Demand for \(\text{Zn}^{2+}\) in Acid-Secreting Gastric Mucosa and Its Requirement for Intracellular \(\text{Ca}^{2+}\)
Demand for Zn\textsuperscript{2+} in Acid-Secreting Gastric Mucosa and Its Requirement for Intracellular Ca\textsuperscript{2+}

JingJing Liu\textsuperscript{1}, Jonathan E. Kohler\textsuperscript{1}, Amy L. Blass\textsuperscript{1}, Juliet A. Moncaster\textsuperscript{2}, Anca Mocofanescu\textsuperscript{2}, Matthew A. Marcus\textsuperscript{3,4}, Eleanor A. Blakely\textsuperscript{4}, Kathleen A. Bjornstad\textsuperscript{4}, Chitra Amarasiriwardena\textsuperscript{5}, Noel Casey\textsuperscript{2,6}, Lee E. Goldstein\textsuperscript{3,6,9}, David I. Soybel\textsuperscript{1,7,9}

1 Department of Surgery, Brigham and Women’s Hospital, Harvard Medical School, Boston, Massachusetts, United States of America, 2 Molecular Aging & Development Laboratory, Boston University School of Medicine, Boston, Massachusetts, United States of America, 3 Advanced Light Source, Lawrence Berkeley National Laboratory, Berkeley, California, United States of America, 4 Life Sciences Division, Lawrence Berkeley National Laboratory, Berkeley, California, United States of America, 5 Channing Laboratories, Brigham and Women’s Hospital, Harvard Medical School, and the Harvard School of Public Health, Boston, Massachusetts, United States of America, 6 Center for Biometals and Metallomics, Boston University, Boston, Massachusetts, United States of America

Abstract

**Background and Aims:** Recent work has suggested that Zn\textsuperscript{2+} plays a critical role in regulating acidity within the secretory compartments of isolated gastric glands. Here, we investigate the content, distribution and demand for Zn\textsuperscript{2+} in gastric mucosa under baseline conditions and its regulation during secretory stimulation.

**Methods and Findings:** Content and distribution of zinc were evaluated in sections of whole gastric mucosa using X-ray fluorescence microscopy. Significant stores of Zn\textsuperscript{2+} were identified in neural elements of the muscularis, glandular areas enriched in parietal cells, and apical regions of the surface epithelium. In *in vivo* studies, extraction of the low abundance isotope, \textsuperscript{70}Zn\textsuperscript{2+}, from the circulation was demonstrated in samples of mucosal tissue 24 hours or 72 hours after infusion (250 \textmu g/kg). In *in vitro* studies, uptake of \textsuperscript{70}Zn\textsuperscript{2+} from media was demonstrated in isolated rabbit gastric glands following exposure to concentrations as low as 10 nM. In additional studies, demand of individual gastric parietal cells for Zn\textsuperscript{2+} was monitored using the fluorescent zinc reporter, fluozin-3, by measuring increases in free intracellular concentrations of Zn\textsuperscript{2+} ([Zn\textsuperscript{2+}]\textsubscript{i}) during exposure to standard extracellular concentrations of Zn\textsuperscript{2+} (10 mM) for standard intervals of time. Under resting conditions, demand for extracellular Zn\textsuperscript{2+} increased with exposure to secretagogues (forskolin, carbachol/histamine) and under conditions associated with increased intracellular Ca\textsuperscript{2+} ([Ca\textsuperscript{2+}]\textsubscript{i}). Uptake of Zn\textsuperscript{2+} was abolished following removal of extracellular Ca\textsuperscript{2+} or depletion of intracellular Ca\textsuperscript{2+} stores, suggesting that demand for extracellular Zn\textsuperscript{2+} increases and is influenced by extracellular Ca\textsuperscript{2+}.

**Conclusions:** This study is the first to characterize the content and distribution of Zn\textsuperscript{2+} in an organ of the gastrointestinal tract. Our findings offer the novel interpretation, that Ca\textsuperscript{2+} integrates basolateral demand for Zn\textsuperscript{2+} with stimulation of secretion of HCl into the lumen of the gastric gland. Similar connections may be detectable in other secretory cells and tissues.

Introduction

For many years, investigation of Zn\textsuperscript{2+} transport in the gastrointestinal tract has focused on nutritional requirements that maintain body stores and pathologic consequences of inadequate intake [1,2,3]. An overall deficiency of Zn\textsuperscript{2+} stores within the body has been implicated in the systemic susceptibility to infection [4,5] and in the pathogenesis of some cancers [6,7,8]. Also, an important physiologic role for Zn\textsuperscript{2+} within the lumen of the alimentary canal has been postulated, based on the observations that supplementation of oral diets with Zn\textsuperscript{2+} has beneficial effects on diarrhea [9,10] and inflammatory conditions [11,12,13,14] of the gastrointestinal tract.

Recent reports have begun to explore the mechanisms that regulate cellular homeostasis of Zn\textsuperscript{2+} in mucosal cells of the gastrointestinal tract [15,16,17,18,19] and its potential influence on mucosal integrity and function [20,21]. In gastric mucosa, adequate intracellular stores and luminal content of Zn\textsuperscript{2+} may regulate integrity of [19] and acid secretion by [18,22] the gastric glands and enhance protection of the mucosa as a whole against acid-peptic injury [23,24]. Little is known, however, of the content and distribution of Zn\textsuperscript{2+} within the mucosa, or of the mechanisms that regulate the flow of Zn\textsuperscript{2+} into the parietal cell during secretory stimulation.

In this study, we utilized complimentary approaches to characterize content and distribution, acquisition and demand
for Zn$^{2+}$ in gastric mucosa of the rabbit and in its individual gastric glands, under resting conditions and during secretory stimulation. Our results indicate that there is variation in content and distribution of Zn$^{2+}$ within the gastric wall and mucosa. We find that, in vivo and in vitro, the mucosa and individual gastric glands are capable of extracting Zn$^{2+}$ from extracellular sources even when its concentration may be in the nanomolar range. In the isolated gastric gland, a multicellular model of epithelial secretion, we find that basolateral uptake of Zn$^{2+}$ is modulated by $\text{Ca}^{2+}$, during stimulation with agonists of apical secretion of acid. Our findings thus suggest a novel role for classical second messenger pathways in integrating secretory functions of the apical membrane with supply functions across the basolateral membrane, and may be applicable to a variety of epithelial systems.

