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Gender-Specific Modulation of the Response to Arterial Injury by Soluble Guanylate Cyclase α1

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Abstract: Objective: Soluble guanylate cyclase (sGC), a heterodimer composed of α and β subunits, synthesizes cGMP in response to nitric oxide (NO). NO modulates vascular tone and structure but the relative contributions of cGMP-dependent versus cGMP-independent mechanisms remain uncertain. We studied the response to vascular injury in male (M) and female (F) mice with targeted deletion of exon 6 of the sGCα1 subunit (sGCα1−/−), resulting in a non-functional heterodimer.

Methods: We measured aortic cGMP levels and mRNA transcripts encoding sGC α1, α2, and β1 subunits in wild type (WT) and sGCα1−/− mice. To study the response to vascular injury, BrdU-incorporation and neointima formation (maximum intima to media (I/M) ratio) were determined 5 and 28 days after carotid artery ligation, respectively.

Results: Aortic cGMP levels were 4-fold higher in F than in M mice in both genotypes, and, within each gender, 4-fold higher in WT than in sGCα1−/−. In contrast, sGCα1, sGCα2, and sGCβ1 mRNA expression did not differ between groups. 3H-thymidine incorporation in cultured sGCα1−/− smooth muscle cells (SMC) was 27%±12% lower than in WT SMC and BrdU-incorporation in carotid arteries 5 days after ligation was significantly lower in sGCα1−/− M than in WT M. Neointima area and I/M 28 days after ligation were 65% and 62% lower in sGCα1−/− M than in WT M mice (p<0.05 for both) but were not different in F mice.

Conclusion: Functional deletion of sGCα1 resulted in reduced cGMP levels in male sGCα1−/− mice and a gender-specific effect on the adaptive response to vascular injury.

Key Words: Soluble guanylate cyclase, gender, vascular remodelling, nitric oxide.

INTRODUCTION

Nitric oxide (NO) plays an important role in maintaining normal vascular tone and modulates the function of vascular cells participating in the response to arterial injury. A better understanding of the molecular mechanisms regulating the response to vascular injury is fundamental to develop more effective therapies for atherosclerosis and restenosis following percutaneous vascular interventions. In the vessel wall, NO is produced by endothelial nitric oxide synthase (NOS3). The main intracellular receptor of NO is soluble guanylate cyclase (sGC), a heterodimer composed of α and β subunits that catalyzes the conversion of GTP to cGMP in response to NO. A rise in cGMP can activate cGMP-dependent protein kinase 1 (cGKI) and result in vascular smooth muscle cell (SMC) relaxation. Recently, Mergia et al. reported that the targeted deletion of exon 4 of the sGCα1 subunit results in a greater than 90% reduction in aortic NO-stimulated sGC activity [1].

NO/cGMP signal transduction, one of the critical pathways modulating vascular function, is reduced in atherosclerosis [2] and after balloon injury [3]. Reduced NO bioavailability in the injured vessel wall is thought to contribute to the progression of vascular lesions and has spurred interest in various interventions aimed at augmenting local NO/cGMP signal transduction. Among these, adenovirus-mediated overexpression of NOS3 [4] or both sGC subunits with administration of molsidomine, a NO-donor [5], inhibits SMC proliferation in vivo and neointima formation in vitro in rats, suggesting a vasoprotective effect of NO/cGMP signaling. In line with these observations, ApoE−/− NOS3−/− mice have a larger neointima after femoral cuff placement [6] and carotid artery ligation [7] and show increased plaque development in an ApoE−/− background [8, 9] as compared to WT mice. However, transgenic mice overexpressing NOS3 in the vascular endothelium also show accelerated atherosclerosis [10], whereas mice with a SMC-specific deletion of cGKI (SMC-cGKI−/−) develop less atherosclerosis [11]. These observations suggest that NO-mediated signal transduction can have variable effects on the response to vascular injury, which might be related to an altered balance between cGMP-dependent signaling and cGMP-independent mechanisms including S-nitrosylation of proteins and the generation of reactive oxygen species [12].
The role of cGMP in modulating the response to vascular injury has not been systematically studied. We, therefore, investigated the effect of reduced endogenous NO/cGMP signal transduction on the response to vascular injury in male (M) and female (F) mice with a targeted deletion of exon 6 of sGCα1 (sGCα1-/-). This deletion results in a non-functional α1β1 heterodimer and is associated with systemic hypertension in male but not female sGCα1-/- mice [13]. We observed gender- and genotype-specific alterations in aortic cGMP levels, corresponding to the impaired neointimal response to carotid injury in male sGCα1-/- mice. Intact sGC gene function critically determines a gender-specific adaptive response to vascular injury.

