°C, pH 6.5–7.5. The cells were then washed twice with HBSS. This labeling protocol keeps the cells viable and in a resting state (16). Approximately 0.8 × 10<sup>6</sup> cells incorporating 50 μCi were injected i.v. into apoE<sup>−/−</sup> mice fed Western diet for 50 weeks. The total amount of activity that was injected into each animal was measured with a radiospectrotr camera (Capintec Inc.). For flow cytometric studies, approximately 4 × 10<sup>6</sup> Ly-6C<sup>hi</sup>CD45.2<sup>−/−</sup> monocytes were injected to CD45.1<sup>−/−</sup>apoE<sup>−/−</sup> mice fed Western diet for 50 weeks. After 24 hours, all animals were euthanized with CO<sub>2</sub>. Aortas were perfused in situ with 10 ml HBSS via the left ventricle and excised from the root to the bifurcation. All heart and fat tissue was excised to eliminate confounding signal. Aortas were then subjected to digestion (flow cytometric studies, see above). For radioactive studies, cpm for each aorta as well as blood were measured using a Wallac 1480 WIZARD gamma counter (PerkinElmer). Percent injected dose per gram tissue was calculated after correcting for...
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