**Methods**

**Animals and tissue procurement**

Anesthesia and euthanasia for New Zealand White rabbits were approved according to policies of Harvard Medical School (Harvard Medical Area (HMA) Standing Committee on Animals; Protocol 03359). As described previously [15,18], rabbits (female, ~2 kg) were anesthetized with ketamine and pentobarbital, undergoing midline laparotomy in order to harvest stomach tissues and glands. For studies of fixed tissue, full thickness squares (7 mm to 10 mm each side) of gastric wall from the acid secreting regions (body/fundus) were obtained, then cut into small strips and frozen in liquid nitrogen. For gland isolations, the aorta was perfused retrograde with warmed (37°C) phosphate-buffered saline, as described previously [15]. The gastric mucosa was separated from underlying muscularis. Isolated glands were prepared using published methods [15,25]. Collagenase Type I (Sigma Chemical, St. Louis, MO) was used for ~60 min digestion with BSA in Dulbecco’s Modified Eagle Medium (DMEM, Sigma Chemical, with 100 μg/mL cimetidine, pH 7.4). Glands were used within 8 hr of isolation.

**Micro X-Ray Fluorescence Studies.** To survey the distribution of metals in the gastric mucosa, we utilized micro X-Ray Fluorescence microscopy ($\mu$XRF) [26]. In this technique, a micron-size X-ray beam is rastered over the sample. The incident X-rays excite fluorescence from elements in the sample such as Zn provided the incident energy is above the relevant absorption edge. For Zn, the edge is at 9.66 keV and the Ka fluorescence at 8.62 keV. A solid-state energy-resolving detector picks up the fluorescence and the counts in energy regions corresponding to elements of interest are recorded for each pixel. Our samples are thin enough to be transparent to both the incident and fluorescence X-rays from the detected elements. In this regime, the signals are proportional to the column densities ($\mu$g/cm$^2$) of each element, with a different sensitivity for each element.

For full thickness samples, a novel fast-freeze method was developed for near instantaneous tissue vitrification. Fixation and mounting of oriented specimens was performed on silicon wafers (Platypus Technologies, Madison, WI). Silicon substrates were used because they exhibit low background signals in $\mu$XRF and have high thermal conductivity. Sequential cryosections (50 μm) were stained with hematoxylin/eosin and then analyzed by X-ray fluorescence ($\mu$XRF) [26]. In this technique, a micron-size X-ray beam is rastered over the sample. The incident energy 11.5 keV, dwell time 100 msec/pixel, pixel size 20×20 μm$^2$, and incident beam size 16×7 μm$^2$ FWHM. For the finer map in Figure 2, the parameters were: incident energy 12.5 keV, dwell time 400 msec/pixel, pixel size 3×5 μm$^2$, and incident beam size 7×7 μm$^2$ FWHM. Fluorescence counts were recorded in energy bands corresponding to K, Ca, Fe, Cu, Ni and Zn, though no significant Ni was found. Since the signals from some elements appear in more than one band, e.g. the Kβ fluorescence of Cu in the Zn Kα band, certain fractions of the signal from such interfering channels were subtracted from the channels interfered with. The coefficients for this subtraction were derived by mapping standards containing some but not all relevant elements, e.g. Cu with no Zn.

**High Resolution Magnetic Sector Inductively Coupled Plasma Mass Spectrometry (HR-ICP-MS).** Among the five stable zinc isotopes, $^{64}\text{Zn}$, $^{66}\text{Zn}$, $^{68}\text{Zn}$, $^{69}\text{Zn}$ and $^{70}\text{Zn}$, $^{70}\text{Zn}$ is the most commonly isotopic tracer due to its low natural abundance and was used for all described experiments. The natural relative abundance for $^{70}\text{Zn}$ and $^{69}\text{Zn}$ are 0.6% and 18.8% respectively with an isotopic ratio of 0.0319 and any increase in this isotopic ratio is indicative of $^{70}\text{Zn}$ uptake in biospecimens. Unless otherwise noted, all reagent and stock solutions were prepared in ultra-trace metal grade nitric acid (Fisher Brand, Fisher Scientific, USA) made from de-ionized water (Millipore, Billerica, MA, USA) in order to match the acid matrix resulting from the sample preparation procedure. Zinc enriched in $^{70}\text{Zn}$ (99%) was obtained (Cambridge Isotope, MA, USA) and was dissolved in HCl and diluted with de-ionized water to prepare an $^{70}\text{Zn}$-enriched spiking solution of 1000 mg/L in 1% HCl/citrate. This stock solution was used to prepare standard and sample solutions for all experiments. Standard-sized tissues samples (~25 mm$^3$) or volumes of isolated glands (0.3 ml or ~2,000 glands, following settling and aspiration of supernatant media) were acid digested in aliquots of concentrated nitric acid and made up to a total volume 2 ml using de-ionized water. Analysis for isotopes $^{68}\text{Zn}$ and $^{70}\text{Zn}$ were performed concurrently on 1 ml samples, at the ICP-MS facility of the Institute of Marine and Coastal Sciences, Rutgers the State University, New Brunswick, NJ. An analytical methodology was developed to scan zinc isotopes using a double-focusing, single-collector Thermo Element 2 ICP-MS (Thermo, Waltham, MA, USA). Settings are shown in Table 1. Sample solutions were introduced into a spray chamber by a low-flow nebulizer using argon as a carrier gas. The signal intensity was obtained by integration of the counting signal of the scanning mass over a 2–4 min acquisition period. Sample blanks were taken into account and a natural abundance zinc standard solution was analyzed in between sample runs for quality control purposes [28]. Thus, in each sample the ratio for $^{70}\text{Zn}/^{69}\text{Zn}$ was calculated as an index of enrichment [28].