METHODS

Experimental Animals

sGCα1-/- mice with a targeted deletion of exon 6 of the α1 subunit gene were generated as described previously [13]. Experiments were performed in adult male and female sGCα1-/- (12 to 14 weeks old) and WT mice (10 to 16 weeks old) bread in a mixed genetic background (50% Swiss/50% 129Sv). The experiments conformed with the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH Publication No. 85-23, revised 1996) and were approved by the Institutional Committee on Animal Care and Welfare of the University of Leuven (Belgium). A schematic representation explaining the timeline of the experiments is shown in Fig. (1).

Vascular cGMP Concentrations

The aorta was made bloodfree and immediately frozen in liquid nitrogen and kept at -80°C until vascular cGMP concentrations were determined using a non-radioactive enzyme immunoassay according to the manufacturer's instructions (Amersham Biosciences, Little Chalfont, United Kingdom). Protein concentrations were determined with the BCA Protein assay kit (Pierce, Rockford, IL) and results expressed as pmol cGMP/mg protein.

sGC Subunit mRNA Expression

Assessment of mRNA transcript levels of murine sGCα1 (exon 5) sGCα2, sGCβ1, and hypoxanthine-guanine phosphoribosyl-transferase (HPRT) was performed by real-time fluorescence detection using specific primer sets, recognizing the respective murine DNA sequences (see reference [14] for primer and probe sequences). Samples were run in triplicate together with a dilution series of a plasmid standard for each gene to allow comparison of the copy numbers of the different subunits. Results were expressed as copy numbers normalized to mHPRT.

In Vitro SMC Proliferation and Migration

Experiments were performed in early passage aortic SMC (passage 4-6) isolated from WT and sGCα1-/- mice and cultured in DMEM + 10% fetal bovine serum. DNA synthesis and migration in response to serum stimulation were assessed using H-thymidine incorporation and Transwell migration chambers (Corning, Corning, NY), respectively. Results obtained with sGCα1-/- cells were expressed relative to WT cells and represent the mean of 6 independent experiments.

Carotid Artery Ligation

Animals were anesthetized with pentobarbital (65 mg/kg IP) and the common carotid artery was ligated close to the bifurcation as described previously [15]. Animals were euthanized 28 days after ligation to determine neointima formation. To evaluate vascular cell turnover in vivo, 5'-bromo-deoxyuridine (BrdU, 50 mg/kg IP) was injected in a separate group of animals 12 hours and 1 hour before euthanasia at 5 days when SMC turnover is maximal in the carotid artery ligation model [15]. Arteries were fixed in situ with 4% formaldehyde, embedded in paraffin, and 5-μm sections were prepared from the bifurcation towards the aortic arch. The reference point was set at the first section containing an intact internal and external elastic lamina.

Histological Analysis of Injured Carotid Arteries

To quantitate neointima formation, sections were stained at 50 μm intervals with hematoxylin and eosin and analyzed by a blinded observer. Neointima area, media area, and internal elastic lamina (IEL) length were quantified using the KS300 morphometric analysis software (Zeiss, Oberkochen, Germany). The maximum intima to media (I/M) ratio (+50 to +500 μm from the reference point in all animals) and the corresponding neointima area, media area, and IEL length were reported. BrdU-positive cells in the media were detected using a monoclonal rat anti-BrdU antibody (Oxford Biotechnology, Kidlington, UK) as described previously [14] and expressed relative to the total cell number. In addition,
cellular composition in neointimal and medial areas 28 days after ligation was investigated using antibodies directed against α-smooth muscle actin (SMC actin) (Dako, Glostrup, Denmark) and CD45 (Becton Dickinson, Franklin Lakes, NJ), a common leucocyte antigen. Additional stainings without primary antibody were included to control for non-specific staining by secondary antibodies. Finally, the media area and IEL length of the contralateral carotid artery was quantitated to compare the response to increased flow in WT and sGCα1-/- mice.