**Fluorescence Microscopy or Plate-Reader Fluorometry for intracellular Zn$^{2+}$ using fluozin-3.** Concentrated preparations of freshly isolated gastric glands were diluted in DMEM to a final concentration 1.875% and loaded for 30 min to 40 min with fluozin-3AM (8 μM) [18,19]. Following transfer to glass coverslips and equilibration in standard Ringer’s solutions, fluorescence imaging of individual glands was performed as described previously [18]. Glands loaded with fluozin-3AM were excited at 488 nm with emission measurement at 520 nm. Fluorescence was monitored concurrently in 4 to 8 individual parietal cells in each isolated gland. Digital images of glands were captured using a CCD camera (Hamamatsu ORCA-ER). In order to take into account variations in starting levels between individual cells, responses were reported as a normalization to starting values ($F/F_0$) [18,19]. For high-throughput fluorometry, aliquots of glands (200 μl, ~2000 glands/well) were transferred to 96 well plates (total
Figure 1. Mapping metal divalent cations in rabbit gastric mucosa. Figure 1A visualizes the section of interest with hematoxylin/eosin staining. Figure 2B, 2C and 2D provide maps at low resolution ($20 \times 20 \ \mu m^2$ pixel) of Fe, Cu and Zn. In these images, white and black indicate high and low levels of the metal, respectively.

doi:10.1371/journal.pone.0019638.g001

Figure 2. Comparisons of the distributions of Zn$^{2+}$ and other abundant cation species in a smaller region of gastric mucosa. Region of interest is shown in the inset (Figure 2A). The pixel size is $5 \times 5 \ \mu m^2$. See text for interpretation of the color images.

doi:10.1371/journal.pone.0019638.g002

Table 1. Typical Element 2 ICP-MS Instrumental parameters for the determination of zinc isotope ratios.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Forward rf power</td>
<td>1050 W</td>
</tr>
<tr>
<td>Reflected rf power</td>
<td>$&lt;2 \ \text{W}$</td>
</tr>
<tr>
<td>Coolant gas flow rate</td>
<td>16 L/min</td>
</tr>
<tr>
<td>Auxiliary gas flow rate</td>
<td>1 L/min</td>
</tr>
<tr>
<td>Nebulizer gas flow rate</td>
<td>0.8–1.2 L/min</td>
</tr>
<tr>
<td>Focus lens</td>
<td>$-700$–$-900 \ \text{V}$</td>
</tr>
<tr>
<td>Y-Deflection lens</td>
<td>$-1$–$-10 \ \text{V}$</td>
</tr>
<tr>
<td>Mass range</td>
<td>$^{66}\text{Zn}$, $^{70}\text{Zn}$</td>
</tr>
<tr>
<td>Dwell time/point</td>
<td>0.005 s</td>
</tr>
<tr>
<td>Points/peak</td>
<td>10</td>
</tr>
<tr>
<td>Scans/measurement</td>
<td>1500</td>
</tr>
<tr>
<td>Solution uptake rate</td>
<td>20–100 $\mu$L/min</td>
</tr>
<tr>
<td>Scan mode</td>
<td>E-scanning, peak-hopping</td>
</tr>
</tbody>
</table>

doi:10.1371/journal.pone.0019638.t001
volume 0.50 ml) following loading of dye in media and experimental manipulations in standard (in mM: 145 NaCl, 2.5 KH$_2$PO$_4$, 1.0 MgSO$_4$ or 1.0 MgCl$_2$, 1.0 CaCl$_2$, 10 HEPES, and 10 glucose, pH 7.4; concentration of Zn$^{2+}$ measured by atomic absorption is 1.2 μM, kindly determined by Dr. Shannon Kelleher, Pennsylvania State University, University Park, PA) or modified Ringer’s solutions. Readout of fluozin-3 fluorescence was performed (Excitation 485 nm/Emission 520 nm) after correction for background fluorescence of solutions and unlabelled glands [29,30]. Fluorometry was performed using a Synergy$^{\text{TM}}$ 2 Multi-Mode Microplate Reader (BioTek Instruments, Inc.). When comparisons to the starting fluorescence levels were required, responses were reported as a normalization to starting values (F/F$_0$) [18,19]. When comparing responses between groups at a single point time, the responses in total fluorescence are reported. Special Reagents

Thapsigargin and TPEN (tetraakis-[2-pyridylmethyl]ethylenediamine) were obtained from Molecular Probes (Eugene, OR) and were dissolved in Ringer’s solutions directly. Nigericin, DTPA (diethylenetriaminepentaacetic acid) and pyritrione were obtained from Sigma-Aldrich and was dissolved under alkaline conditions, then added to Ringer’s solutions for final dilutions of 1:1000. All solutions were checked for changes in pH and adjusted if necessary to baseline pH. For Ringer’s, calculations of free and bound concentrations of Ca$^{2+}$, Zn$^{2+}$, Fe and K are presented in various combinations as bicolor or tricolor maps. In the bottom-most map (Figure 1d) the red intensity of each pixel represents the relative amount of Fe, the green the amount of Zn and the blue the amount of K. Thus, blue-green colors represent areas containing relatively large amounts of K and Zn together. The surface epithelium is notable for a relatively high content of Zn, compared to the glandular regions deep in the mucosa. Content and distribution of Zn$^{2+}$ is homogeneous throughout the glandular region.

Measurements of $^{70}$Zn$^{2+}$ uptake in gastric mucosa and isolated gastric glands

To demonstrate that gastric mucosa acquires Zn$^{2+}$ from the circulation, in vivo, three pairs of rabbits were injected intravenously with $^{70}$Zn$^{2+}$ in sterile saline (ZnSO$_4$, 250 μg/kg) to one animal and saline alone to the other. Gastric mucosa was harvested 24 hr later, some preserved for analysis and the rest processed for isolation of individual gastric glands. The relative abundance in nature for $^{65}$Zn and $^{69}$Zn are 0.6% and 18.8%; and the ratio of $^{70}$Zn/$^{65}$Zn is 0.0319. Any enrichment of $^{70}$Zn reflected in a higher ratio would reflect uptake in tissues and individual glands. [33]. As shown in Figure 3, ICP-MS analysis of whole mucosa and isolated glands revealed appropriate ratios of $^{70}$Zn/$^{65}$Zn in saline injected animals, along with significant uptake of $^{70}$Zn$^{2+}$ in whole gastric mucosa and isolated gastric glands. Avidity of gastric mucosa for $^{70}$Zn$^{2+}$ is higher than that of peripheral tissues such as skin, retina, cornea and much higher than the avascular lens.

In in vitro studies, we evaluated potential threshold levels of extracellular [Zn$^{2+}$] which would permit detection of $^{70}$Zn$^{2+}$ uptake by gastric mucosa. Isolated gastric glands were exposed to increasing concentrations of $^{70}$Zn$^{2+}$ in Ringer’s solutions (containing EGTA 0.3 mM, with Ca$^{2+}$ adjusted to keep its free concentration 1 mM), with total content of added Zn$^{2+}$ calculated to provide a free [Zn$^{2+}$] of 1 nM, 10 nM or 100 nM. After 1 hour, glands were processed for ICP-MS. In unstimulated glands, evidence of accumulation started to become evident during exposure to extracellular [Zn$^{2+}$] at 10 nM (Figure 3), although accelerated and significant uptake was more consistently observed when [Zn$^{2+}$] reached 100 nM. These data indicate that basolateral uptake processes operate at low threshold concentrations for free extracellular Zn$^{2+}$, consistent with those that have been predicted [34] and measured [35,36] in the circulating plasma.

Fluorescence microscopy in individual parietal cells of isolated gastric glands: dependence of Zn$^{2+}$ uptake on [Ca$^{2+}$]

Flux measurements of $^{70}$Zn$^{2+}$ reliably measure uptake in relation to small extracellular concentrations; however, they do not allow continuous monitoring of intracellular Zn$^{2+}$ accumulation. To explore the conditions regulating acquisition of Zn$^{2+}$ in real time, glands were loaded with fluozen-3 (Ex 485 nm/Em 520 nm) in order to monitor changes in intracellular concentration of Zn$^{2+}$ in individual parietal cells [18]. Following transfer to coverslips on a microscope stage, fluorescence was interrogated for Ca, a metal component of the microtome knife. No Ni was detected (data not shown) indicating that contamination of the specimen with extraneous metals was unlikely.

To further define distribution of Zn within the glandular region of the mucosa (Figure 2), a region 1.8 mm in longest dimension was scanned in more detail, comparing the distribution of Zn with other physiologically relevant elements (K, Ca, Fe). Data for K and Ca are presented in grayscale as in Figure 1, and data for Zn, Fe and K are presented in various combinations as bicolor or tricolor maps. In the bottom-most map (Figure 2d) the red intensity of each pixel represents the relative amount of Fe, the green the amount of Zn and the blue the amount of K. Thus, blue-green colors represent areas containing relatively large amounts of K and Zn together. The surface epithelium is notable for a relatively high content of Zn, compared to the glandular regions deep in the mucosa. Content and distribution of Zn$^{2+}$ is homogeneous throughout the glandular region.

Data Summary and Statistical Analysis

In the microscopy imaging system, fluorescence intensities were monitored continuously throughout each experiment (SimplePCI software). At discrete time intervals, measurements were summarized as means ± SE. For comparison between treatments, unless stated otherwise, measurements in individual parietal cells (4 to 8 cells identified in each isolated gland) were combined to provide a single integrated value at each time point for each gland. Unless stated otherwise, comparisons were performed using analysis of variance for sequential or multiple measurements (Kruskal-Wallis test). Measurements of $^{70}$Zn$^{2+}$ in saline injected animals, along with significant uptake of $^{70}$Zn$^{2+}$ in whole gastric mucosa and isolated gastric glands. Avidity of gastric mucosa for $^{70}$Zn$^{2+}$ is higher than that of peripheral tissues such as skin, retina, cornea and much higher than the avascular lens.

In in vitro studies, we evaluated potential threshold levels of extracellular [Zn$^{2+}$] which would permit detection of $^{70}$Zn$^{2+}$ uptake by gastric mucosa. Isolated gastric glands were exposed to increasing concentrations of $^{70}$Zn$^{2+}$ in Ringer’s solutions (containing EGTA 0.3 mM, with Ca$^{2+}$ adjusted to keep its free concentration 1 mM), with total content of added Zn$^{2+}$ calculated to provide a free [Zn$^{2+}$] of 1 nM, 10 nM or 100 nM. After 1 hour, glands were processed for ICP-MS. In unstimulated glands, evidence of accumulation started to become evident during exposure to extracellular [Zn$^{2+}$] at 10 nM (Figure 3), although accelerated and significant uptake was more consistently observed when [Zn$^{2+}$] reached 100 nM. These data indicate that basolateral uptake processes operate at low threshold concentrations for free extracellular Zn$^{2+}$, consistent with those that have been predicted [34] and measured [35,36] in the circulating plasma.

Results

Metal Maps of Gastric Mucosa using μXRF

Shown in Figure 1 are metal maps for individual divalent metals (Zn, Cu, Fe), demonstrating relative differences in content and distribution in the mucosa. Panel 1A provides a standard tissue frozen section, stained with hematoxylin and eosin, corresponding to a neighboring section that was used to generate metal maps [Panel 1B: Fe; Panel 1C: Cu; Panel 1D: Zn]. In these maps, white represents the maximum intensity while black areas show little or none of the relevant element. Of note, consistently strong signals for the Fe and Zn are detected in region of the surface epithelium, near the luminal interface; signals for Cu are very minimal throughout the section although perhaps more concentrated in the surface epithelium. Relatively homogeneous signals for Fe are observed throughout the glandular mucosa, deep to the surface epithelium, in areas known to be populated by mitochondria-rich parietal cells [31,32]. Signal for Zn appears to be distributed within the mucosa much as that for Fe. In contrast, dense content of Zn was detected in the region of the myenteric nerve plexus, lying between the layers of muscle, whereas there was relatively little signal for Cu or Fe. As a control, samples were
monitored during exposure to standard Ringer’s solutions containing free [Zn\(^{2+}\)] of 10 \(\mu\)M to 50 \(\mu\)M. As shown in 

**Figure 4A**, exposure to standard Ringer’s containing an additional 10 \(\mu\)M Zn\(^{2+}\) leads immediately to increases in [Zn\(^{2+}\)]\(_i\), reaching plateaus within 10 minutes. These responses were abolished during exposure to low concentrations of a membrane-permeable chelator, TPEN (10 \(\mu\)M), indicating that Zn\(^{2+}\) is the source of the fluorescence signal within the cell [37].