Statistics

All values are expressed as mean±SD. A Student t-test was used to compare differences between 2 groups and one-way ANOVA to compare multiple groups. Post-hoc analysis was performed using the Student-Newman-Keuls test or a Bonferroni correction (if variances were significantly different).

An expanded materials and methods section is available as online data supplement.

RESULTS

Effect of sGCα1-/- and Gender on Vascular NO/cGMP Signaling

To investigate the effect of a truncated non-functional sGCα1β1 heterodimer on sGC mRNA expression, quantitative PCR (qPCR) was performed. Relative mRNA transcript levels of the sGC subunits were comparable between the four groups (Table 1).

To investigate gender and genotype-dependent effects on sGC gene function, unstimulated aortic cGMP levels were measured. cGMP levels were 4-fold higher in females than in males in both genotypes (p<0.05 each), and, within each gender, 4-fold higher in WT than in sGCα1-/- (p<0.05 each, Fig. 2).

In Vitro SMC Proliferation and Migration

To evaluate the effect of sGC gene function on SMC proliferation, DNA turnover was quantified by measuring 3H-thymidine incorporation in early passage SMC from sGCα1-/- and WT aortic explants. Proliferation was 27%±12% lower in sGCα1-/- than in WT cells (n=6, p<0.05). In contrast, vascular cell migration was comparable in cells from both genotypes (97%±24%, n=6, p=ns). There was no difference between cells derived from male and female mice.

Table 1. Aortic mRNA Transcript Levels Normalized to HPRT

<table>
<thead>
<tr>
<th></th>
<th>sGCα1</th>
<th>sGCα2</th>
<th>sGCβ1</th>
</tr>
</thead>
<tbody>
<tr>
<td>WT M  (n=7)</td>
<td>11.8±7.0</td>
<td>1.5±0.3</td>
<td>25.4±7.3</td>
</tr>
<tr>
<td>sGCα1-/- M (n=7)</td>
<td>8.4±2.2</td>
<td>1.6±0.5</td>
<td>16.4±5.2</td>
</tr>
<tr>
<td>WT F  (n=6)</td>
<td>13.1±3.9</td>
<td>1.7±0.5</td>
<td>18.3±2.7</td>
</tr>
<tr>
<td>sGCα1-/- F (n=5)</td>
<td>11.5±9.5</td>
<td>1.8±0.9</td>
<td>19.8±10.0</td>
</tr>
</tbody>
</table>

Fig. (2). Vascular cGMP levels in WT (open bars) and sGCα1-/- (closed bars) male mice (M) and female mice (F). Aortic cGMP levels were 4-fold higher in F than in M mice in either genotype, and, within each gender, 4-fold higher in WT than in sGCα1-/- mice (*p<0.05 vs. all).

Neointimal Response to Vascular Injury in sGCα1-/- Mice

To evaluate the role of sGC gene function in the adaptive response to vascular injury, the common carotid artery was surgically ligated in WT and sGCα1-/- mice. The maximum I/M ratio in both genotypes was observed at a distance ranging from +50 to +500 μm proximal to the ligation, while almost no neointima was observed beyond +1000 μm. Maximum I/M ratio 28 days after ligation was lower in sGCα1-/- M than in WT M mice (p<0.05, Figs. 3A and 3B, respectively) but was not different in F mice (Fig. 4A). Neointima area was also lower in sGCα1-/- M compared to WT M or F mice of either genotype (p<0.05 vs. all). Media area and IEL length did not differ between groups (Table 2). The neointimal cells 4 weeks after ligation showed marked SMC actin immunoreactivity (Fig. 3C), while almost no CD45-positive inflammatory cells were observed in the neointima or the media (Fig. 3D). CD45 immunoreactivity was predominantly observed in the adventitia.
We next investigated whether carotid artery ligation would differentially alter the response to increased flow in the contralateral carotid artery in both genotypes. There was no difference in media area or IEL length of the contralateral carotid artery in M or F mice (Table 2).

**BrdU-Incorporation 5 Days After Ligation**

To investigate genotype-associated changes in SMC turnover following vascular injury, BrdU was injected 12 hours and 1 hour before sacrifice in WT and sGCα1−/− mice.

The proliferative index, expressed as % BrdU positive cells, was lower in sGCα1−/− M compared to WT M mice (*p<0.05, Fig. 4B), which contributed to a reduced I/M ratio 28 days after ligation. There was no difference between sGCα1−/− F and WT F mice.