Utilizing fluozin-3 measurements, previous studies in our laboratory [18] have consistently detected **intracellular accumulation** of labile Zn\(^{2+}\) in isolated gastric glands when extracellular Zn\(^{2+}\) was at least 50 \(\mu\)M, and this was observed in the presence or absence of extracellular Ca\(^{2+}\). We also observed an increase in the rate of Zn\(^{2+}\) uptake when glands were exposed to the secretagogue, forskolin (10 \(\mu\)M), a finding consistent with the idea that stimulation of acid secretion across the apical membrane leads to increased demand for Zn\(^{2+}\) across the basolateral membrane. These observations were corroborated by preliminary studies indicating that exposure to forskolin (10 \(\mu\)M) enhances influx of isotopic \(^{70}\)Zn\(^{2+}\), more than three-fold over baseline influx (extracellular concentration 50 \(\mu\)M, data not shown). In these *in vitro* studies, when extracellular Ca\(^{2+}\) was removed, responses became less consistent, particularly if extracellular Zn\(^{2+}\) concentrations were 10 \(\mu\)M or lower. These observations led us to hypothesize that uptake of Zn\(^{2+}\) into the parietal cell might be connected to intracellular Ca\(^{2+}\) homeostasis.

To assess whether intracellular uptake of Zn\(^{2+}\) requires the presence of Ca\(^{2+}\), a protocol was devised to totally deplete Ca\(^{2+}\) from extracellular solutions and intracellular stores. Glands preloaded with fluozin-3 were perfused with Ca\(^{2+}\)-depleted Ringer’s (0.3 mM EGTA) containing 2 \(\mu\)M thapsigargin to deplete the intracellular Ca\(^{2+}\) store, followed by exposure to 10 \(\mu\)M Zn in Ca\(^{2+}\)-depleted Ringer’s. As shown in a representative recording from 5 parietal cells in a single gland in **Figure 4B**, Zn\(^{2+}\) accumulation was abrogated under these conditions, indicating a requirement for intracellular Ca\(^{2+}\). Exposure to thapsigargin depletes intracellular, membrane-bound stores of Ca\(^{2+}\), but activates store-operated channels at the plasma membrane [38,39]; and exposure to both thapsigargin and physiologic concentrations of extracellular Ca\(^{2+}\) lead to increases in [Ca\(^{2+}\)]\(_i\), without any re-filling of stores [40]. We therefore asked whether uptake of extracellular Zn\(^{2+}\) can occur when intracellular, membrane-bound stores are depleted but store-operated entry of Ca\(^{2+}\) is present (**Figure 4C**). Gastric glands loaded with fluozin-3 were exposed first to Ringer’s containing thapsigargin (2 \(\mu\)M) and no added Ca\(^{2+}\) to deplete intracellular and extracellular sources of Ca\(^{2+}\). Under these conditions no accumulation was observed in the presence of extracellular Zn\(^{2+}\) (10 \(\mu\)M). When Ca\(^{2+}\) was subsequently restored to the perfusate (1 \(\mu\)M) in the presence of thapsigargin, increases in [Zn\(^{2+}\)]\(_i\) were observed (**Figure 4C**). Thus, the cell’s ability to take up extracellular Zn\(^{2+}\) requires some level of intracellular Ca\(^{2+}\) content, but not necessarily the presence of functionally intact intracellular stores of Ca\(^{2+}\).

**Ca\(^{2+}\)-dependent accumulation of intracellular [Zn\(^{2+}\)] is due to influx of Zn\(^{2+}\) from extracellular sources**

We next performed studies to confirm that Ca\(^{2+}\)-dependent increases in intracellular Zn\(^{2+}\) is due to influx from extracellular sources rather than release from intracellular pools. Glands loaded with fluozin-3 were exposed initially to Ringer’s solutions containing and thapsigargin (2 \(\mu\)M) and no added Ca\(^{2+}\), in order to deplete intracellular stores. These solutions also contained DTTPA (10 \(\mu\)M), a chelator of Zn\(^{2+}\) that is membrane-impermeable and therefore depletes free Zn\(^{2+}\) in the extracellular perfusate but not within the cell. Under these conditions, no increase in [Zn\(^{2+}\)] was observed. Glands were then exposed to solutions containing Zn\(^{2+}\) (10 \(\mu\)M) and Ca\(^{2+}\) (1 mM) and, as expected, increases in intracellular Zn\(^{2+}\) were observed. These increases were, however, arrested when the chelator DTTPA (10 \(\mu\)M) was added (**Figure 5**). These findings indicate that the increases in [Zn\(^{2+}\)] are due to influx from extracellular sources and not release from intracellular pools.

---

**Figure 3. Acquisition of \(^{70}\)Zn in tissues and isolated gastric glands of the rabbit.** Measurements of isotopic Zn species by ICP-MS are reported as the ratio of \(^{70}\)Zn/\(^{68}\)Zn (no dimensions) means ± SD. *p<0.05 compared to control measurements. **Protocol A.** Measurements in tissues harvested from rabbits: three control (black columns) and in three rabbits 18 hrs after intravenous infusion of \(^{70}\)ZnCl\(_2\) (250 \(\mu\)g/kg). **Protocol B.** Measurements performed in aliquots of gastric glands exposed to standard Ringer’s containing EGTA, Ca\(^{2+}\) (calculated free concentration 1 mM) and reagent ZnCl\(_2\) (control) or \(^{70}\)ZnCl\(_2\) (calculated free concentrations 1 nM, 10 nM or 100 nM).

<table>
<thead>
<tr>
<th>Zn(^{2+}) Concentration</th>
<th>i.v. (^{70})Zn(^{2+}) injected rabbits (250 (\mu)g/kg)</th>
<th>isolated rabbit glands in vitro exposure</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 nM</td>
<td>non-enriched</td>
<td>non-enriched</td>
</tr>
<tr>
<td>10 nM</td>
<td>(^{70})Zn(^{2+})-enriched</td>
<td>(^{70})Zn(^{2+})-enriched</td>
</tr>
<tr>
<td>100 nM</td>
<td>(^{70})Zn(^{2+})-enriched</td>
<td>(^{70})Zn(^{2+})-enriched</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Tissue/Sample Type</th>
<th>[^{70}]Zn(^{2+}) Accumulation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cornne</td>
<td></td>
</tr>
<tr>
<td>Retina</td>
<td></td>
</tr>
<tr>
<td>Lens</td>
<td></td>
</tr>
<tr>
<td>Skin</td>
<td></td>
</tr>
<tr>
<td>Gastric mucosa</td>
<td></td>
</tr>
<tr>
<td>Gastric glands</td>
<td></td>
</tr>
</tbody>
</table>


DOI: 10.1371/journal.pone.0019638.g003

---

Figure 3. Acquisition of \(^{70}\)Zn in tissues and isolated gastric glands of the rabbit. Measurements of isotopic Zn species by ICP-MS are reported as the ratio of \(^{70}\)Zn/\(^{68}\)Zn (no dimensions) means ± SD. *p<0.05 compared to control measurements. **Protocol A.** Measurements in tissues harvested from rabbits: three control (black columns) and in three rabbits 18 hrs after intravenous infusion of \(^{70}\)ZnCl\(_2\) (250 \(\mu\)g/kg). **Protocol B.** Measurements performed in aliquots of gastric glands exposed to standard Ringer’s containing EGTA, Ca\(^{2+}\) (calculated free concentration 1 mM) and reagent ZnCl\(_2\) (control) or \(^{70}\)ZnCl\(_2\) (calculated free concentrations 1 nM, 10 nM or 100 nM).

doi:10.1371/journal.pone.0019638.g003
Zn$^{2+}$ Demand and Ca$^{2+}$ in Gastric Mucosa

Figure 5. Ca$^{2+}$-dependent increases in [Zn$^{2+}$], are attributable to influx from extracellular sources. Recording from imaging of a single gland (4 individual parietal cells) loaded with fluo3-3. The recording begins with perfusion in standard Ca-Ringers solution, followed by Ca Ringer’s containing 2 mM thapsigargin, which depletes intracellular calcium stores. During exposure to 10 mM Zn$^{2+}$, in the presence of Ca$^{2+}$, a monotonic increase in [Zn$^{2+}$], follows, similar to that observed in Figure 4C. With addition of an excess of DTPA (100 μM), a high affinity and membrane impermeable chelator of heavy metals (but not Ca$^{2+}$ or Mg$^{2+}$), the accumulation of Zn$^{2+}$ is arrested. In 18 cells monitored in 5 glands, this arrest was always observed, demonstrating that Ca$^{2+}$ dependent loading of Zn$^{2+}$ was due solely to influx from extracellular sources.

doi:10.1371/journal.pone.0019638.g005

Fluorometric measurements of [Zn$^{2+}$]: evidence for Ca$^{2+}$/Zn$^{2+}$ exchange

In fluorescence microscopy, there is a possibility of inadvertent bias in selection of individual glands for study. To be confident of the overall characteristics of the response to depletion and restoration of [Ca$^{2+}$]$_i$, we adapted our fluorescence-based protocol of monitoring demand for Zn$^{2+}$, using a 96-well platform, which permits assessment of responses in populations of glands (~2,000 glands/well).

A first set of studies (Figure 6A) was performed to confirm that accumulation of Zn$^{2+}$ in a population of glands was impaired in the absence of Ca$^{2+}$. Glands were exposed, in sequence to: standard Ringer’s solutions containing 1 mM Ca$^{2+}$ (CaR, 30 min), then Ringer’s depleted of Ca$^{2+}$ and containing 300 μM EGTA (0CaR TG, 30 min), then 0CaR TG solutions containing 10 μM Zn$^{2+}$ (0CaR TG 10 μM Zn) for 30 min with replacement by fresh solution at 30 min. As shown, resting levels of [Zn$^{2+}$], were decreased and very little uptake was detected when extracellular and intracellular sources of Ca$^{2+}$ had been depleted.

A second set of studies (Figure 6B) was then performed to confirm that uptake of Zn$^{2+}$ could be observed despite functional impairment of intracellular Ca$^{2+}$ stores. To establish baseline conditions with depletion of intracellular stores, glands were exposed to CaR (30 min) and then 0CaR with TG (30 min). Glands were then exposed to solutions in which Ca$^{2+}$ was restored (CaR, TG and Zn$^{2+}$ 10 μM) for 30 min, with replacement by fresh solution at 30 min. As in studies of individual glands (Figures 4 and 5), when Ca$^{2+}$ was restored to the system, uptake of Zn$^{2+}$ was significantly enhanced and continued to the last period of observation (Figure 6B).

A third set of studies (Figure 6C) was then performed to confirm reversibility of Ca$^{2+}$-dependent uptake of Zn$^{2+}$. Baseline conditions were established with depletion of intracellular stores (CaR for 30 min followed by 0CaR-TG for 30 min). Glands were then exposed to solutions in which Ca$^{2+}$ had been restored (CaR, TG and Zn$^{2+}$ 10 μM) for 30 min, followed by exposure to solutions in which Ca$^{2+}$ was again depleted (0CaR, TG and Zn$^{2+}$ 10 μM) for 30 min. As previously observed, when Ca$^{2+}$ was restored to the system, uptake of Zn$^{2+}$ was significantly enhanced; however, when Ca$^{2+}$ was again removed accumulation of Zn$^{2+}$ was impaired and actually decreased. These findings suggest that Zn$^{2+}$ accumulation is responsive to Ca$^{2+}$ availability in the cytoplasm, by a mechanism that is consistent with exchange of intracellular Ca$^{2+}$ for extracellular Zn$^{2+}$.