**DISCUSSION**

NO/cGMP signal transduction plays an important role in maintaining normal vascular function and structure and modulates the response to arterial injury. In the present study, we investigated the role of sGC in vascular tone and injury.

**Fig. (3).** The average maximum I/M ratio in WT male mice (panel A) was significantly higher than in sGCα1−/− male mice (panel B). The neointima (NI) was almost exclusively composed of SMC actin positive cells (panel C), while almost no CD45-positive cells (panel D) could be detected in the neointima or the media (M) between the internal and external elastic laminae (arrowheads, panel D). CD45 immunoreactivity was predominantly observed in the adventitia (A).

**Fig. (4).** Maximum I/M ratio 28 days after ligation was reduced in sGCα1−/− M compared to WT M mice and F mice of either genotype (*p<0.05 vs. all, panel A). Proliferation 5 days after ligation, expressed as % BrdU-positive cells/total number of cells, was also significantly lower in sGCα1−/− M than in WT M mice (*p<0.05 vs. WT M mice, panel B), while there was no difference in F mice.
study, a targeted deletion in the sGCα1 subunit resulting in a non-functional α1β1 heterodimer, was associated with a reduced response to vascular injury in male but not in female sGCα1−/− mice compared to WT mice. Male sGCα1−/− mice showed a significant reduction in DNA turnover in the remodeling arterial segment 5 days after carotid artery ligation compared to male WT mice and a smaller neointimal area 28 days after ligation. No differences were observed between female sGCα1−/− and WT mice. These data suggest an important gender-specific effect of endogenous sGC gene function on the response to systemic vascular injury.

To investigate the effect of reduced sGC gene function on baseline NO/cGMP signal transduction in the vessel wall, we determined aortic cGMP concentrations in male and female WT and sGCα1−/− mice. Aortic cGMP levels were significantly lower in sGCα1−/− M than in sGCα1−/− F and WT F mice and accompanied by a reduced response to arterial injury. In contrast, higher cGMP levels in WT F mice compared to WT M mice did not translate into a difference in the response to injury, suggesting a threshold cGMP level beyond which the vascular phenotype is no longer affected. Of note, these gender-specific differences were not observed in isolated aortic rings in a recent study by Nimmgeers et al. [16]. This is most likely due to the equilibration of aortic rings in an organ bath for 30 minutes before cGMP production in female rat aortas [17]. Moreover, the relative is consistent with previous observations of increased NO production in female rat aortas [17].

The reduced NO/cGMP signal transduction in sGCα1−/− mice did not result in altered mRNA transcript levels of the sGC subunits. Similarly, increased aortic cGMP levels in transgenic mice overexpressing NOS3 in the vascular endothelium did not affect sGCβ1 protein expression [19, 20]. These observations demonstrate that the expression of sGC subunits in the aorta is not modulated via a NO or cGMP-mediated feedback mechanism. In contrast, previous in vitro studies reported reduced sGCα1 mRNA stability and protein expression in cultured SMC following the administration of an NO-donor [21] or a sGC activator [22], underlining the difficulty of extrapolating in vitro observations in cultured SMC to in vivo NO/cGMP signaling in the vessel wall.

Vascular injury results in reduced NO/cGMP signal transduction in the vessel wall [5]. Strategies aimed at increasing NOS3 expression [4] or reducing NOS uncoupling by administration of L-arginine have successfully reduced neointima formation in rats [23]. The role of endogenous NO-mediated signaling in the adaptive response to injury and the control of vascular SMC proliferation on the other hand remains unclear. The analysis of transgenic mice has shown that NO can both promote and inhibit vascular remodeling [12]. The opposing effects of NO most likely depend on the spatiotemporal profile of NO-production (including cellular source and quantity) and are probably mediated by different cGMP-dependent and cGMP-independent mechanism [24].