Demand for extracellular Zn$^{2+}$ during secretory stimulation

In a final series of studies, we evaluated demand for extracellular Zn$^{2+}$ during maximal secretory stimulation, exposing glands to the
muscarinic agonist carbachol (CCh, 1 mM) and the cAMP/PKA agonist forskolin (FSK, 10 μM) [25,41,42]. Glands were loaded with fluozin-3, then pre-stimulated with CCh/FSK or vehicle (1:1000 DMSO) in media for 30 min before being loaded into wells containing Ringer’s solutions with 10 μM Zn. Responses were read 10 minutes after placement in these solutions, in order to obtain immediate estimates of labile Zn accumulation within the cell.

As shown in Figure 7, under un-stimulated conditions, intracellular content of Zn was not influenced by the absence of extracellular Ca++. During exposure to secretory agonists (CCh/FSK), accumulation of Zn was significantly enhanced if Ca++ was present in the Ringer’s solution, but significantly impaired if Ca++ was absent. These findings confirm that, during secretory stimulation, intracellular content of Zn depends on influx of extracellular Ca++. In an additional experiment, also shown in Figure 7, we found that pretreatment with thapsigargin (TG) did not impair accumulation of Zn during exposure to CCh/FSK, if Ca++ was present in the Ringer’s solution, indicating that functional intracellular stores of Ca++ are not, per se, required for the ability of the gland to take up Zn during secretory stimulation. In contrast, pretreatment with TG in Ca++-free Ringer’s enhanced the failure of the glands to preserve content of Zn++, a finding consistent with the hypothesis that accumulation of Zn++ is even more completely impaired by profound depletion of Ca++ from the extracellular compartment, intracellular stores and the cytoplasm.

Discussion

Work reported previously from our laboratory indicates that the transport of Zn by the parietal cell is a critical feature of its ability to regulate acidity within its secretory compartment, which includes the tubulovesicles of the parietal cell and the lumen of the gland. Chelation of Zn within these compartments led to marked alkalization, suggesting that the presence of Zn insulates these compartments against leakage of H+ ions [15]. Recently we have reported evidence that the secretory compartments serve as a reservoir for Zn++, accumulating it in response to proton gradients and failing to accumulate it when secretion is blocked by proton pump inhibition [18].

The studies reported here provide three sets of observations with respect to utilization of zinc within the gastric mucosa: first, as demonstrated by μXRF, that zinc accumulates differently in distinct micro-anatomic regions of the mucosa; second, as observed following in vivo intravenous injection, the non-radioactive isotope 70Zn accumulates readily within both gastric mucosa and in the acid-secreting gastric glands; and third, as suggested by in vitro studies of isolated gastric glands, that exposure to well-recognized secretory agonists accelerates the uptake of Zn from the nutrient side of the epithelium and requires intact intracellular stores of Ca++. The studies reported here refine and extend the concept that demand for Zn from the circulation and nutrient compartment may be regulated acutely by alterations in H+ secretion to the lumen.
Heterogeneous distribution of Zn\(^{2+}\) within the gastric mucosa

To our knowledge, this is the first report of the use of \(\mu\)XRF to evaluate the distribution of Zn\(^{2+}\) and other cations in the mucosa of a specific region of the gastrointestinal tract. The basis of the method is fluorescence produced by synchrotron-derived x-rays, which can be focused to localize elemental content within isolated cells [43,44] or tissues [45,46]. \textit{In vitro}, quantification within individual cells and resolution of individual isotopes of different metals is feasible [26,43,44]. However, interpretation of differences in distribution within tissues depends on unique structural features [45] or co-localizing markers [46] for cellular or sub-cellular areas of interest.

The gastric mucosa is divided into two spatially and functionally distinct regions [31,47,48]: the gastric glands, which secrete acid and pepsinogen; and the surface epithelium, which secretes mucus and bicarbonate and is thought to provide the protective barrier. The gastric glands, during exposure to secretagogues, 

Heterogeneous distribution of Zn\(^{2+}\) within the gastric mucosa

To our knowledge, this is the first report of the use of \(\mu\)XRF to evaluate the distribution of Zn\(^{2+}\) and other cations in the mucosa of a specific region of the gastrointestinal tract. The basis of the method is fluorescence produced by synchrotron-derived x-rays, which can be focused to localize elemental content within isolated cells [43,44] or tissues [45,46]. \textit{In vitro}, quantification within individual cells and resolution of individual isotopes of different metals is feasible [26,43,44]. However, interpretation of differences in distribution within tissues depends on unique structural features [45] or co-localizing markers [46] for cellular or sub-cellular areas of interest.

The gastric mucosa is divided into two spatially and functionally distinct regions [31,47,48]: the gastric glands, which secrete acid and pepsinogen; and the surface epithelium, which secretes mucus and bicarbonate and is thought to provide the protective barrier. The gastric glands, during exposure to secretagogues, demonstrate enhanced fluorescence imaging and fluorometric methods. Utilizing these methods, we were able to confirm enhanced uptake of Zn\(^{2+}\) by isolated glands, observing that it can be detected within minutes when extracellular concentrations are in the 10 nM range and very consistently when concentrations are 100 nM. Moreover, rates of uptake are accelerated during stimulation with a powerful secretory agonist, forskolin. The use of rare, non-radioactive isotopes such as \(\text{Zn}^{64}\) permits investigation of Zn\(^{2+}\) distribution into whole tissues and cell preparations, without the restrictions required when radioactive isotopes (e.g., \(\text{Zn}^{65}\)) would be used. The specificity of these methods for individual metal species not only permits quantification of rates of entry into cells and tissues, but also increases the confidence in designing studies utilizing potentially less specific reporters of Zn\(^{2+}\) movements, such as the fluorescent reporter fluo-3.