Wolfsgruber and colleagues showed that activation of cGKI in vitro promotes vascular SMC growth and that cGKI is critically involved in the ingrowth of SMC in vascular lesions during atherogenesis in an ApoE−/− background [11]. Doubly deficient ApoE−/− SMC-cGKI−/− had significantly smaller vascular lesions than ApoE−/− mice. These results suggest that endogenous NO/cGMP/cGKI signaling stimulates rather than inhibits vascular SMC proliferation and that the anti-proliferative effects are mediated by cGMP-independent effects or cGMP-dependent cGKI-independent effects [12]. Stimulation of vascular SMC proliferation is potentially beneficial in atherosclerosis as SMC ingrowth could increase plaque stability and reduce the risk of an atherothrombotic event following plaque rupture. In our study, we observed a reduced response to vascular injury in sGCα1−/− M mice compared to WT M mice. Male NOS3−/−
mice, in contrast, show increased neointima formation after carotid artery ligation [7]. The systemic hypertension in male sGCα1-/- M mice may have impacted on the response to vascular injury. Hypertensive Male NOS3 +/- mice do, however, show an increased response to vascular injury, while neointima formation is reduced in male sGCα1-/- mice. Taken together, these results support the view that endogenous NO/cGMP/cGKI signaling stimulates rather than inhibits vascular SMC proliferation and indicates that the antiproliferative effect of NO is most likely mediated via cGMP-independent pathways such as direct S-nitrosation of proteins or the formation of peroxynitrite [25].

Studies in transgenic mice have also demonstrated that gender and pregnancy can affect the response to mechanical vascular injury. Male gender enhances the neointima formation in NOS3 +/- mice after femoral cuff placement, while pregnancy almost completely abolished the intimal response to vascular injury [6]. After femoral cuff placement, the neointima is almost exclusively composed of SMC as is the case after carotid artery ligation (Fig. 3C) [26]. Our results indicate that sGC could play a role in mediating gender-specific effects on the response to vascular injury. Further research is, however, warranted to better characterize the underlying molecular mechanisms. It remains to be determined whether the gender-specific effect of the functional deletion of the sGCα1 subunit on the response to vascular injury is testosterone-dependent as shown for systemic blood pressure [13]. Orchidectomy and treatment with an androgen receptor antagonist prevented hypertension in male sGCα1-/- mice, while ovariectomy did not influence systemic blood pressure in female mice. Moreover, chronic testosterone treatment increased blood pressure in ovariectomized sGCα1-/- mice, confirming that the effect of sGC on systemic blood pressure was testosterone-dependent.

The carotid artery ligation model used in this study has different characteristics than traditional injury models including wire injury. A study in ApoE-/- mice after bone marrow transplant with GFP and LacZ-positive bone marrow cells showed that there was little contribution of bone-marrow derived cells in the carotid artery ligation model compared to wire injury [26]. This model is therefore not fully representative of endovascular injury in patients. Our study on the other hand allows a relatively specific assessment of the effects of reduced NO/cGMP signal transduction on vascular SMC function after vascular injury without endothelial denudation with only limited interference from other cell types invading from the circulation (Figs. 3C and 2D). Studying the effect on SMC proliferation is of particular interest for the prevention of in-stent restenosis. While stenting prevents elastic recoil and in-stent restenosis, it is associated with increased SMC proliferation and migration [27]. Further research will be needed to improve our knowledge of NO-mediated cGMP-dependent and cGMP-independent mechanisms in the response to vascular injury to determine whether pharmacologic modulation of NO/cGMP signal transduction could be used to limit restenosis following PCI and to favor optimal healing of stented segments.

In summary, functional deletion of the sGCα1 subunit gene resulted in an impaired response to carotid artery ligation in male sGCα1-/- mice but not in female sGCα1-/- mice compared to WT mice. The targeted deletion of sGCα1 elicits an important gender-specific effect on the response to vascular injury and may impact on future innovative therapies for arterial injury.

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ABBREVIATIONS

ApoE = Apolipoprotein E
BCA = Bicinchoninic acid
BrdU = 5'-bromo-deoxyuridine
cGK = cGMP-dependent protein kinase
cGMP = Cyclic guanosine monophosphate
DMEM = Dulbecco's modified eagle medium
GTP = Guanosine triphosphate
HPRT = Hypoxanthine-guanine phosphoribosyltransferase
IEL = Internal elastic lamina
I/M ratio = Intima-to-media ratio
IP = Intraperitoneal administration
mRNA = Messenger ribonucleic acid
NI = Neointima
NO = Nitric oxide
NOS = Nitric oxide synthase
qPCR = Quantitative polymerase chain reaction
sGC = Soluble guanylate cyclase
SMC = Smooth muscle cell
WT = Wild type

REFERENCES