Connection between intracellular Ca\(^{2+}\) and uptake of Zn\(^{2+}\) across the basolateral membrane of the gastric parietal cell

Based on studies with \(\text{Zn}^{64}\), we were able to design experiments to study conditions of uptake under baseline conditions and during exposure to well-recognized secretory agonists, using utilizing relatively inexpensive and rapidly responsive fluorescence imaging and fluorometric methods. Utilizing these methods, we were able to confirm enhanced uptake of Zn\(^{2+}\) across the basolateral membrane during secretory stimulation. In addition, we found that manipulation of Ca\(^{2+}\) homeostasis leads to alterations in the ability of the parietal cell to take up and preserve Zn\(^{2+}\). Thus, intracellular accumulation of Zn\(^{2+}\) is nearly abolished when intracellular stores of Ca\(^{2+}\) are depleted by exposure to thapsigargin and Ca\(^{2+}\)-depleted Ringer’s. Following exposure to thapsigargin, uptake of Zn\(^{2+}\) was observed—albeit at a reduced level—if Ca\(^{2+}\) is present in the extracellular solution. This uptake was arrested when the chelator DTPA (10 \(\mu\)M) was added. These observations indicate that the increases in [Zn\(^{2+}\)], are indeed due to influx from extracellular
Zn\(^{2+}\) Demand and Ca\(^{2+}\) in Gastric Mucosa

Sources and not release from intracellular stores. They argue that, under baseline conditions, uptake of Zn\(^{2+}\) across the basolateral membrane depends on adequate stores of intracellular Ca\(^{2+}\). With stimulation by powerful agonists such as forskolin and carbachol, demand for extracellular Zn\(^{2+}\) increases and depends on influx of extracellular Ca\(^{2+}\). In addition, these studies illustrate physiologically relevant approaches for using both real-time imaging of individual glands, which is time-intensive, and 96-well platforms, which would be amenable to rapid through-put approaches for small molecule screens.

Additional considerations: technical
The fluorescent reporter utilized here was fluo-3, which has been utilized in its free acid form for monitoring extracellular secretion of Zn\(^{2+}\) from insulin-secreting beta cells [30]. When conjugated to an acetoxyethyl ester group, it has proven useful for monitoring intracellular concentrations of free Zn\(^{2+}\) in different cell types [29,49], including epithelial cells of the gastrointestinal tract [16,18]. Importantly, this reporter is unresponsive to Ca\(^{2+}\) under a number of experimentally relevant conditions [18,49,50]. However, it is possible for release or sequestration of other divalent cations to influence responses of the reporter [51]. It is for this reason that caution must be used in applying relationships that have been used to calculate free concentration of Zn\(^{2+}\) from the reported \(K_d\) and from maximum and minimum fluorescence responses, as was originally proposed for Ca\(^{2+}\) reporters [52]. Thus, we have refrained from providing direct estimates of [Zn\(^{2+}\)], based on fluo-3 measurements.

Additional considerations: connection between intracellular Ca\(^{2+}\) and movements of Zn\(^{2+}\) across the basolateral membrane of the gastric parietal cell
In the current set of studies, we find that Ca\(^{2+}\) facilitates optimal uptake of Zn\(^{2+}\) across the cell membrane, implying that it is either a counter-ion in exchange or it is acting as a regulatory second-messenger. Membrane proteins that facilitate Zn\(^{2+}\) transport constitute the SLC30A (ZnT) and SLC39A (ZIP) gene families [53,54]. To date, fourteen proteins that facilitate import of Zn\(^{2+}\) to the cytoplasm (either from extracellular sources or possibly intracellular compartments) have been identified in mammals [53,54]. Current information on the mechanisms of Zn\(^{2+}\) transport does not implicate Ca\(^{2+}\) in the former role and emerging information for ZIP, ZnT, and related transporters in yeast suggest that counter exchanging ions are likely to be protons [53]. Based on the well-recognized role of Ca\(^{2+}\) as a second messenger in responses to secretagogues [32,38,40,56,57,58,59], our findings offer the novel conclusion that Ca\(^{2+}\) is a second messenger that can match the basolateral demand for Zn\(^{2+}\) with the secretory response to physiologic stimulation. It seems likely that similar connections will be found in other secretory cells and tissues—neural, endocrine and exocrine.

Additional considerations: pathophysiological implications of Ca\(^{2+}\)-regulated Zn\(^{2+}\) uptake
Compared to levels of free Ca\(^{2+}\), the amount of free Zn\(^{2+}\) within the cytoplasm is even more tightly controlled. Our studies have suggested that, in different epithelial cells, the baseline concentration of [Zn\(^{2+}\)], is in the nanomolar range [16,18]. This consideration alone suggests tight regulation of the activities of transporters that modulate demand for Zn\(^{2+}\) and its disposal from the cytoplasm. Observations in other cell types such as neurons suggest that exposure to elevated levels of Zn\(^{2+}\) can be cytotoxic, within minutes [60,61]. Preliminary studies in epithelial cells of the gastrointestinal tract confirm that oxidant stress induces increases in [Zn\(^{2+}\)], [16] that can influence pathways of cell death and the balance between necrosis and apoptosis [19,62]. Dysregulation of Ca\(^{2+}\) homeostasis is also a consequence of such oxidant-related stress [16,39]. The findings of the current study suggest that oxidative disruption of intracellular Zn\(^{2+}\) could be amplified by dysregulation of intracellular Ca\(^{2+}\) homeostasis. Further studies will help to clarify this potential relationship, not only in gastric mucosa but also in other secretory cell types or tissues.

Author Contributions
Conceived and designed the experiments: JJL, AM, MAM, EB, CA, LEG, DIS. Performed the experiments: JJL, JELK, ALB, JAM, EAB, KAB, CA, NC, LEG, DIS. Analyzed the data: JJL, ALB, MAM, CA, NC, DIS. Contributed reagents/materials/analysis tools: JAM, AM, MAM, EAB, KAB, CA, NC. Wrote the paper: JJL, JELK, MAM, EB, CA, EB, LEG, DIS. Senior authorship and responsibility for this work: LEG, DIS.

References


